

Synthesis of and Complexation Studies with Polyaminopolycarboxylic Acid Crown Ethers

A Thesis

Submitted for the Degree of

Doctor of Philosophy

By

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*Dedicated to
my father B. Devasahayam,
my mother B. Danamma
and to my sister Chanti*

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DECLARATION

*I hereby declare that the matter embodied in this thesis is the **result of investigations** carried out by me in the School of Chemistry, University of Hyderabad, Hyderabad under the supervision of Professor M. Nagarajan.*

*In keeping with the general practice of reporting **scientific** observations, due acknowledgements have been made wherever the work described is based on the findings of other investigators.*

B. L. A. Prabhavathi Devi

B. L. A. Prabhavathi Devi.

CERTIFICATE

This is to certify that the work described in this thesis entitled
SYNTHESIS OF AND COMPLEXATION STUDIES WITH
POLYAMINOPOLYCARBOXYLIC ACID CROWN ETHERS *has*
been carried out by Ms. B. L. A. Prabhavathi Devi, under my supervision
and the same has not been submitted elsewhere for any degree.



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ABBREVIATIONS

aq	aqueous
Bn	benzyl
Bu	butyl
Bz	benzoyl
CDTA	1,2-diaminocyclohexanetetraacetic acid
cyclam	1,4,7,10-tetraazacyclotetradecane
DBU	1,8-diazabicyclo[5.4.0]undec-7-ene
DBN	1,5-Diazabicyclo[4.3.0]non-5-ene
DCC	1,3-dicyclohexylcarbodiimide
DCTA	1,2-diaminocyclohexanetetraacetic acid
dien	diethylenetriamine
DOTA	1,4,7,10-tetraazacyclododecane-N,N',N'',N'''-tetraacetic acid
DTPA	diethylenetriaminepentaacetic acid
DTPA-BAM	N,N''-bis[(methylcarbamoyl)methyl]-N,N',N'''-tris(carboxymethyl)diethylenetriamine
DMAP	4-dimethylaminopyridine
DMF	dimethylformamide
EDTA	ethylenediaminetetraacetic acid
EGTA	ethylene glycol(2-aminoethyl ether)tetraacetic acid
Et	ethyl
g	gram
HEHA	1,4,7,13,16-hexaazacyclohexadecane-N,N',N'',N''',N''''-hexaacetic acid

LAH	lithium aluminium hydride
Me	methyl
min	minute(s)
ml	millilitre(s)
mmol	millimole(s)
Ms	methanesulfonyl (mesyl)
NOTA	1,4,7-triazacyclononane-N,N',N''-triacetic acid
PCC	pyridiniumchlorochromate
PDC	pyridiniumdichromate
PEPA	1,4,7,10,13-pentaazacyclopentadecane-N,N',N'',N''',N''''-pentaacetic acid
Ph	phenyl
Pr	propyl
Py	pyridine
R ₁	longitudinal relaxivity
TAAHA	tris-(2-aminoethyl)aminohexaacetic acid
rt	room temperature
TEA	triethylamine
TETA	1,4,8,11-tetraazacyclotetradecane-N,N',N'',N'''-tetraacetic acid
TFA	trifluoroacetic acid
THF	tetrahydrofuran
TMS	trimethylsilyl
Ts	<i>p</i> -toluenesulfonyl (tosyl)

ABSTRACT

This thesis deals with the synthesis of and complexation studies with polyaminopolycarboxylic acid crown ethers, the crown diamine 71, the crown tetraamine 78, the crown diaminetetraacetic acid 79 and the crown tetraaminehexaacetic acid 82. The thesis consists of three sections, namely, introduction, results and discussion and experimental. The results and discussion is divided into two parts. Part-1 deals with the synthesis of the ligands 71, 78, 79 and 82. Part-2 deals with the determination of the dissociation constants of 71, 79 and 82 and the stability constants of these ligands with different metal ions.

The introductory chapter is divided into two sections. The first section presents a brief survey of some of the salient features of crown ether chemistry. In the second section the synthesis of polyaminopolycarboxylic acid chelating agents of medicinal interest, both diagnostic and therapeutic, are covered. A brief survey of their utility in medicine is also included.

Multidentate polyamino crown ethers 71 and 78 were synthesized in high yields starting from easily available pentaerythritol 2. Utilising the compounds 71 and 78, we prepared the target crown diaminetetraacetic acid 79 and the crown tetraaminehexaacetic acid 82. The general methods of N-alkylation of 71 and 78 with bromoacetic acid as well as with ethyl bromoacetate followed by hydrolysis of the respective ethyl esters 80 and 83 to obtain 79 and 82 in pure form were unsuccessful. Finally, the

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polyaminopolycarboxylic acid crown ethers 79 and 82 were obtained in pure form, by alkylating 71 and 78 with benzyl bromoacetate followed by hydrogenolysis of the respective benzyl esters 81 and 84. Attempts at the synthesis of multiloop spiro crown ethers 85, 87 and 89 were however unsuccessful.

In order to examine the chelating abilities of the ligands 71 and 79, we prepared some solid metal complexes such as the crown diamine-copper (II) complex 92, the crown salicylaldimine nickel (II) complex 94 and the crown diamine-tetraacetic acid-gadolinium complex 95

Using the potentiometric titration method, the dissociation constants of 71, 79 and 82 were evaluated employing the computer program SCOGS 2. The same technique was adapted to determine the stability constants of the complexes formed from 71, 79 and 82 with different metal ions.

From our results, it is evident that the first acid dissociation constant ($pK_1 = 2.86$) of the crown diamine 71 is very low when compared with that of 1,3-propanediamine ($pK_1 = 8.36$). This indicates that the diprotonated species of crown diamine 71 is more acidic than that of 1,3-propanediamine. The large difference in pK_1 's may be due to the effect of the crown ether moiety that is present in the crown diamine 71. The monoprotonated species of the crown diamine formed from the diprotonated species by loss of one proton, is stabilised by intramolecular complexation of the crown with the primary ammonium ion, thus shifting the equilibrium towards the

monoprotonated species and resulting in a low value for pK_1 . This type of large differences in acid dissociation constants are not observed in the case of crown diaminetetraacetic acid **79** as well as in crown tetraminehexaacetic acid **82**, when compared to trimethylenediamine tetraacetic acid (H_4L) and triethylenetetraamine hexaacetic acid (H_6L), because in these cases intramolecular complexation with the crown is not a factor

It was also found that the crown diamine **71** forms 1 : 1 and 2 : 1 complexes with nickel and copper, whereas with cobalt and lanthanides it forms 3 : 1 complexes. The stability constants ($\log K_1$ and $\log K_2$) of the crown diamine **71** with Cu^{2+} are 4.80 and 9.95, respectively and $\log K_N$ is 14.75. Among the lanthanides, crown diamine **71** was found to complex best with Gd^{3+} ion ($\log K = 15.40$). We also studied the synergistic effect of sodium ions on the complexation ability of the crown diamine **71** with transition and lanthanide ions. It was found that in the presence of sodium ions, the stability constants of crown diamine **71** with transition and lanthanide metal ions were reduced. For example, the stability constant of Cu^{2+} with crown diamine **71** in the absence of sodium ion was found to be 14.75, whereas in the presence of sodium ion it was 10.05. This effect is attributed to the participation of the amine group in the stabilization of sodium-crown complexation. In the case of the tetraacid **79**, this kind of effect was not significant, where the stability constants with Cu^{2+} in the absence of sodium ion was found to be 12.81, whereas in the presence of sodium ion it was 12.67. This is perhaps due to the greater role played by the carboxylate groups in metal ion complexation in the crown tetraacid **79** than

the amino groups in the crown diamine 71. It was also found that the crown tetraaminehexaacetic acid 82 is a better ligand than the crown diaminetetraacetic acid 79 and the crown diamine 71 with transition and lanthanide metal ions. The stability constant ($\log K$) of 82 with Gd^{3+} was 19.75 and with Cu^{2+} was 21.76, whereas for 79 and 71 the corresponding values with Gd^{3+} are 10.77 and 15.40 and with Cu^{2+} are 12.67 and 15.04.

In the experimental section, full details regarding the synthesis of the compounds as well as the measurements made with them are presented. A bibliography of all the relevant references concludes the thesis.

We have accomplished a novel and facile synthesis of crowns with pendant amino groups (71 and 78) and polyaminopolycarboxylic acid arms (79 and 82). The results obtained indicate that these ligands are good metal chelating agents with transition and lanthanide metal ions. These complexes have potential use as MRI agents and as NMR shift reagents for NMR active alkali metal ions.

Intrdoduction

INTRODUCTION

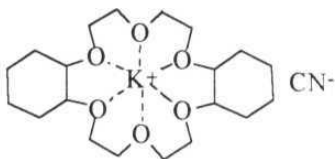
The discovery of dibenzo-18-crown-6 by Charles J Pedersen¹ in 1967 is one of the landmarks in the history of chemistry and also marks the beginning of synthetic host-guest chemistry. In 1987, the Noble prize in chemistry was shared by Pedersen, Cram and Lehn for their contributions to the field of supramolecular chemistry and host-guest chemistry. "Supramolecular chemistry" has been defined by Lehn² as "chemistry beyond the covalent bond or the chemistry of associates with well defined structure". The gross structure of the associates is governed by relatively weak forces such as hydrogen bonds, ion-dipole and dipole-dipole interactions and van der Waals interactions. Although supramolecular chemistry has rapidly expanded into a burgeoning research area encompassing a multitude of disciplines,³ in this introduction the attention is specifically on crown ethers.

This thesis, as the title itself indicates, deals with the synthesis of polyaminopolycarboxylic acid crown ethers and a study of their complexing abilities, especially towards rare earth metal ions. The potential utility of such complexes as contrast agents for magnetic resonance imaging (MRI) provided the impetus for undertaking the present work. In keeping with this objective, the introductory chapter is divided into two sections. The first section presents a brief survey of some of the salient features of crown ether chemistry. Due to the exhaustive nature of the subject, the coverage, as might be expected, is very selective and is restricted to crown ethers with multiple cation binding sites and their role in metal ion transport. Synthesis

of polyaminopolycarboxylic acids of medicinal interest, both diagnostic and therapeutic, are covered in the second section along with their applications. These include the use of metal complexes of polyaminopolycarboxylic acids as contrast reagents for MRI, in radioimmunotherapy, in tumour targeting, as models for calcium binding proteins and as antiviral agents against the viruses HIV-1 and 2

Most of the initial work on crown ethers was concentrated on the design of selective receptors for either alkali and alkaline earth cations⁴ and very little is known even today about receptors (host molecules) which can simultaneously and selectively complex with several different types of alkali, alkaline earth and heavy metal ions. Among the many applications of selective complexation of cations by crown ethers are chemical sensors,⁵ selective removal of poisonous or radioactive metal cations from waste streams,⁶ membrane transport,⁷ immobilisation of radioisotopes⁸ and as phase-transfer catalysts⁹

Crown ethers have found much use in organic synthesis as phase-transfer catalysts for nucleophilic substitutions.¹⁰ In a nucleophilic substitution reaction, the organic substrate is usually insoluble in water and other polar solvents, while the nucleophile is often an anion, which is soluble in water but not in the substrate or other organic solvents. When these kinds of reactions are carried out in the presence of crown ethers, the rate of the reaction is accelerated. Thus, for example, a salt like KCN is converted by

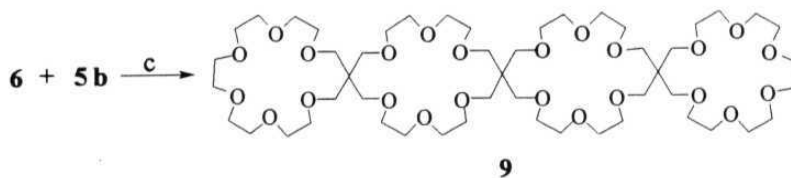
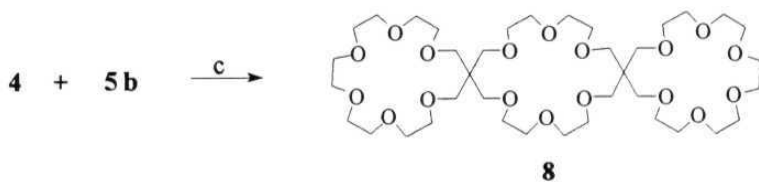
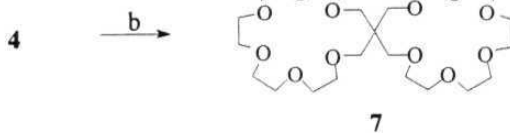
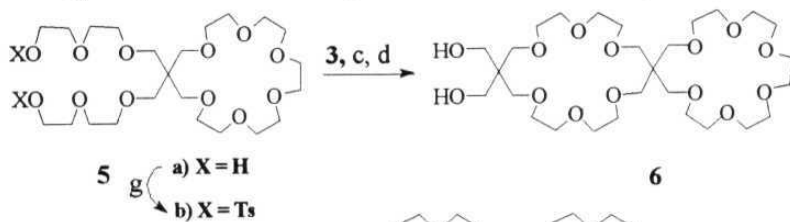
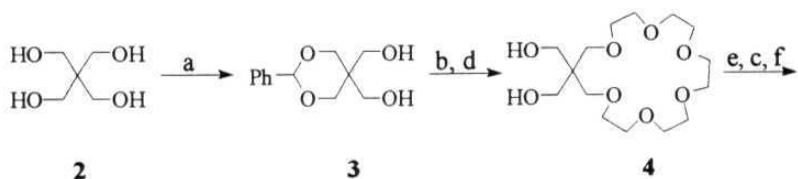


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Figure 1

dicyclohexano-18-crown-6 into a new salt **1** (Fig 1) whose anion is the same, but whose cation is now a much larger species with the positive charge spread over a larger volume, and hence much less concentrated. This larger cation is much less soluble in water than K^+ and more soluble in organic solvents, thus bringing the anion, by an ion-pairing mechanism, to the organic media. Also, the anion is not solvated by the organic solvent unlike in water and is therefore a "free" species which is more reactive. Any reaction that needs an insoluble anion to be solubilized in organic solvents can be accelerated by an appropriate phase-transfer catalyst.

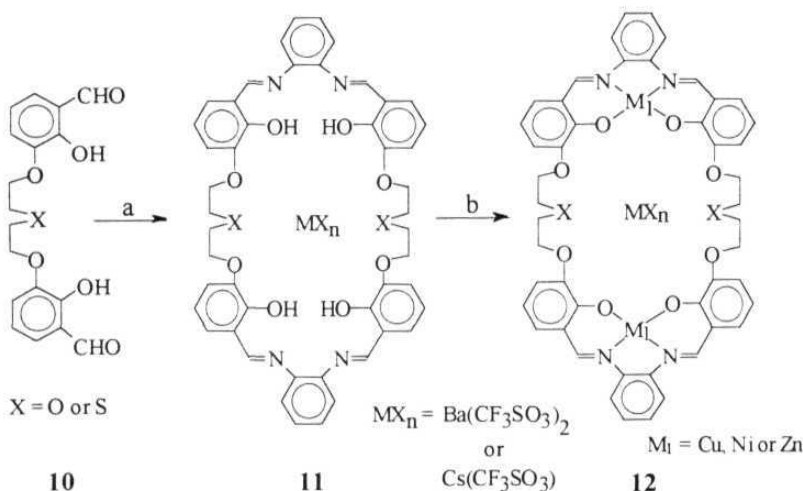
Most studies on complexation phenomena with crown ethers involve a single host and a single guest but molecular receptors with multiple interacting binding sites are of current interest. In 1982, Weber¹¹ reported the synthesis and selective complex formation of spiro-linked multiloop crown compounds **7**, **8** and **9**. These compounds were prepared by stepwise cyclisation, making use of blocking/deblocking and high-dilution techniques,



Scheme 1. *Reagents and conditions:* a) PhCHO, HCl, b) pentaethylene glycol ditosylate, NaH, THF, c) NaH, THF, d) H₂, Pd/C, EtOH, 3 atm, e) ClCH₂CH₂OCH₂CH₂OTHP, f) HCl, EtOH, g) *p*-TsCl, Py

starting from pentaerythritol (2) as the basic building block as shown in Scheme 1. It was found that the symmetrically looped molecules can incorporate two or three identical metal cations and asymmetrical loop molecules allow the simultaneous uptake of different metal ions of the alkali and alkaline earth families.

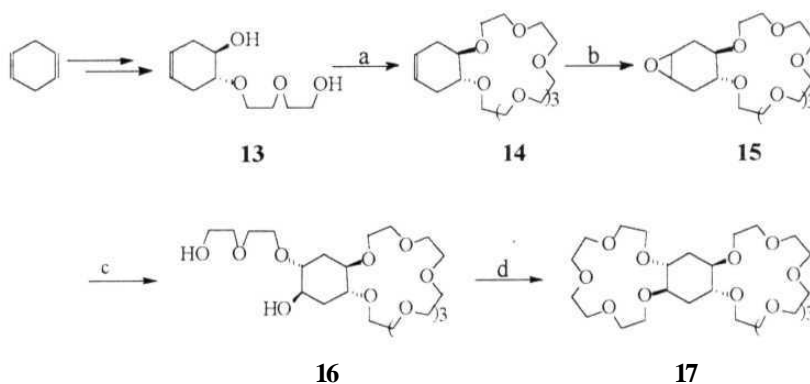
The propensity for complex formation in transition metal ions has prompted the synthesis of ligands with suitable donor atoms in the macrocycle. In 1991, Reinhoudt¹² synthesised macrocyclic hetero-trinucleating ligands that have one compartment for complexation of alkali or alkaline earth cations and two compartments for transition metal cations. The hetero-trinucleating ligands were prepared by (2+2) macrocyclisation of



Scheme 2. *Reagents and conditions:* a) *o*-phenylenediamine, $1/2$ MX_n , MeOH, heat, b) $M_I(OAc)_2$, MeOH

a salicylaldehyde derivative **10** and *o*-phenylenediamine in the presence of $Ba(CF_3SO_3)_2$ or $Cs(CF_3SO_3)$ as the template salt in methanol as shown in Scheme 2. The complexes **11** were converted into the hetero-trinuclear complexes **12** by reacting **11** with 2 equiv. of copper, nickel or zinc acetate in methanol. Dinuclear copper complexes without a bridging ligand are interesting as models for copper proteins.¹³

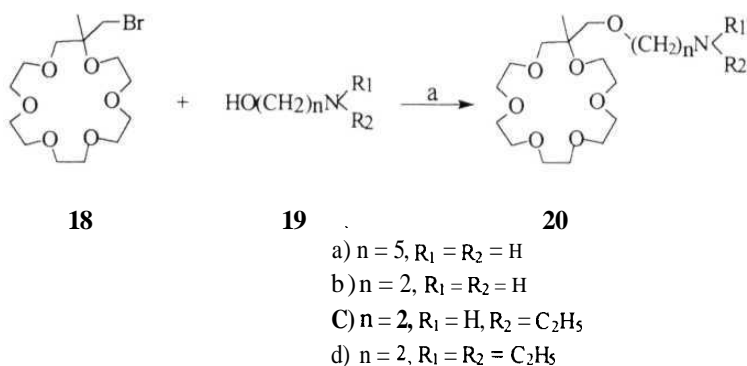
Bis-macrocyclic ethers with positive and negative allosteric cooperativity were developed by Costero¹⁴ A system in which binding at one site increases binding elsewhere displays positive cooperativity and the opposite result, negative cooperativity The bis-macrocyclic ether **17** was synthesized by converting 1,4-cyclohexadiene to its monoepoxide by treatment with hydrogen peroxide and ethyl chloroformate The monoepoxide was opened with diethylene glycol to the trans hydroxy ether **13**, from which the first ethereal cavity was constructed by condensation with the appropriate glycol ditosylate as depicted in Scheme 3 Repetition of the sequence leads to trans diaxial opening of the epoxide **15** to the second diol **16**, necessary for the construction of second cavity as exemplified by **17**



Scheme 3. *Reagents and conditions:* a) $\text{TsOCH}_2(\text{CH}_2\text{OCH}_2)_3\text{CH}_2\text{OTs}$, NaH, THF; b) H_2O_2 , ethyl chloroformate, c) $\text{HOCH}_2\text{CH}_2\text{OCH}_2\text{CH}_2\text{OH}$, d) $\text{TsOCH}_2\text{CH}_2\text{OCH}_2\text{CH}_2\text{OTs}$, NaH, THF.

It was shown that the bis-macrocyclic ether **17** was able to transport double the amount of Na^+ and K^+ cations when compared to the corresponding monocyclic systems because of the negative allosteric cooperativity

In ion transport systems, macrocyclic polyethers are excellent candidates for the recognition of the binding sites of carriers towards specific cations because of their selective complexation properties. Okahara¹⁵ and co-workers reported the synthesis, complexation and application in ion transport of a new series of 18-crown-6 ethers containing an amino group. Thus, the ionophores **20a-d** were prepared by the reaction of 2-(bromomethyl)-2-methyl-18-crown-6 (**18**) with the sodium alkoxide of the appropriate amino alcohols **19a-d** for 4 h at 100-120° (Scheme 4). These ionophores display better transport of K^+ than Na^+ . The foregoing brief



Scheme 4. Reagents and conditions: a) Na, 100-120°, 4 h

account underlines the importance of crown ethers in multi-cation complexation and metal ion transport

In recent times, rapid advances have occurred in medical diagnostics, amongst which, perhaps, MRI occupies the pride of place. MRI permits the study of different types of tissues and enables the identification of diseased tissues. For this purpose, nuclear magnetic resonance (NMR) imaging contrast agents are used in cancer and tumour treatment to detect the infected tissue. Complexes of paramagnetic transition metal and lanthanide ions, which can decrease the relaxation times of adjacent nuclei via dipolar interactions, have received most attention as potential contrast agents in magnetic resonance imaging (MRI). Paramagnetic contrast agents are unique amongst diagnostic agents as in the tissues these agents are not visualized directly on the NMR image, but are detected indirectly by virtue of changes in proton relaxation behaviour in their vicinity. The development of these agents offers an intriguing challenge for the investigators in the fields of chemical, physical and biological sciences. These include the design and synthesis of stable, non-toxic and tissue specific metal complexes and the quantitative understanding of their effect on the nuclear relaxation behaviour in solution and in tissue.

The need for NMR contrast agents and the interesting research problem associated with their development has produced an active research area. Bloch¹⁶ first described the use of a paramagnetic salt, ferric nitrate, to enhance the relaxation rates of water protons. The first human NMR imaging

study involving a paramagnetic agent was performed by Young¹⁷ The diagnostic potential of paramagnetic agents was first demonstrated in patients by Carr¹⁸ Gadolinium (III) diethylenetriaminepentaacetate $[[\text{Gd}(\text{DTPA})(\text{H}_2\text{O})]^{2-}]$ was administered intravenously to patients with cerebral tumours, providing enhancement of the lesion in the region of cerebral capillary breakdown. This is the only MRI agent currently in clinical use.

One of our objectives in the course of this research was to synthesize suitable ligands for use as complexing agents of metal ions, especially Gd^{3+} , so as to investigate their utility as MRI agents. Therefore, the requirements that a metal complex should possess for use as an MRI agent are briefly discussed

General requirements for metal complexes to function as NMR contrast agents:

NMR imaging contrast agents must be biocompatible Pharmaceuticals in addition to being nuclear relaxation probes. Besides the standard pharmaceutical features such as water solubility and shelf stability, the requirements relevant for metal complex based agents can be classified into three general categories 1) Relaxivity 2) Specificity in vivo distribution and 3) Non toxicity, in vivo stability and excreatability. Each of these is now discussed in greater detail.

1). Relaxivity: The efficiency with which the complex enhances the proton relaxation rates of water, referred to as relaxivity, must be sufficient to significantly increase the relaxation rates of the target tissue. This directly contributes to the sensitivity of the observation. In the specific case of Gd^{3+} , the hydrated ion has a longitudinal relaxivity of $9.1 \text{ mM}^{-1} \text{ s}^{-1}$ at 20 MHz and 35° , whereas the corresponding values for $\text{Gd}(\text{EDTA})^-$ and $\text{Gd}(\text{DTPA})^{2-}$ are 6.6 and $4.1 \text{ mM}^{-1} \text{ s}^{-1}$, respectively, under the same conditions¹⁹

2). Specific in vivo distribution: Ideally, to be of diagnostic value, the complex should localise for a period of time in the target tissue or tissue compartment in preference to nontarget regions, and thus enhance relaxation rates. This is not only a function of the concentration of the agent in the desired tissue, but more importantly, also of the difference in relaxivities of the desired tissue over the others. The choice of suitable MRI agent in this case is largely governed by the type of tissue to be studied.

3). Non-toxicity, in vivo stability and excretability: At the dosage required for MRI studies, lanthanide metal ions are known to be toxic. The toxicity is reduced by complexation with ligands which form stable and kinetically inert complexes with the metal ion. This forms the basis for detoxification of metal ion poisoning by chelation therapy. While the high thermodynamic stability reduces the degree of complex dissociation, the kinetic inertness precludes loss of metal ion due to substitution processes. Finally, the MRI agent should be excreted rapidly from the system.

In medical sciences, chelating agents are used in both diagnostic and therapeutic applications.²¹ Among the chelating compounds used either for chemical or medicinal purposes, polyaminopolycarboxylic acids stand out as archetypical metal binding agents. Some of the linear polyaminopolycarboxylic acid chelating agents (Fig. 2) are ethylenediaminetetraacetic acid (EDTA, **21**), 1,2-diaminocyclohexanetetraacetic acid (DCTA, **22**), tris-(2-aminoethyl) aminehexaacetic acid (TAAHA, **23**) and diethylenetriaminepentaacetic acid (DTPA, **24**).

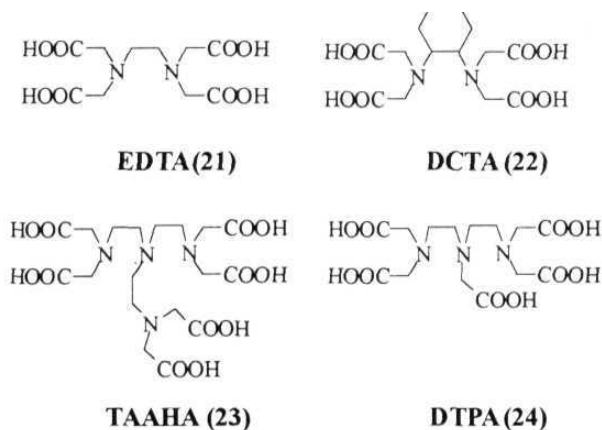


Figure 2

EDTA forms stable complexes with a number of physiologically important cations such as Ca^{2+} , Mn^{2+} , Cu^{2+} , Zn^{2+} , Fe^{2+} and Fe^{3+} and is used to protect against toxic elements in cationic forms. DTPA is superior to EDTA for polyvalent cations such as lanthanides and actinides because of its octadenticity.

Amongst the metal ions, Gd^{3+} ion appears to be the most useful paramagnetic²⁴ species because of its high magnetic moment and efficient relaxation. However, this ion is too toxic to be used in vivo and it must be reacted with a chelating agent before being injected into the blood stream in order to facilitate its rapid excretion through the kidneys. Safe and effective contrast agents containing gadolinium should not dissociate in the body, and thus should be highly stable and kinetically inert towards metal ion release. High stability is achieved with the Gd^{3+} complex of DTPA, and $\text{Gd}(\text{DTPA})^{2-}$ is the most commonly used MRI contrast agent today.

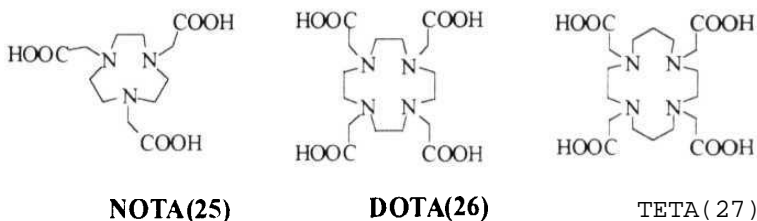


Figure 3

Chelating agents based on azacrown ethers bearing acetic acid moieties such as 1,4,7-triazacyclononane- N,N',N'' -triacetic acid (NOTA, 25), 1,4,7,10-tetraazacyclododecane- N,N',N'',N''' -tetraacetic acid (DOTA, 26) and 1,4,8,11-tetraazacyclotetradecane- N,N',N'',N''' -tetraacetic acid (TETA, 27, Fig 3) have attracted considerable interest in the design of NMR contrast agents because of their multidentate ligating sites.²⁵ Stetter and Frank²⁶ reported that DOTA 26 forms the most stable Ca^{2+} complex known, with a log K value of 16.5.²⁷ Kinetic investigation of the lanthanide DOTA chelates by Desreux²⁸ has shown that $\text{Gd}(\text{DOTA})^-$ dissociates more slowly than $\text{Gd}(\text{DTPA})^{2-}$ even in acidic media $\text{Gd}(\text{DOTA})^-$ is kinetically remarkably inert and also the complexes of lanthanides with DOTA have formation constants that are several orders of magnitude higher than both the EDTA and TETA. The stability constant for the formation of $\text{Gd}(\text{DOTA})^-$, for example, was found to be 10^{27} .

However, one of the drawbacks of these macrocyclic ligands is their slow complexation rate. The larger the ring size, the slower is the encapsulation rate.²⁸ In an attempt to design new ligand systems which could give a faster rate of complexation, Kodama^{29a} synthesized the macrocyclic pentaazapentacarboxylic acid ligand (PEPA, 28) and hexaazahexacarboxylic acid ligand (HEHA, 29, Fig.4) and studied their complexation with lanthanides (La^{3+} - Lu^{3+} , except Pr^{3+}). The stabilities of these complexes are almost parallel to those of DTPA and DOTA. In contrast to DOTA and TETA, PEPA and HEHA interact much faster with lanthanides. A comparison of the stability constants of the lanthanide

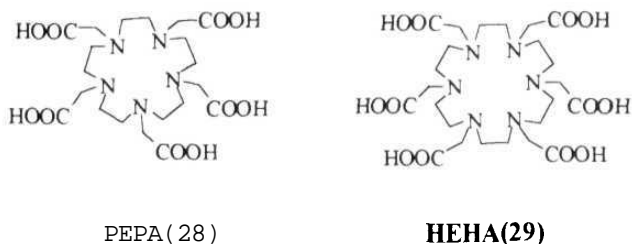


Figure 4

complexes of these macrocycles indicates that as the nitrogen atoms in the macrocycle increase, the 1 : 1 complex becomes more stable. The stability constants of these macrocycles are presented in Table 1.^{29b}

Table 1
Stability Constants (log K_{ML}) of Macrocyclic Complexes of
Lanthanides at 25° and $\mu=0.2$ M NaNO₃.

Metal ion	TETA	PEPA	HEHA	DTPA	DOTA
La ³⁺	12.74	13.57	19.11	19.50	–
Ce ³⁺	13.12	14.16	19.59	20.30	23.00
Nd ³⁺	13.76	14.85	20.36	21.60	–
Sm ³⁺	14.47	15.35	21.24	22.30	-
Eu ³⁺	14.66	15.59	22.68	22.40	28.20
Gd ³⁺	14.73	15.88	22.95	22.50	24.60

Martell³⁰ prepared a multidentate ligand containing hydroxypyridyl donor groups (Fig. 5), namely, N,N'-bispyridoxylethylenediamine-N,N'-diacetic acid (PLED, 30). This hexadentate ligand was found to have high affinity for trivalent metal ions [Ga(III) and Fe(III)], low ligand protonation constants and high water solubility. **Rocklage**³¹ synthesized the 5,5'-

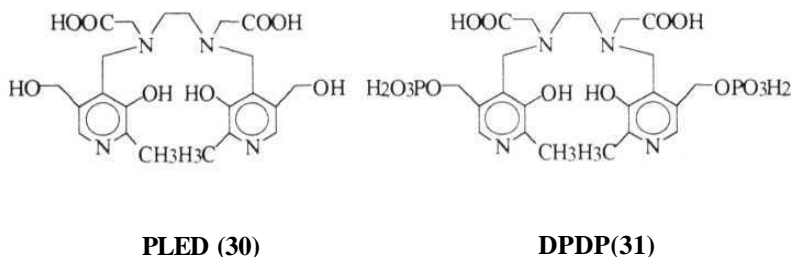
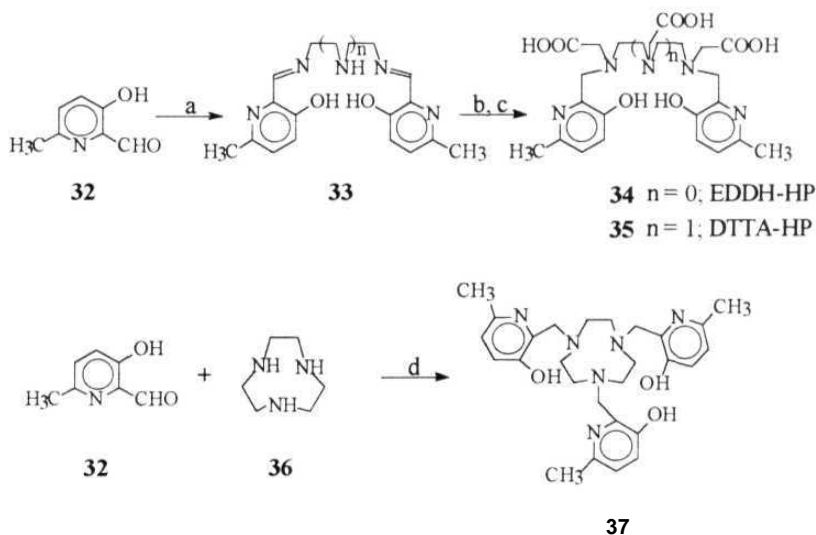


Figure 5

bisphosphate analogue of PLED, N,N'-dipyridoxylethylenediamine-N,N'-diacetic acid 5,5'-bisphosphate (DPDP, 31), and studied its divalent metal ion complexes for NMR imaging contrast enhancement. The disadvantage of these ligands is that they contain sterically demanding methyl groups and the pyridine nitrogens lower the pK_a 's of the ligands, making them more acidic. To overcome these problems, **Martell**³² prepared septadentate ligands 34 and 35 and one octadentate ligand 37 (Scheme 5) and studied their complexation properties with Ga(III), Fe(III) and In(III). The ligands 34 and

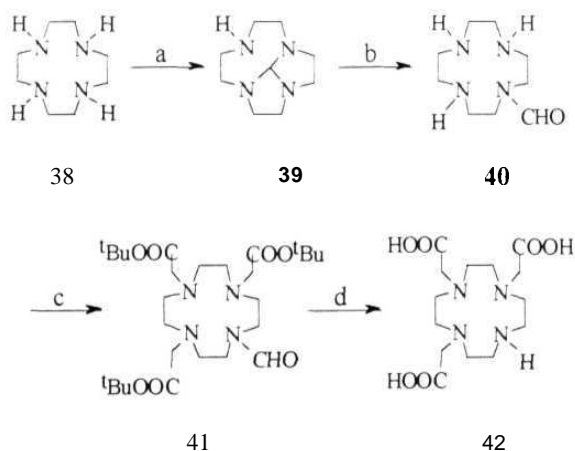
35 were prepared starting from a simple aldehyde, 3-hydroxy-6-methyl-2-pyridinecarboxaldehyde (32), by a modified Schiff's base formation, followed by reduction and alkylation with bromoacetate. The ligand 37 was prepared from triazacyclononane (36), formaldehyde and 3-hydroxy-6-methylpyridine (32) by the Mannich reaction as shown in Scheme 5. Preliminary investigations showed that Ga(III) and Fe(III) complexes of 37 are extremely stable when compared to those derived from 34 and 35.



Scheme 5. *Reagents and conditions:* a) diethylenetriamine, MeOH, b) NaBH_4 ; c) $\text{BrCH}_2\text{COONa}$, 6 N HCl ; d) CH_2O , EtOH

Gadolinium complexes of DTPA, DOTA and EDTA ligands are charged under physiological conditions, and the requirement of nonparamagnetic cationic counterions increases the osmolality of the solution. Minimum osmolality is predicted for a neutral complex, which behaves in aqueous solution as one species and this aspect has been addressed by several workers.

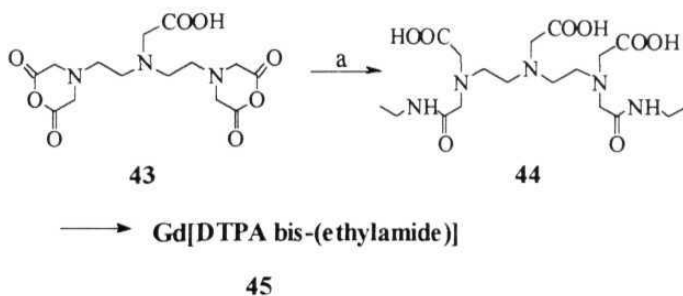
Tweedle³³ prepared a new synthetically useful ligand, 1,4,7-tris (carboxymethyl)-1,4,7,10-tetraazacyclododecane, D03A 42, by several methods. One amongst them was by carboxymethylation of 40 with chloroacetic acid or tert-butyl bromoacetate followed by removal of the



Scheme 6. Reagents and conditions: a) $(\text{CH}_3)_2\text{NCH}(\text{OCH}_3)_2$, cyclohexane, b) H_2O ; c) $\text{BrCH}_2\text{CO}_2^t\text{Bu}$, toluene, NaOH, H_2O ; d) H_2SO_4

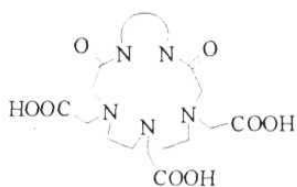
protecting groups as shown in Scheme 6. The ligand DO3A 42 is heptadentate and can be easily derivatized on the unique nitrogen of the macrocyclic ring, providing potentially octadentate ligands, which, upon reaction with gadolinium, form neutral Gd(III) complexes that are highly water soluble, inert to dissociation and substitution for Gd(III), and effective as proton relaxation agents.

In order to prepare a neutral gadolinium complex that retains high water solubility and relaxivity, DTPA-bis(ethylamide) 44 was prepared by Raymond³⁴ by treatment of DTPA-bis(anhydride) 43 with 70% ethylamine (Scheme 7). On reacting the ligand 44 with Gd_2O_3 in H_2O , the gadolinium complex 45 was obtained, whose crystal structure showed that the amide carbonyl oxygens play a prominent role in metal ion coordination.

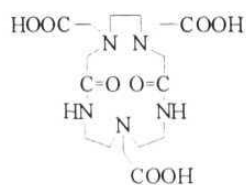


Scheme 7. *Reagents and conditions:* a) ethylamine, H_2O ; b) Gd_2O_3 , H_2O , heat

In order to understand the ligand selectivity in greater detail and to design newer generation of MRI contrast agents, several macrocyclic bis



DTPA . bis (amide)(46)



EDTA . DAM (47)

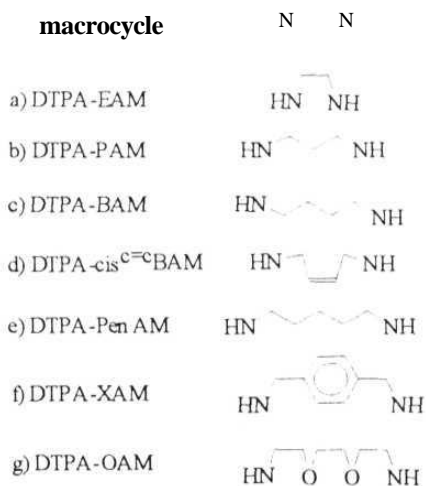
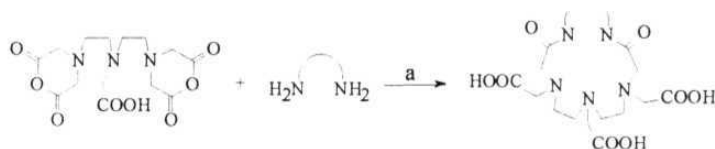


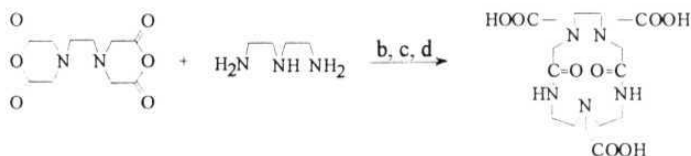
Figure 6

(amide) derivatives of DTPA and EDTA (Fig 6) were prepared by Carvalho³⁵ as illustrated in Scheme 8. Macrocyclic ligands have several advantages over acyclic ones as they are "preorganized" for metal complexation³⁶ and have the ability to structurally discriminate in binding various metal ions.³⁷

It was found that the stability of the Gd^{3+} complexes increases with increasing size of the DTPA-bis(amide) macrocycle series, because of the enhanced participation of the amide carbonyl oxygens in metal ion



DTPA . bis (amide) macrocycle (46)



EDTA . DAM (47)

Scheme 8. *Reagents and conditions:* a) DMSO, DMF, DBN; b) DBN, DMSO, c) $BrCH_2COO^tBu$, $i-Pr_2EtN$, MeOH; d) Trifluoroacetic acid

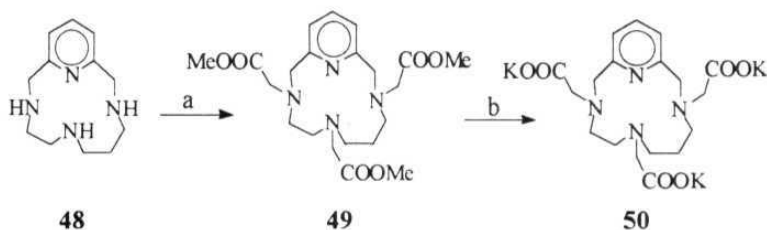
Table 2
Metal Chelate Stability Constants [log K, 25°, $\mu=0.10$ M (KCl)]j.

metal ion	DTPA-EAM	DTPA-PAM	DTPA-BAM ^a	DTPA-cis ^{C=C} BAM	DTPA-PenAM	DTPA-XAM	DTPA-0 AM	EDTA-DAM
Gd ³⁺	11.15	14.49	1685	15 56	15.94	12.34	1744	15.14
Zn ²⁺	12.08	11.74	1204	12 19	11 46	1065	12.19	9.03
(Gd ³⁺ /Zn ²⁺)	-0.94	2.75	4.81	3.38	448	1.69	5.25	6.12

^aRef 38

coordination (Table 2) The greater the thermodynamic stability, the lesser the degree of complex dissociation which can potentially lead to toxicity However, a metal complex in vivo may also undergo other types of reactions including ligand displacement and transmetallation Thus, for example, endogenously available metal ions such as Cu²⁺ and Zn²⁺ can potentially compete for the ligand of a contrast agent and promote the release of Gd³⁺ Gd(DTPA-BAM), the bis(methylamide) derivative of DTPA was the first nonionic MRI contrast agent developed for clinical application and its logarithmic selectivity constant for Gd³⁺ over Zn²⁺ is 4 81.³⁸ EDTA-DAM ligand was found to have highest selectivity for Gd³⁺ over Zn²⁺, presumably through a more favourable structural discrimination by this ligand, and the structural isomer DTPA-EAM showed less favourable selectivity for Gd³⁺ over Zn²⁺.

Recently, Aime reported the synthesis of a novel heptadentate chelating ligand **50** and its lanthanide complexes with improved relaxation efficiencies.³⁹ The ligand **50** was prepared by reacting **48** with methyl bromoacetate in the presence of silver (I) carbonate followed by hydrolysis



Scheme 9. *Reagents and conditions:* a) $\text{BrCH}_2\text{CO}_2\text{Me}$, Ag_2CO_3 , DMF, b) KOH, MeOH

with KOH as in Scheme 14. Ln^{3+} complexes of ligand **50** were prepared by mixing stoichiometric amounts of the ligand and the lanthanoid (III) chloride at neutral pH. Thermodynamic formation constant (K_f) for the Eu^{3+} complex was estimated through competitive NMR experiments between the ligand **50** and Eu^{3+} -1,2-diaminocyclohexane- N,N,N',N' -tetraacetic acid (**22**), and was found to be $3 \pm 2 \times 10^{18}$ and the relaxivity of Gd^{3+} -**50** complex was $6.3 \text{ m M}^{-1} \text{ s}^{-1}$ (20 MHz, 25°), about 35% higher than for Gd-DOTA and similar to that of Gd-DO3A

Complexes of polyazamacrocycles (bearing a C-substituted functional group for antibody attachment) exhibit remarkable kinetic inertness and have found applications in radioimmunotherapy. For example, Mears⁴⁰ showed that the copper complex of 2-(*p*-nitrobenzyl)-1,4,8,11-tetraazacyclotetradecane-*N,N',N'',N'''*-tetraacetic acid (nitrobenzyl-TETA 52, Fig. 7) is very stable in human serum under physiological conditions. The conjugate of this complex with a monoclonal antibody has been tested as well in tumour bearing mice. These types of macrocyclic polyamines are the key precursors to macrocyclic bifunctional chelating agents conjugated to monoclonal antibodies. Chelators that can hold radiometals such as ⁹⁰Y or

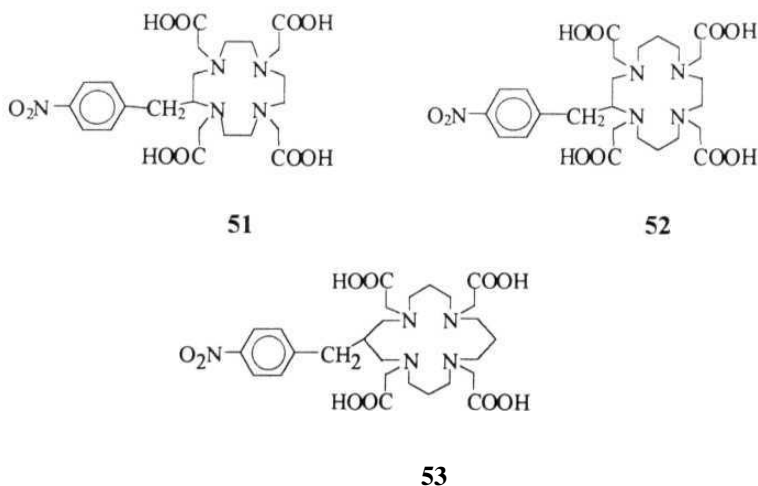
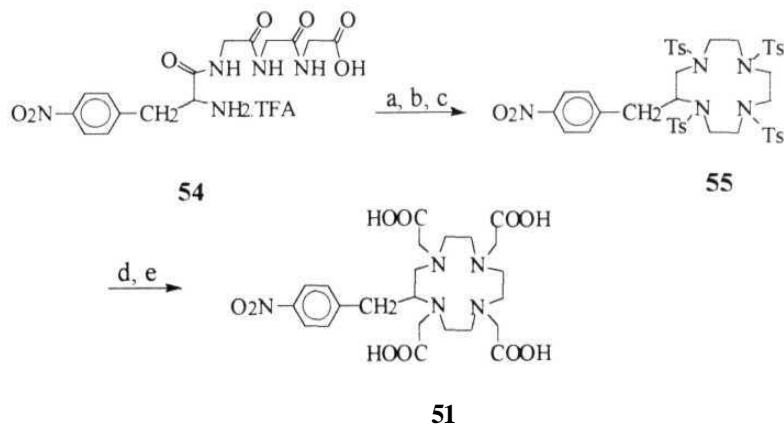


Figure 7

^{67}Cu with high stability under physiological conditions are essential to avoid excessive radiation damage to nontarget cells in radioimmunotherapy

In this connection, Mears⁴¹ prepared in good yields, polyazamacrocyclic bifunctional chelating agents such as the 12-membered 2-(*p*-nitrobenzyl)-1,4,7,10-tetraazacyclotetradecane-*N,N',N'',N'''*-tetraacetic acid (nitrobenzyl-DOTA, 51) via peptide synthesis and intramolecular tosylamide ring closure as shown in Scheme 10. The $^{88}\text{Y(III)}$ complex of this octadentate ligand (nitrobenzyl-DOTA, 51) forms in a few hours at



Scheme 10. *Reagents and conditions:* a) BH_3 , THF, reflux, 3h, b) *p*-TsCl, CH_3CN , Et_3N , rt, 8h; c) Cs_2CO_3 , DMF, 60° , 5h, d) 96% H_2SO_4 , $\text{C}_6\text{H}_5\text{OH}$, 100° , 48h, e) BrCH_2COOH , pH 10, 70° , 3h

room temperature in 0.1 M ammonium acetate at pH 5 and is quite stable, whereas under the same conditions nitrobenzyl-TETA 52 or nitrobenzyl-HETA 53 do not form $^{88}\text{Y(III)}$ complexes in significant yields.

Derivatives of EDTA and DTPA are also used as bifunctional chelating agents with applications in tumour targeting via monoclonal antibody conjugates. In 1974 Mears⁴² synthesized an EDTA derivative bearing a *p*-aminophenyl substituent **56a** (Fig. 8) that may be coupled to proteins under mild conditions. Benzyl (**56b**, **56c**), phenethyl (**56d**) and 2-carboxyethyl (**57**) analogues have subsequently been described. Recently, Gansow⁴³ reported the preparation of the DTPA analogue *p*-SCN-Bz-DTPA 60 according to Scheme 11

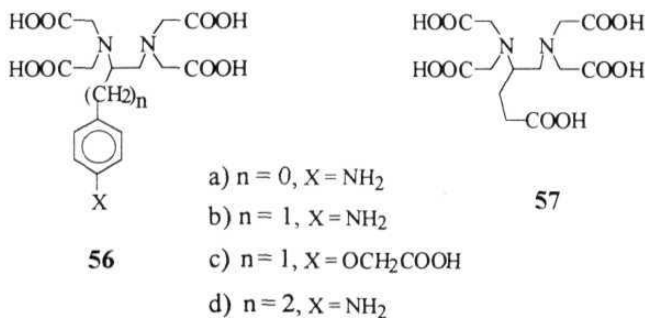
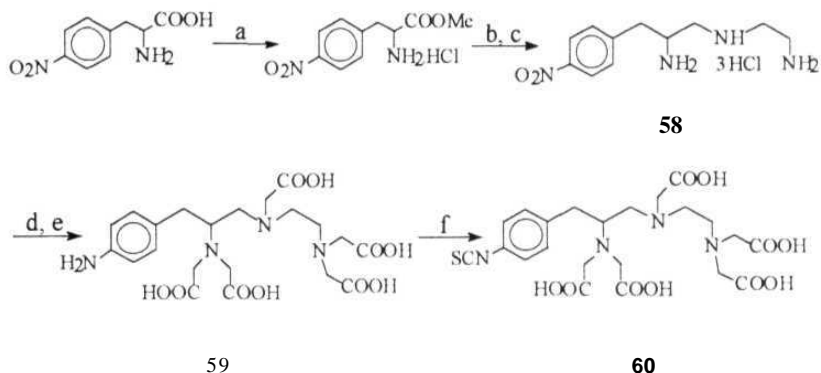
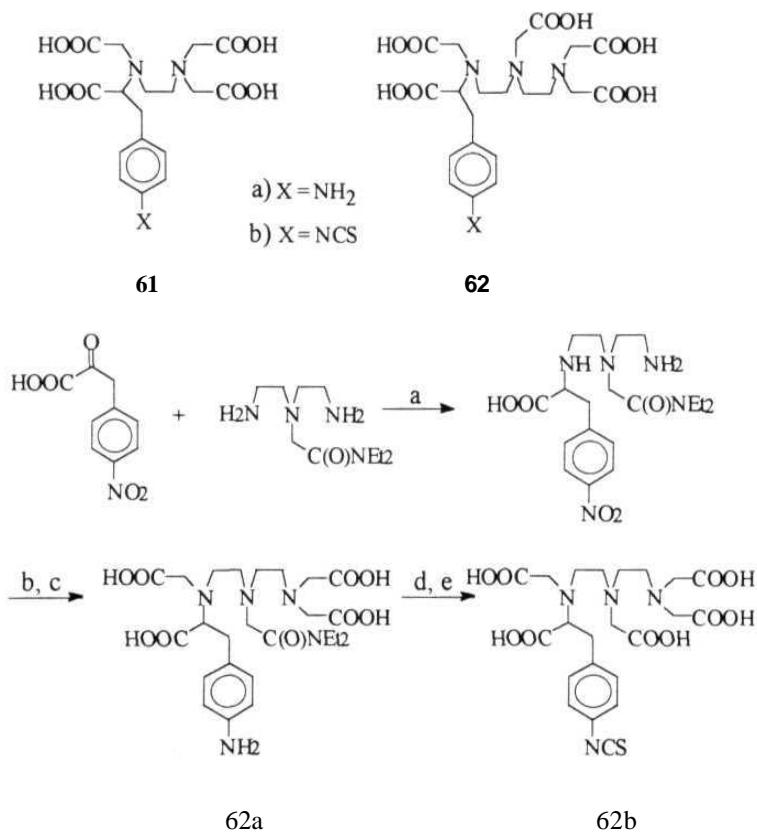


Figure 8



Scheme 11. *Reagents and conditions:* a) HCl, MeOH; b) ethylenediamine, TEA, c) BH_3 THF, HCl, d) BrCH_2COOH , KOH, e) H_2 , Pd/C, pH 10, f) C(S)Cl_2 , pH 8.5.

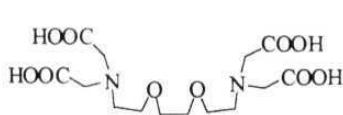
A common feature in all these structures is the attachment of the protein reactive **function** at a methylene carbon away from the polyamine **backbone**. In seeking a more flexible, generic route for introducing a protein reactive moiety into a wide variety of polyaminopolycarboxylate frameworks, Johnson⁴⁴ synthesized another class of bifunctional chelating agents of the types 61 and 62, in which the protein reactive site is incorporated at a position common to, and accessible in all such chelators, namely, on one of the carboxymethyl arms, as shown in Scheme 12. All these



Scheme 12. *Reagents and conditions:* a) NaOH, NaBH₃CN/HCl; b) BrCH₂COONa, c) H₂, Pd/C, H₂O/HCOOH, d) NaOH, e) C(S)Cl₂/HCl, CCl₄.

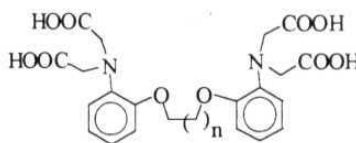
bifunctional chelators are used in preparing monoclonal antibody conjugates for tumour targeting

Polyaminopolycarboxylate type chelators exert interesting patterns in selectivity for binding a series of alkaline earth metal ions from Mg^{2+} to Ba^{2+} . Interest has been focussed on Ca^{2+} because of its importance in physiological activities such as muscle contraction, neurotransmitter release, hormonal response and blood clotting. Calcium binding proteins, such as troponin C⁴⁵ and calmodulin,⁴⁶ have high affinity for Ca^{2+} over Mg^{2+} and other alkaline earth metal ions because of their polycarboxylate binding sites. In 1980, Schauer and Anderson⁴⁷ synthesized 3,6-dioxaoctane-1,8-diamine- N,N,N',N' -tetraacetic acid (H_4 egta, 63, Fig.9) and showed it to be a useful model for the Ca^{2+} binding site of proteins



H_4 egta

63



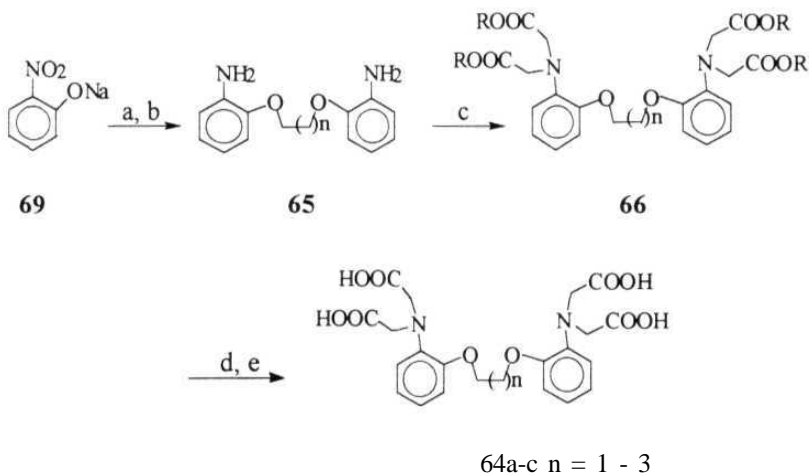
a) $n=1$, H_4 bapta

64 b) $n=2$, H_4 baptpa

c) $n=3$, H_4 bapbta

Figure 9

Tsien⁴⁸ synthesized 1,2-bis(*o*-aminophenoxy)ethane-*N,N,N',N'*-tetraacetic acid (H₄ bapta, 64a), an H₄ egta like chelator in which the 1,2-disubstituted benzene rings hold the amino nitrogens in the vicinity of the ethereal oxygens (Scheme 13). In addition, H₄ bapta has some useful properties. The existence of two phenyl groups in the molecule endows H₄ bapta with an appropriate absorption maximum in its UV spectrum. Thus,



Scheme 13. *Reagents and conditions:* a) $\text{Br}(\text{CH}_2)_n\text{Br}$, DMF, 100° ; b) Sn, con.HCl, EtOH, reflux; c) 1,8-bis(diethylamino)naphthalene, $\text{BrCH}_2\text{CO}_2\text{R}$ (R = Me or Et), CH_3CN , reflux, d) KOH, EtOH/ H_2O , 60° , e) Dowex 50wX8, H_2O

the chelated and free states of H_4 bapta can be followed quite conveniently by UV spectroscopy. In addition, H_4 bapta complexes faster with Ca^{2+} than does H_4 egta, with about the same selectivity for Ca^{2+} over Mg^{2+} as H_4 egta.

In order to study the structural interaction relationships on selective chelation towards metal ions, Ohno⁴⁹ prepared 1,3-bis(*o*-aminophenoxy)propane- N,N,N',N' -tetraacetic acid (H_4 bappta, **64b**) and 1,4-bis(*o*-aminophenoxy)butane- N,N,N',N' -tetraacetic acid (H_4 bapbta, **64c**) following the procedure reported by Tsien as depicted in Scheme 13 and showed that the Ca^{2+}/Mg^{2+} selectivity of bappta⁴⁻ resembles that of the Ca^{2+} -transporting protein, calmodulin.

A series of bis-tetraazamacrocycles such as the bicyclam JM2763 (65) and JM3100 (66, Fig 10), are a novel class of antiviral agents that exhibit potent inhibitory effects on HIV-1 and HIV-2 replication with high selectivity. These have been reported by De Clercq.⁵⁰

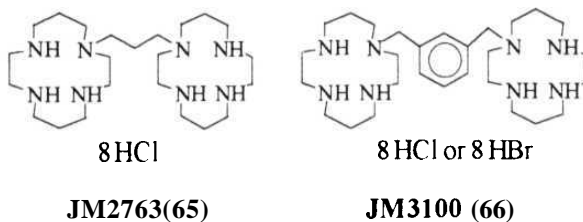


Figure 10

As mentioned above, polyaminopolycarboxylic acid chelating agents such as EDTA, DTPA, DOTA and TETA and their derivatives form stable complexes with a number of transition and lanthanide metal ions which are useful in MRI studies and cancer treatment, whereas the crown ether complexes with alkali and alkaline earth metal ions are important from the physiological viewpoint. Pendant macrocyclic compounds containing both crown ether and polyaminopolycarboxylic acid moieties are unknown. Such compounds may exhibit some interesting properties which may be useful in biological systems. Thus, we planned to prepare some polyaminopolycarboxylic acid crown ethers and study their complexation abilities with transition and lanthanide metal ions. The synthesis *of* these compounds, and the results *of* their complexation studies, are discussed in the next chapter.

RESULTS AND DISCUSSION

Results and discussion

As mentioned in the introductory chapter, the potential utility of chelating agents containing pendant polyaminopolycarboxylic acid arms in magnetic resonance imaging, cancer diagnosis and therapy prompted us to prepare some compounds containing both crown ether and polyaminopolycarboxylic acid units. Two prototype structural units A and B incorporating both crown ether and polyaminopolycarboxylic acid moieties shown in Fig. 11 were initially identified for further analysis. The presence of two α -amino ether units in structure A suggests that it would be susceptible to acid catalysed hydrolysis and hence attention was focussed on structure B. A straightforward retrosynthetic analysis of B led to the conclusion that pentaerythritol would be a convenient starting material.

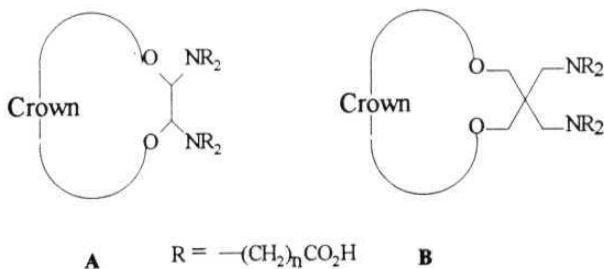
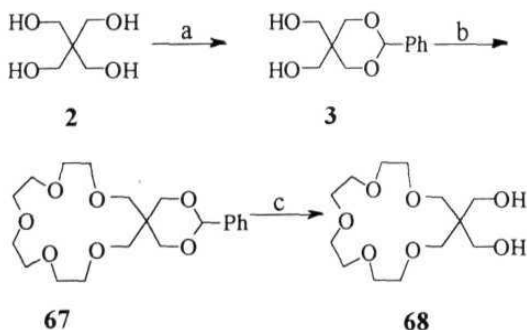


Figure 11

Weber has reported the synthesis of multi-loop spiro crowns 7, 8 and 9 containing the crown diol moiety 4. We realized the utility of 4 and related structures in the synthesis of our polyaminopolycarboxylic acid crown ethers since they provide a crown ether cavity as well as the hydroxyl functionality which can further be transformed into the polyaminopolycarboxylic acid unit.

Thus, the crown diol 68 was prepared starting from the easily available pentaerythritol [2, Scheme 14]. Treatment of pentaerythritol (2) with benzaldehyde in the presence of conc. HCl furnished monobenzalpentaaerythritol 3 which on treatment with tetraethylene glycol ditosylate in the presence of sodium hydride in THF at reflux temperature

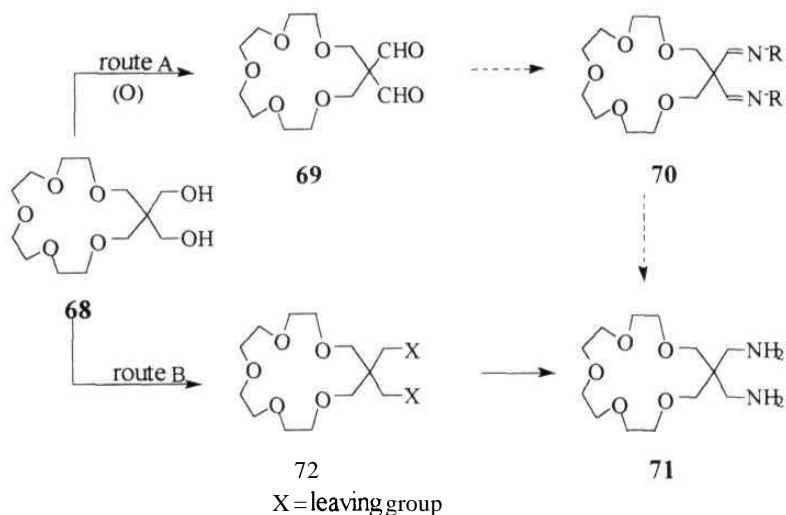


Scheme 14. *Reagents and conditions:* a) PhCHO, HCl; b) tetraethylene glycol ditosylate, NaH, THF, reflux, 12 h; c) 0.1 N H₂SO₄, heat, 12 h

afforded the 3-phenyl-2,4,8,11,14,17,20-heptaoxaspiro[5.15]heneicosane (67). This upon acid hydrolysis afforded the required crown diol 68. The physical and spectral properties of 68 were in agreement with those reported in the literature.

At this point, starting from the crown diol 68 several options were considered to prepare the crown diamine 71, which could serve as the starting material for the synthesis of the target polyaminopolycarboxylic acid crown ethers. In the first approach, synthesis of the crown dialdehyde 69 was envisaged to be followed by its condensation with a suitable amine to provide the Schiff's base 70 (Scheme 15). Subsequent removal of the R group would then lead to the crown diamine 71. This route was planned so as to avoid a nucleophilic substitution of the hydroxyl group of 68, a neopentyl type substrate, by a nitrogen nucleophile. An alternate method for the synthesis of 71 involves a nucleophilic substitution as the key step, although it suffers from the drawback mentioned earlier (route B).

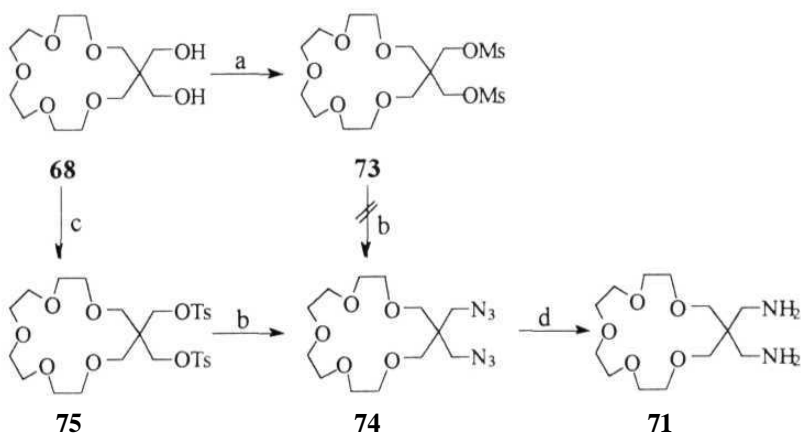
Oxidation of the crown diol 68 with several oxidising agents such as pyridinium chlorochromate (PCC), pyridinium dichromate (PDC), chromic acid in pyridine and N-chlorosuccinimide/dimethyl sulfide, all lead to complex mixtures. Examination of the IR spectra of these mixtures revealed the presence of the carbonyl group. However, no separations of the mixtures could be achieved.



Scheme 15.

As oxidation of **68** did not provide the required dialdehyde **69** in pure form, the second approach was adopted. Thus, as shown in Scheme 16, the crown diol **68** on treatment with methanesulfonyl chloride provided the crown dimesylate **73** which, without further purification, was reacted with sodium azide to afford a mixture of products from which the required crown diazide **74** was not separable in pure form by column chromatography. The IR spectrum of the mixture showed the characteristic azide absorption at 2100 cm^{-1} and mesylate bands at 1370 and 1190 cm^{-1} , indicating the presence of both product and starting material. This incomplete reaction is

probably due to the sluggish nucleophilic displacement of **the mesylate** group. In order to make the nucleophilic displacement more facile, we planned to use the crown ditosylate **75** instead of the dimesylate. An initial attempt at tosylation of **68** was by treating it with *p*-toluenesulfonyl chloride in pyridine. The reaction gave a mixture and the required product could not be obtained in a pure form. Under the conditions employed by Ouchi, we were able to tosylate the crown diol **68** in good yield (80%) and the desired product **75** was obtained as a white crystalline solid. The product was characterized by its spectral as well as by its analytical data. The IR spectrum



Scheme 16. *Reagents and conditions:* a) methylenesulfonyl chloride, DMAP, pyridine, b) NaN_3 , DMF, 120° , 72 h, c) *p*-toluenesulfonyl chloride, NaOH, THF/ H_2O , 5° , 1 h; d) LAH, THF, reflux, 12 h.

of the compound showed sulfonate bands at 1353 and 1187 cm^{-1} and no band corresponding to the -OH groups, substantiating the ditosylation of the crown diol 68. The ^1H and ^{13}C NMR spectra were also fully in consonance with the assigned structure.

Nucleophilic displacement of the crown ditosylate 75 with azide was performed by refluxing it with 6 eq. of sodium azide in DMF for 3 days. Under these conditions, the crown diazide 74 was obtained in almost quantitative yield. The IR spectrum of crown diazide 74 clearly showed the azide absorption at 2100 cm^{-1} .

Our initial attempts at reducing the diazide 74 to diamine 71 with 2.5 eq. of LAH at rt or at reflux were unsuccessful. Subsequently, other methods of reduction of the crown diazide 74 such as catalytic transfer hydrogenation using 10% Pd/C in cyclohexene⁵² or triphenylphosphine/hydrobromic acid/acetic acid⁵³ were found to be of no avail.

Surprisingly, when the reduction of 74 was carried out with 6 fold excess of LAH in THF under reflux, the required crown diamine 71 was obtained in quantitative yield. The diamine 71 was completely characterized by its spectral and analytical data. Its IR spectrum showed amine absorptions at 3315, 939 and 879 cm^{-1} and the absence of the azide band at 2100 cm^{-1} , indicating the complete reduction. The ^1H NMR spectrum contained a singlet at 2.89 ppm corresponding to the methylene protons adjacent to the amino group, as well as signals at 3.46 and 3.60-3.65 ppm that of the crown

protons. Finally the ^{13}C NMR spectrum displayed resonance at 72.24, 70.77, 64.59, 44.59 and 35.59 ppm, conforming the assigned structure.

The reactions of two other nitrogen nucleophiles, ethyl urethane and benzyl amine on the crown dithiolate **75** were both unsuccessful. Only the unreacted starting material could be recovered in each case.

At this stage, our attention was focussed on the synthesis of polyaminopolycarboxylic acid crown ethers starting from the crown diamine **71**. The crown diamine **71**, at the most, can only lead to a crown diaminetetraacetic acid with 6 binding sites. In order to obtain compounds with more ligating sites (lanthanides are known to form complexes with higher coordination numbers like 8-12),²⁴ synthesis of the crown tetraamine **78** was planned, which in turn can be transformed to a crown tetraaminehexaacetic acid having 10 binding sites.

Synthesis of the crown tetraamine **78** can be conceived of in two possible ways (Fig 12). The first approach consists of the nucleophilic displacement of the crown dihalide **72** with ethylenediamine. The second route involves incorporation of two more nitrogens into the crown diamine moiety. The former suffers from the drawback that it can yield more than one product leading to problems in purification. Therefore, the latter approach was adopted since the formation of a mixture of products can be avoided by selecting suitable reagents.

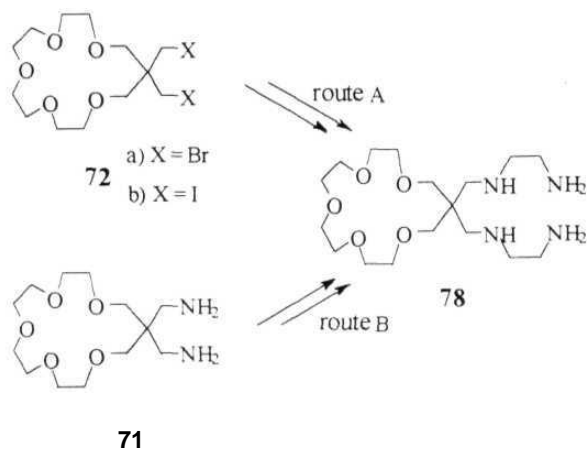
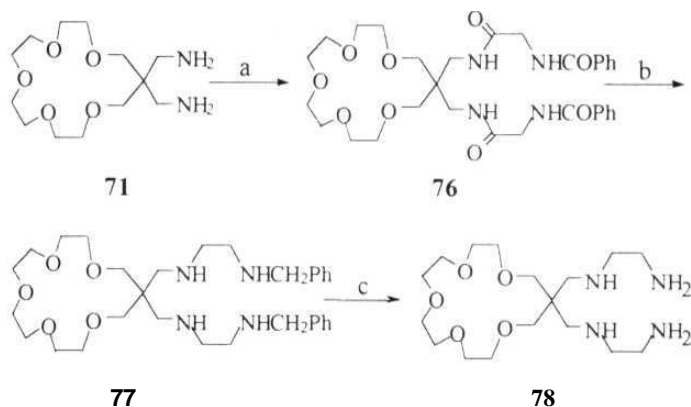


Figure 12

Thus, according to Scheme 17, the crown diamine 71 was condensed with hippuric acid in the presence of DCC to afford the diamide 76 in 75% yield. Compound 76 was fully characterised by elemental and spectral data. The IR spectrum of 76 contains the characteristic amide absorption bands at 1650 and 1550 cm^{-1} and the ^1H NMR spectrum showed a doublet at 4.12–4.15 ppm corresponding to methylene protons adjacent to the carbonyl group (COCH_2NH). The ^{13}C NMR spectrum indicates the presence of two types of carbonyl carbons at 170.19 ppm (NHCOCH_2) and 167.50 ppm (NHCO^iPh).



Scheme 17. *Reagents and Conditions:* a) hippuric acid, DCC, dichloromethane, 5°; b) LAH, THF, reflux, 12 h; c) H₂, 20% Pd(OH)₂/C, EtOH, 65 psi, 5 h

Diamide 76 on reduction with 6 eq of LAH in THF at reflux afforded the crown N,N'-dibenzyltetraamine 77 in quantitative yield as a brown coloured syrup. An excess of LAH was needed for the complete reduction of 76 to 77. The IR spectrum of 77 showed the absence of amide and carbonyl bands, indicating the complete reduction of 76. The FAB mass spectrum contains the (M + H)⁺ peak at m/z = 559. Inadequate purification of 77 due to its high polar nature precluded its elemental analysis. Hence its hydrochloride salt was prepared by passing dry HCl gas into a dichloromethane solution of 77. The salt, however, was highly sensitive to

moisture Finally, 77 was characterized as its (tetrakis)hexafluorophosphate salt, a white solid prepared by treating the hydrochloride salt of 77 with 4 eq of ammonium hexafluorophosphate

Removal of the N-benzyl groups of 77 was attempted next No cleavage occurred when 77 was treated with: 1) 4 4% formic acid in methanol and 10% Pd/C⁵⁴ (quantitative amount), 2) sodium in liq ammonia,⁵⁵ 3) ammonium formate and 10% Pd/C⁵⁶, 4) hydrogenation with 10% Pd/C in the presence of acetic acid⁵⁷ and 5) hydrogenation with 10% Pd/C in ethanolic HCl ⁵⁸ Finally, cleavage of the N-benzyl group was attained using 20% Pd(OH)₂/C (Pearlman **catalyst**).⁵⁹ Thus, when the N,N'-dibenzyltetraamine 77 was hydrogenated in ethanol using 2 eq of 20% Pd(OH)₂/C (by weight) at 65 psi for 4 h, the desired crown tetraamine 78 was obtained in good yield as a hygroscopic foamy material During the course of this reaction, it was observed that the amount of 20% Pd(OH)₂/C used was a decisive factor When catalytic or 1 eq of 20% Pd(OH)₂/C was used, cleavage of the N-benzyl group was not effected This is attributed to the complexation of the crown tetraamine 78 with palladium, thus poisoning the catalyst for further reaction

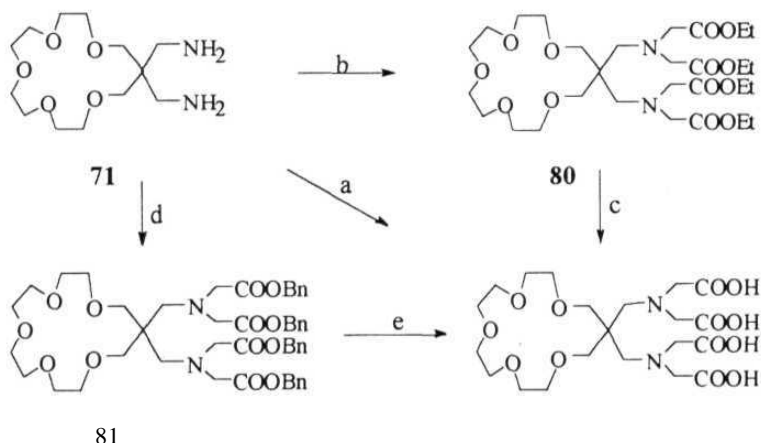
The hygroscopic nature of 78 precluded direct elemental analysis For the complete characterization of 78, preparation of its hexafluorophosphate salt was attempted, but without success However, the bis(tetraphenylborate) salt of 78 could be prepared by treating 78 in 0.1 M HCl solution with an aq solution of sodium tetraphenylborate This was

identified by its elemental and spectral data. The ^1H NMR spectrum showed no signals due to the benzyl groups, indicating complete cleavage.

Having the crown diamine 71 and the crown tetraamine 78 in hand we endeavoured to prepare the corresponding tetra- and hexaacetic acids as depicted in Schemes 18 and 19. Initially, direct N-alkylation using bromoacetic acid was attempted.⁴³ Thus, when the crown diamine 71 and the crown tetraamine 78 were subjected to N-alkylation with bromoacetic acid in the presence of potassium hydroxide at 5° , the crude crown diaminetetraacetic acid 79 and the crown tetraaminehexaacetic acid 82 were obtained. The IR spectra of 79 and 82 clearly indicate the acid hydroxyl and carbonyl absorptions at 3408 and 1734 cm^{-1} . Purification of 79 and 82 by either chromatography or ion exchange techniques was, however, unsuccessful.

An alternate method, also reported in the literature, involves the use of ethyl bromoacetate as the alkylating reagent, followed by hydrolysis to afford the free acid⁶⁰. Accordingly, the crown ethyl esters 80 and 83 were obtained by N-alkylation of the corresponding amines 71 and 78 with ethyl bromoacetate in the presence of potassium carbonate. These compounds 80 and 83 were characterized by their spectral and analytical data. The IR spectrum of 80 showed the ester carbonyl band at 1734 cm^{-1} and the ^1H NMR spectrum indicated the presence of ethyl ester signals as a quartet at 4.09-4.19 ppm and a triplet at 1.22-1.25 ppm, along with crown proton signals at 3.48-3.64 ppm as a multiplet. The ^{13}C NMR spectrum showed the

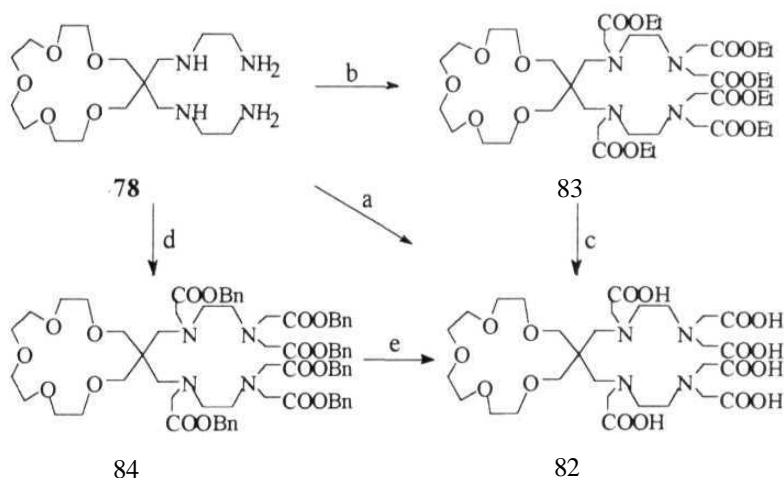
ester carbonyl carbon at 172 ppm and finally the FAB mass spectrum displayed the $(M + H)^+$ peak at $m/z = 637$. In the IR spectrum of 83, a strong ester carbonyl absorption at 1732 cm^{-1} was seen. The ^1H NMR spectrum showed signals at



Scheme 18. *Reagents and conditions:* a) BrCH_2COOH , KOH , H_2O ; b) $\text{BrCH}_2\text{COOEt}$, K_2CO_3 , DMF , 100° , 48 h, c) KOH , H_2O , reflux, 12 h, d) $\text{BrCH}_2\text{COOBn}$, K_2CO_3 , DMF , 120° , 72 h, e) H_2 , 20% $\text{Pd}(\text{OH})_2/\text{C}$, EtOH , 65 psi, 3 h.

4.13-4.20 and at 1.22-1.29 ppm corresponding to the ethyl ester protons and the crown protons at 3.53-3.63 ppm as a multiplet. The ^{13}C NMR spectrum

contains two carbonyl carbon peaks at 172 and 171 ppm and the FAB mass spectrum gave the $(M + H)^+$ peak at $m/z = 895$



Scheme 19. *Reagents and conditions*: a) BrCH_2COOH , KOH , H_2O ; b) $\text{BrCH}_2\text{COOEt}$, K_2CO_3 , DMF, 100° , 72 h; c) KOH , H_2O , reflux, 12 h; d) $\text{BrCH}_2\text{COOBn}$, K_2CO_3 , DMF, 120° , 72 h; e) H_2 , 20% $\text{Pd}(\text{OH})_2/\text{C}$, EtOH, 65 psi, 5 h.

Basic hydrolysis of 80 and 83 yielded the crude crown diaminetetraacetic acid 79 and the crown tetraaminehexaacetic acid 82, respectively. In this case also purification of 79 and 82 was unsuccessful either by chromatography or ion-exchange techniques. This difficulty

perhaps arises due to the use of strongly basic and acidic conditions in the course of hydrolysis and workup. In order to avoid acidic and basic conditions in the preparation and purification of 79 and 82, either in reaction or in the workup procedure, we adopted a new method for their synthesis. This method comprises the preparation of benzyl esters of 79 and 82, followed by hydrogenolysis. Accordingly, 71 and 78 were reacted with benzyl bromoacetate in the presence of potassium carbonate to obtain the corresponding benzyl esters 81 and 84 in 54% and 12-15% yields, respectively. Attempts to improve the yield of 84 by treating 78 with benzyl iodoacetate/potassium carbonate or cesium carbonate in DMF as well as in acetonitrile under reflux conditions for 3 days were of no avail. These benzyl esters 81 and 84 were identified by their elemental and spectral data. The IR spectra of 81 and 84 showed the carbonyl absorption at 1743 cm^{-1} and ^1H NMR spectrum clearly indicated the benzyl methylene protons at 5.10 ppm as a singlet. In the ^{13}C NMR spectrum of 81, the carbonyl signal was observed at 171.76 ppm whereas in 84 two carbonyl signals at 171.14 and 170.60 ppm were observed.

Finally, synthesis of the desired crown diaminetetraacetic acid 79 and the crown tetraaminehexaacetic acid 82 were accomplished by hydrogenolysis of 81 and 84, respectively, using 2 eq. of 20% $\text{Pd}(\text{OH})_2/\text{C}$ (by weight) in ethanol at 65 psi. These compounds 79 and 84 were fully characterised by their elemental and spectral data. The IR spectrum of 79 showed the acid OH and carbonyl absorption bands at 3422 and 1739 cm^{-1} , respectively. In the ^1H NMR spectrum, the benzyl ester signals of 81 were

absent and the ^{13}C NMR spectrum clearly displayed the acid carbonyl carbon at 170 ppm. The IR spectrum of 82 showed acid OH and carbonyl absorption bands at 3429 and 1736 cm^{-1} , respectively. The ^1H NMR spectrum showed the absence of benzyl ester signals of 84, indicating complete benzyl group cleavage.

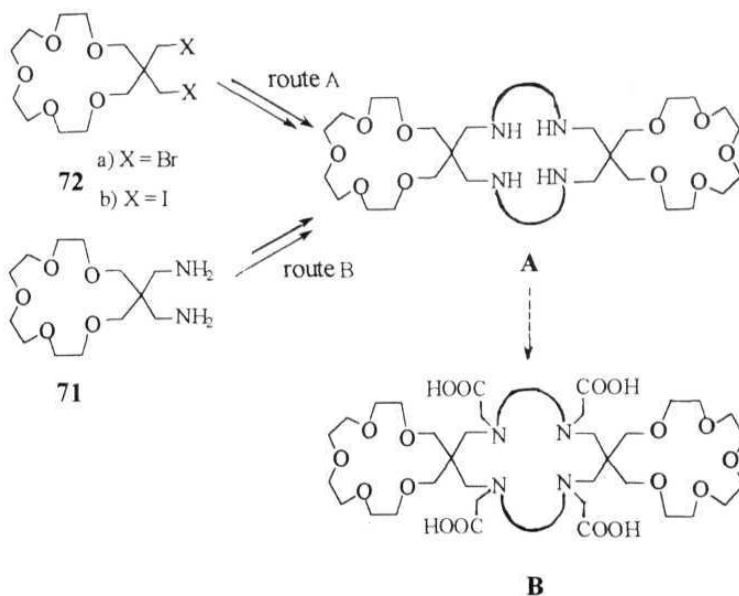
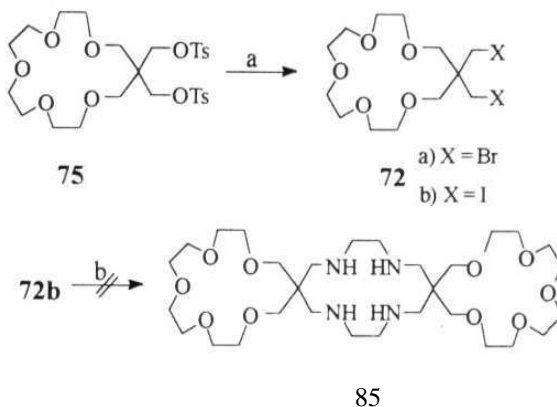


Figure 13

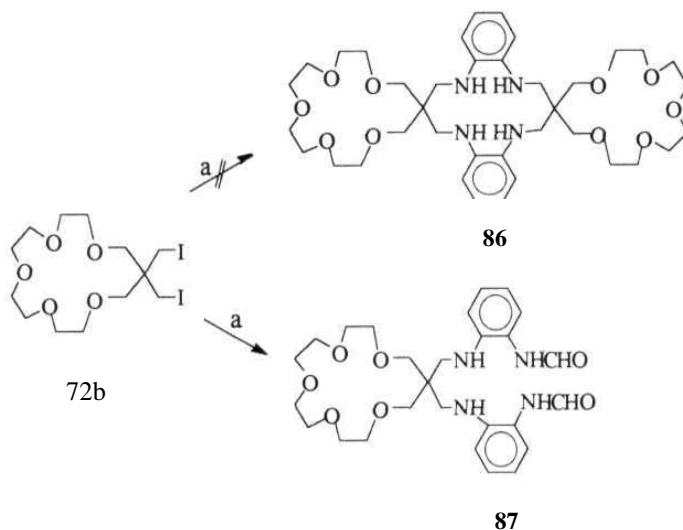
The successful realization of the synthesis of 79 and 82 prompted us to take up the synthesis of multiloop aza crown ethers of the type A (Fig 13). These compounds could then serve as precursors to the polyaminopolycarboxylic acids B, analogous to the tetraazamacrocyclic compounds DOT A 26 and TETA 27. As depicted in Fig. 13, this was sought to be achieved from either the crown dihalides 72a, b or the crown diamine 71. Accordingly, the dibromide 72a and the crown diiodide 72b were prepared from the crown ditosylate 75 (Scheme 20). Thus, the crown ditosylate 75 was reacted with sodium bromide or sodium iodide in diethylene glycol at 150° to furnish the crown dibromide 72a or the crown diiodide 72b in good yields. The products were characterized by elemental analysis and spectral data. In the ^1H NMR and ^{13}C NMR spectra, the tosylate signals of 75 were absent 72a & b showed the bromomethyl carbon at 35.9 ppm and iodomethyl carbon at 13.58 ppm, respectively, in the ^{13}C NMR spectra.

With the halides 72a and 72b in hand, the linking process to generate a tetraazamacrocyclic cycle was attempted. On reaction with ethylenediamine, 72b gave a complex, inseparable mixture. When nickel (II) sulphate was added to act as a template in the above reaction, a more polar, complex mixture resulted. Once again, no separation could be achieved. Examination of the crude IR and ^1H NMR spectra of these products indicated the presence of crown ether protons. No further structural information could be obtained



Scheme 20. *Reagents and conditions:* a) NaBr or NaI, diethylene glycol, 50°, 12 h, b) ethylenediamine, 100°, 12 h.

The crown diiodide **72b** on heating with *o*-phenylenediamine in DMF furnished a complex mixture (Scheme 21). This mixture was partially separated by successive extraction with dichloromethane and methanol. The IR and ^1H NMR spectra of both dichloromethane and methanol soluble fractions showed signals corresponding to aromatic and crown ether protons. It was observed that the ^1H NMR spectra of these products contain a signal at 8.13 ppm corresponding to a formyl proton. From this it was deduced that the reaction resulted only in the formylated product **87** and that none of the desired product **86** was formed. Further purification and characterization of **87** was not attempted.



Scheme 21. *Reagents and conditions:* a) *o*-phenylenediamine, DMF, 120°, 72 h.

In order to avoid the formation of a mixture of products, the second approach was adopted starting from crown diamine 71. It is known in the literature that 1,3-diamines on reaction with biacetyl in the presence of nickel (II) sulphate and zinc (II) chloride form the nickel (II) complexes of [14]-cyclam 88⁶¹ (Fig. 14). Accordingly, when the crown diamine 71 was treated with one eq. of biacetyl, under the same conditions (Scheme 22), a buff

coloured solid was obtained. However, its elemental analysis and spectral data did not correspond to the expected nickel complex of the spiro crown

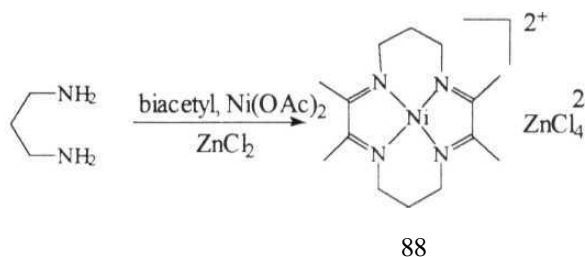
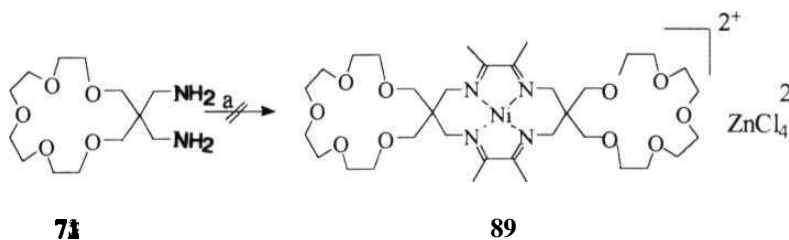


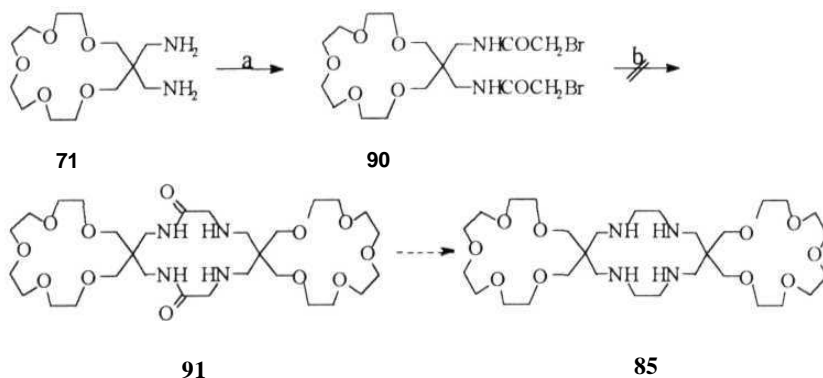
Figure 14

compound 89. The IR spectrum of the product mixture has absorption bands at 3134, 3047 and 1610 cm^{-1} and a strong ether band at 1093 cm^{-1} . The ^1H and ^{13}C NMR spectra do not display the signals corresponding to methyl protons of **89**.



Scheme 22. *Reagents and conditions:* a) biacetyl, HCl, MeOH, NiSO_4 , ZnCl_2 .

In another method, condensation of the crown diamine **71** with bromoacetic acid in the presence of DCC in dichloromethane afforded the crown bis(bromoacetamide) **90** in 60% yield (Scheme 23). The IR spectrum of **90** showed a strong absorption band at 3400 and 1760 cm^{-1} , indicating the presence of a secondary amide and the ^{13}C NMR spectrum had signals from the amide carbonyl carbon at 168.6 ppm and halocarbon at 29.12 ppm. When the crown bis(bromoacetamide) **90** was treated with the crown diamine **71** in ethanol at reflux temperature a complex resulted mixture from which the required product was not isolable. The IR spectrum of the crude mixture showed the carbonyl band at 1660 and the crown ether band at 1100 cm^{-1} .

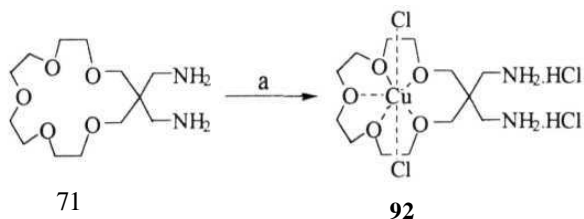


Scheme 23. Reagents and conditions: a) BrCH_2COOH , DCC, dichloromethane, 5° , 12 h; b) **71**, EtOH, reflux, 12 h

Finally, in order to obtain the spiro crown system present in 85, the crown diamine 71 was treated with oxalyl chloride in dichloromethane as well as with diethyl oxalate in ethanol at reflux temperature. Both reactions, however, resulted in complex mixtures and their separation was not attempted.

At this stage we did not pursue further the synthesis of spiro crowns in the light of all these failures.

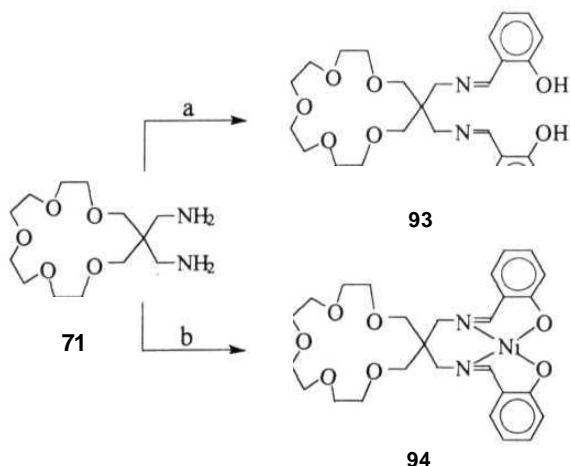
In order to investigate the complexation ability of the crown diamine 71, preparation of some of its metal complexes were attempted. When a 1 : 1 mixture of the crown diamine 71 and cupric chloride in ethanol was heated, a light yellow coloured crystalline material of the crown diamine-copper complex 92 was obtained (Scheme 24). The IR spectrum of 92 showed the crown ether bands at 1100 cm^{-1} along with primary ammonium bands at $3100\text{-}3000\text{ cm}^{-1}$. In the UV spectrum of 92, recorded in methanol,



Scheme 24. *Reagents and conditions:* a) $\text{CuCl}_2 \cdot 2\text{H}_2\text{O}$, EtOH, heat

an absorption maximum at 899 nm was seen. The complex analysed for $C_{13}H_{30}N_2O_5CuCl_4$, suggesting that both amine nitrogens are protonated. A plausible structure, with the copper (II) ion in the middle of the cavity, is shown in **92**.

When the crown diamine **71** was treated with salicylaldehyde in ethanol at reflux temperature for 24 h, the crown salicylaldimine **93** was obtained as a brown coloured syrup, found to be unstable on long standing (Scheme 25). The structure of **93** was confirmed by the presence of a strong phenolic OH absorption band at 3300 cm^{-1} in the IR spectrum and also by



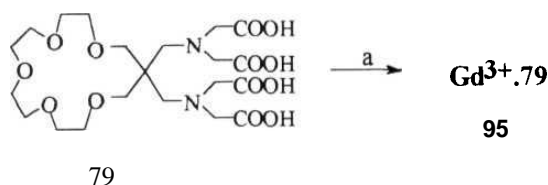
Scheme 25. Reagents and conditions: a) salicylaldehyde, EtOH, heat, 12 h; b) salicylaldehyde, NiSO₄, EtOH, heat, 48 h.

the presence of phenolic OH protons, aromatic proton and imine proton signals at 8.38 , at 7.27-7.35 and at 6.84-6.97 ppm, respectively, along with the crown proton signals in the ^1H NMR spectrum

It is known in the literature that nickel (II) complexes of Schiff's bases can be prepared using nickel (II) sulfate or nickel (II) acetate as a template.⁶² Thus, the crown salicylaldimine nickel (II) complex 94 was prepared by refluxing the crown diamine 71 and salicylaldehyde in ethanol, followed by the addition of nickel (II) sulfate. The resulting dark green complex was purified by column chromatography. Identification of the product rests on its spectral data. The UV spectrum of the complex 94 in methanol showed an absorption maximum at 585.4 nm ($\epsilon = 68$), typical of a d-d transition.⁶² The ^1H NMR spectrum contains signals at 6.99-7.22 ppm corresponding to the aromatic protons, the imine protons as a triplet at 6.50 ppm along with the crown proton signals at 3.41-3.64 ppm. It was also observed that in the ^1H NMR spectrum of 94, a signal at 8.38 ppm corresponding to the phenolic OH proton of the free crown salicylaldimine 93 was absent, indicating the participation of the phenolic oxygen in complexation.

The crown diaminetetraacetic acid- Gd^{3+} complex 95 was prepared by treating 79 with gadolinium oxide (Scheme 26). The light pink coloured solid Gd^{3+} complex 95 was identified by the presence of a carboxylate absorption band at 1600 cm^{-1} in the IR spectrum. Broadening of signals in the ^1H NMR spectrum, because of paramagnetic nature of the Gd^{3+} ion, was

observed The ESR spectrum of 95 also showed a broad symmetric signal with a g value of 2.06, close to the reported Gd^{3+} g value of 1.991 for gadolinium trichloride doped in lanthanum trichloride.⁶³



Scheme 26. *Reagents and conditions:* a) Gd_2O_3 , H_2O , heat.

Determination of Acid Dissociation and Stability Constants.

Due to the importance of gadolinium complexes of polyaminopolycarboxylic acid chelating agents like DTPA 24, DOTA 25 and TETA 26 in MRI studies, we synthesized novel chelating ligands containing both crown and polyaminopolycarboxylic acid functionalities, such as the crown diamine 71, the crown diaminetetraacetic acid 79, the crown tetraamine 78 and the crown tetraaminehexaacetic acid 82 and studied their complexation abilities with different metal ions in solution. Crown ethers are known to form stable complexes with lanthanide metal ions with different stoichiometries⁶⁴. Our attention in the study of the complexation abilities of 71, 79 and 84 was also focussed mainly on the lanthanides. The acid dissociation and chelate stability constants were measured by the

potentiometric titration method All the constants were determined by using the computer program SCOGS 2 ⁶⁵

Initially, the acid dissociation constants were calculated and were used for determining the stability constants The dissociation constants were calculated using the methods of Jonassen⁶⁶ and Martell⁶⁹ and were used as rough estimates for the input data in the program SCOGS 2 The measurements and calculations are discussed in the following paragraphs

1. Crown Diamine 71.

a) Determination of acid dissociation constants:

Using the potentiometric data (please see experimental for details) the acid dissociation constants of the protonated form of 71 were calculated by Jonassen's method⁶⁶ and pK_1 was obtained as 3.31 and pK_2 as 8.76. These values were used as input in the program, and after refinement, the acid dissociation constants obtained were $pK_1 = 2.86$ and $pK_2 = 8.79$.

From our results, it is evident that the first acid dissociation constant ($pK_1 = 2.86$) of the crown diamine 71 is very low when compared with that of 1,3-propanediamine ($pK_1 = 8.36$ and $pK_2 = 10.52$).⁶⁸ This indicates that the diprotonated species of crown diamine 71 is more acidic than that of 1,3-propanediamine The large difference in pK_1 's may be due to the effect of the crown ether moiety that is present in **71** It is known in the literature that in the case of 18-crown-6 ethers containing a pendant primary ammonium

group, hydrogen bonding is possible between the ammonium moiety and the crown cavity.¹⁵ A similar phenomenon is likely with the crown diamine 71 also. The monoprotonated species of the crown diamine 97 formed from the diprotonated species 96 by loss of one proton is stabilised by intramolecular complexation of the crown with the primary ammonium ion as shown in 98 (Fig. 15), thus shifting the equilibrium towards 97 and resulting in a low value for pK_1

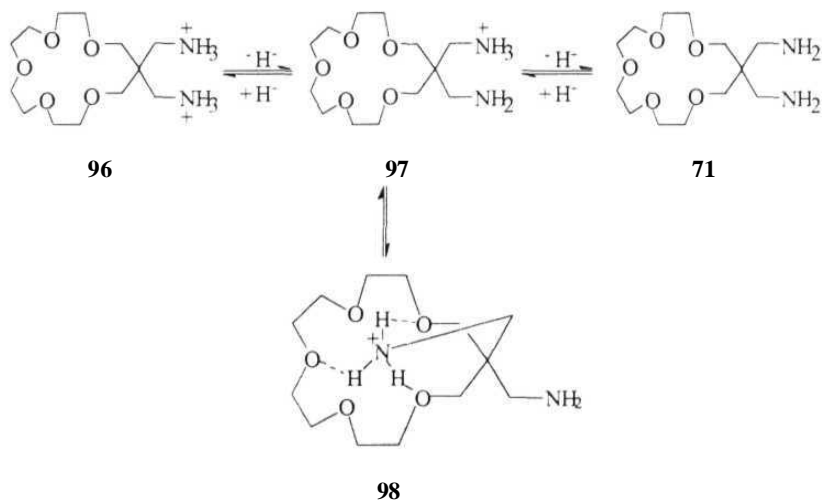


Figure 15

b) Determination of stability constants:

The potentiometric titrations of the ligand solution were performed in the presence of metal ions and the stability constants were determined by processing the potentiometric data using the SCOGS 2 program and the results obtained are listed in Table 3.

Table 3

Stability constants of the crown diamine 71 [25°, $\mu = 0.10 \text{ M}$ (Et₄NClO₄)]

	Co ²⁺	Ni ²⁺	Cu ²⁺	La ³⁺		Nd ³⁺	Sm ³⁺	Eu ³⁺	Gd ³⁺
log K ₁	9.74	8.38	9.95	3.01	5.97	3.48	3.55	3.85	3.62
log K ₂	3.95	3.64	4.80	4.51	4.05	2.34	6.01	4.45	6.41
log K ₃	1.35	—	·	3.60	4.60	5.90	5.21	4.90	5.37
log K _N	15.04	12.02	14.75	11.12	14.62	11.80	14.77	13.42	15.40

It was found that the crown diamine 71 forms 1 : 1 and 2 : 1 complexes with nickel and copper, whereas with cobalt and lanthanides it forms 3 : 1 complexes. It was also observed that the stability constants of the crown diamine 71 decreased when the titration was carried out using sodium perchlorate to maintain the ionic strength instead of tetraethylammonium perchlorate (Table 4). This effect of sodium may be attributed to the participation of the amino group in stabilisation of the sodium-crown complex.

Table 4
Stability constants of the crown diamine 71 at 25°

Metal ion	log K	
	$\mu = 0.10 \text{ M}$ Et_4NClO_4	$\mu = 0.10 \text{ M}$ NaClO_4
Cu^{2+}	14.75	10.05
Sm^{3+}	14.77	12.91
Gd^{3+}	15.40	12.94

2) Crown diaminetetraacetic acid 79.

a) Determination of acid dissociation constants:

The titration curve of the crown diaminetetraacetic acid 79 (pH vs a, where a is the number of moles of base added per mole of 79) showed the saturation point at $a = 6$ instead of 4, as might be expected for a tetrabasic acid. The reason for this behaviour is not clear. Therefore, 79 was first neutralised with 6 molar eq of sodium hydroxide and then titrated with perchloric acid. The acid dissociation constants were calculated according to Martell's and Schwarzenbach's graphical method⁶⁹ and the values obtained are $\text{pK}_1 = 1.60$, $\text{pK}_2 = 2.82$, $\text{pK}_3 = 6.20$ and $\text{pK}_4 = 9.43$. These values were used as input in the computer program SCOGS 2 and after refinement the values obtained are listed in Table 5

Table 5
Stability constants of crown diaminetetraacetic acid 79 [25°, $\mu = 0.10$
M (NaClO₄)]

pK₁	2.12
pK₂	2.80
pK₃	6.07
	9.35

The acid dissociation constants of the crown diaminetetraacetic acid 79 are comparable with that reported for trimethylenediaminetetraacetic acid

b) Determination of stability constants:

Processing the potentiometric titration data for the complexation of 79 with Cu²⁺ gave the value of log K as 12.67. Similarly, the stability constants of the crown diaminetetraacetic acid 79 with other metal ions were obtained and are listed in Table.6

Table 6
Stability constants of the crown diaminetetraacetic acid 79 (25°, μ = 0.10 M (NaClO₄)]

No. of displaceabl c protons	log K							
	metal ion							
	Ni ²⁺	Cu ²⁺	La ³⁺	Ce ³⁺	Nd ³⁺	Sm ³⁺	Eu ³⁺	Gd ³⁺
3	–	–	11.05	11.20	11.45	10.80	10.70	11.84
4	12.25	12.67	9.96	9.80	10.44	9.82	9.56	10.77

When the crown diaminetetraacetic acid 79 was titrated in the presence of cobalt perchlorate at pH 7.90, the metal ion started precipitating out and hence its stability constant was not processed. It was found that when the number of displaceable protons were given as 3 or 4 in the input to the SCOGS 2 program, the log K values obtained were different as shown in Table 6. However, it should be noted that with Ni²⁺ and Cu²⁺, convergencies were obtained only when the number of displaceable protons was taken as 4. Since DTPA 24 and DOT A 25 form charged complexes with lanthanides, it is more appropriate to use the data for log K corresponding to 4 displaceable protons for the acid 79 as well. It was also observed that in the case of the crown diaminetetraacetic acid 79, the effect of sodium ions was not significant (Table 7). This is perhaps due to the greater role played

by the carboxylate groups in metal ion complexation in 79 than the amino groups in 71.

Table 7
Stability constants of the crown diaminetetraacetic acid 79 at 25°

Metal ion	log K (No of displaceable protons 4)	
	$\mu = 0.10 \text{ M}$ (NaClO ₄)	$\mu = 0.10 \text{ M}$ (Et ₄ NClO ₄)
Cu ²⁺	12.67	1281
Sm ³⁺	982	10 40
Eu ³⁺	9.56	9.83
Gd ³⁺	10.77	10.38

3. Crown tetraaminehexaacetic acid 82.

a) Determination of acid dissociation constants:

When the titration data was processed with the computer program SCOGS 2, the last protonation constant $\text{p}K_6$ was found not to converge and hence $\text{p}K_6$ was calculated according to Martell's method.⁶⁹ The results obtained are listed in Table 8. The acid dissociation constants of the crown tetraaminehexaacetic acid 82 are also comparable with the reported data for triethylenetetraaminehexaacetic acid (H₆L)⁷⁰

Table 8
Acid dissociation constants of the crown tetraaminehexaacetic acid 82
[25°, $\mu = 0.10$ M (NaClO₄)]

pK ₁	2.14
pK ₂	3.16
pK ₃	4.54
pK ₄	7.06
pK ₅	9.51
pK ₆	10.85

b) Determination of stability constants:

The titrations were done in the presence of metal ions and the data was processed with the computer program and the stability constants thus obtained are tabulated in Table 9.

Table 9
Stability constants of the crown tetraaminehexaacetic acid 82 [25°, μ = 0.10 M (NaClO₄)]

Metal ion	log K (No. of displaceable protons)			
	6	5	4	3
Cu ²⁺	6.97	13.10	21.76	22.40
	4.44	11.33	19.60	21.32
Ce ³⁺	5.30	9.01	18.91	21.06
Nd ³⁺	5.48	10.00	19.70	21.46
Sm ³⁺	6.30	11.14	19.35	21.25
Eu ³⁺	6.64	10.48	19.75	21.53
Gd ³⁺	6.56	10.87	19.75	21.65

From these results, it is observed that the crown tetraaminehexaacetic acid 82 has better ligating ability with transition and lanthanide metal ions than the tetraacid 79 when the number of displaceable protons are 4. However, it was found that the chelate stability constants of the crown tetraaminehexaacetic acid 82, when calculated according to Martell's method,⁶⁷ were close to those of the stability constants obtained from the SCOGS 2 program when the number of displaceable protons are 4. However, since we did not prepare and isolate any of the metal complexes with the ligand 82, the stoichiometry of the complexes is not clear at this point

In conclusion, from our results, it is clear that the ligands 71, 79 and 82 have good ligating abilities. Additionally, they are highly soluble in water. It was observed that the crown diamine 71 forms 1 : 1 and 2 : 1 complexes with Ni^{2+} and Cu^{2+} , whereas with Co^{2+} and lanthanides it forms 3 : 1 complexes also. The stability constants of the crown diamine 71 with transition and lanthanide metal ions were found to be better than that of those with 1,3-propanediamine. In the presence of sodium ions, the stability constants were significantly reduced. This is attributed to the involvement of the amino groups in stabilizing the sodium complex with the crown ether moiety. The stability constants of the ligand 82 with lanthanides were close to those of DTPA (24) and HEHA (29). As expected, the crown tetraaminehexaacetic acid 82 is a better chelator than the crown diaaminetetraacetic acid 79.

Our preliminary results indicate that the lanthanide complexes of 71, 79 and 82 meet the basic requirements to function as MRI agents. Proton relaxation studies are needed before their suitability as MRI agents can be properly assessed. Toxicity tests of these compounds may also be required as crown ethers have been reported to be mildly toxic.⁷⁴

A second potential application of these complexes is as NMR shift reagents for NMR active alkali metal ions.^{29b} Since the crown ether moiety offers a ready site for alkali metal ion complexation, these complexes should act as efficient shift reagents. Their charge and side chains are added points in their favour.

Work on these lines is being currently pursued in our laboratory

EXPERIMENTAL

Experimental

All moisture sensitive reactions were carried out under nitrogen atmosphere. Reagents were transferred using standard septa-syringe techniques. All solvents were distilled from appropriate drying agents prior to use. All reagents were purified by appropriate methods before use. All organic extracts were dried using anhydrous MgSO_4 .

Solvents used for column chromatography were of commercial grade and were fractionally distilled. Column chromatography was performed using ACME 100-200 mesh silica gel and ACME alumina. Analytical thin layer chromatography (TLC) was performed on home made plates using ACME silica gel GF 254 grade containing 13% calcium sulphate as binder and were developed by shining ultraviolet light and/or by exposure to iodine vapours.

Melting points were determined by using a SUPERFIT melting point apparatus and are uncorrected. IR spectra were recorded on a Perkin-Elmer model 1310 spectrophotometer or JASCO FT-IR 5300 instrument. Solid samples were prepared as KBr wafers and liquid samples as a film between NaCl plates. ^1H NMR (100 and 200 MHz) and ^{13}C NMR (25 and 50 MHz) spectra were obtained with JEOL FX-100 and Bruker AF 200 instruments. All spectra were recorded using chloroform- d as solvent, unless otherwise mentioned. Chemical shifts are reported as δ values in parts per million relative to tetramethylsilane (0.0 ppm) as internal standard (^1H and ^{13}C).

Data are reported as follows: chemical shifts (multiplicity, integrated intensity, assignment). Elemental analyses were performed using a Perkin-Elmer 240C CHN analyser. The FAB mass spectra were obtained from CDRI Lucknow.

Preparation of Monobenzalpentaerythritol 3:

In an open 1 L three necked round bottomed flask pentaerythritol (80.0 g, 0.587 mole) and water (600 ml) were taken and the flask was equipped with an efficient mechanical stirrer and a dropping funnel containing benzaldehyde (67.0 g, 0.62 mole). The flask was heated until all the solid dissolved and was then allowed to cool undisturbed. When the solution was at rt, stirring was started and 4.5 ml of concentrated hydrochloric acid was added through the open neck of the flask, followed by 10 ml of benzaldehyde from the dropping funnel. When the precipitate of monobenzalpentaerythritol started forming, benzaldehyde was added dropwise. After the addition of benzaldehyde was complete, the mixture was stirred for an additional 2 h. The precipitate was collected on a Buchner funnel and washed with ice-cold water, made slightly alkaline with sodium carbonate. The solid was transferred to 1 L round bottomed flask, 350 ml water, slightly alkaline with sodium carbonate, was added and the mixture was heated to 100°. After about 10 min at this temperature, the hot mixture was filtered quickly through a fluted filter paper. The solid remaining on the paper was washed with 50 ml of hot water (made slightly alkaline with sodium carbonate). The combined aq. filtrates were cooled in an ice bath for several hours, and the crystals were collected on a Buchner funnel and dried

The dry product was heated under reflux for 15 min in an Erlenmeyer flask with 95 ml of toluene, and the hot mixture was allowed to cool to rt with continuous agitation to prevent formation of hard lumps. Finally, the mixture was cooled in an ice bath for 5 h, and the solid product was collected on a Buchner funnel and dried.

Yield: 23.0 g.

Solid, mp: 132-134°, (lit 134)¹¹

3-Phenyl-2,4,8,11,14,17,20,-heptaoxaspiro [5.15] heneicosane (67):¹¹

Monobenzalpentaoerythritol 3 (11.2 g, 50 mmol) and tetraethylene glycol ditosylate (25.1 g, 50 mmol) in separate 250 ml portions of THF were simultaneously added over a period of 8 h to a vigorously stirred, refluxing suspension of sodium hydride (3.0 g, 125 mmol) in 1 L of THF (high dilution conditions). After refluxing for an additional 12 h, the mixture was allowed to cool to rt and then quenched with methanol. The solvent was removed under reduced pressure, and the resulting residue was extracted with chloroform (3 x 250 ml). The combined extracts were evaporated and the residue was purified by chromatography on a silica gel column. First, 1 L of hexane was passed through the column to remove the mineral oil (NaH suspension). The product was then eluted with ethyl acetate.

Yield: 16.25 g (85%)

IR (neat): 3020, 2860, 1460, 1360, 1100, 750, 700 cm⁻¹.

¹H NMR (100 MHz):

6 7.18-7.50 (m, 5H, *ArH*), 5.38 (s, 1H, *PhCH*), 3.43-3.78 (m, 24H, *OCH₂CH₂O* and dioxane *CH₂*)

^{13}C NMR (25 MHz):

138.07, 128.83, 128.18, 126.06, 101.65, 71.30, 71.00, 70.35,
69.94, 69.18, 38.71 ppm.

15,15-Bis (hydroxymethyl)-1,4,7,10,13-pentaoxacyclohexadecane (68): ¹¹

A mixture of 67 (19.94 g, 0.052 mol) and 100 ml of aq. 0.1 N H_2SO_4 was refluxed for 12 h. The solution then was neutralised with aq. barium hydroxide, filtered and evaporated to dryness. The crude product was extracted with chloroform and then purified by column chromatography using acetone as eluent to obtain the crown diol 68 as a colourless, viscous oil.

Yield: 14.5 g (95%).

IR (neat): 3360, 2870, 1110 cm^{-1} .

^1H NMR (100 MHz):

5.368 (m, 24H, $\text{OCH}_2\text{CH}_2\text{O}$), 2.84 (br, 2H, OH)

^{13}C NMR (25 MHz):

72.90, 71.90, 65.20, 46.50 ppm

Attempted oxidation of the crown diol 68 to crown dialdehyde 69:

1. Pyridinium chlorochromate (PCC) Oxidation:

To a stirred suspension of PCC (220 mg, 1.02 mmol) in dichloromethane (1 ml) at rt was added the crown diol 68 (100 mg, 0.34 mmol) in dichloromethane (1 ml) and the contents stirred for 3 h. The resulting reaction mixture was passed through a fluorosil column and eluted with dichloromethane and the solvent was evaporated. The IR spectrum of

the residue obtained showed a strong **O-H** band (3350 cm^{-1}) and a weak carbonyl band (1740 cm^{-1}). However, **tlc** showed a complex mixture and further purification was not attempted.

When pyridinium dichromate (PDC) was used as the oxidising agent in dichloromethane, a complex mixture resulted. Chromic acid oxidation of the above crown diol 68 in pyridine/dichloromethane also did not give the required product

2. Oxidation of the crown diol 68 using *N*-chlorosuccinimide/dimethyl sulphide:

A solution of *N*-chlorosuccinimide (34 mg, 0.255 mmol) and dimethyl sulphide (0.025 ml, 0.425 mmol) in toluene (0.10 ml) was cooled to -25° . To this, a solution of crown diol 68 (50 mg, 0.17 mmol) in toluene (0.20 ml) was added and cooling was continued for a further 30 min. Triethylamine (25 mg, 2.50 mmol) in toluene (0.10 ml) was then added and the cooling bath was removed. After 5 min, diethyl ether (2 ml) was added and the organic layer was washed with 0.40 ml of 1% HCl followed by water and then dried, filtered and concentrated. The IR spectrum of the resulting residue showed both **O-H** and carbonyl bands and its further purification was not attempted.

IR (neat): $3350, 2950, 1720, 1470, 1110\text{ cm}^{-1}$.

Preparation of crown dimesylate 73:

To a stirred, ice cold solution of crown diol 68 (1.25 g, 4.25 mmol) in pyridine (30 ml) were added a few crystals of 4-(N,N-dimethylamino) pyridine followed by methanesulfonyl (III) chloride (1.95 g, 0.017 mol). The reaction mixture was stirred overnight and then poured into a mixture of dil HCl/ice-water mixture and extracted with dichloromethane. The organic layer was washed with dil. HCl solution, followed by saturated sodium bicarbonate solution and then dried, filtered and concentrated. The residue thus obtained was chromatographed to obtain the required crown dimesylate 73 as a syrup.

Yield: 1.13 g (60%).

IR(neat): 2900, 1470, 1360, 1180, 1110, 960, 850 cm^{-1} .

Reaction of crown dimesylate 73 with sodium azide 74:

To a solution of the crown dimesylate 73 (1.13 g, 2.50 mmol) in dimethylformamide (10 ml) was added sodium azide (490 mg, 7.53 mmol) and maintained at 60° overnight. The resulting reaction mixture was poured into water and extracted with diethyl ether. The organic layer was dried, filtered and concentrated. The IR spectrum of the resultant residue showed both azide and mesylate bands and further purification was not attempted.

IR (neat): 2900, 2100, 1470, 1370, 1190, 1115 cm^{-1} .

Preparation of crown ditosylate 75:

Sodium hydroxide (1.20 g, 30.0 mmol) dissolved in water (6 ml) and crown diol 68 (3.0 g, 10.2 mmol) in THF (12 ml) were placed in a flask and

the mixture was cooled in an ice bath with magnetic stirring. After 0.5 h of cooling, *p*-toluenesulfonyl chloride (4.08 g, 21.4 mmol) was added in small portions and the reaction mixture was stirred for 1 h at 5°. The reaction mixture was then poured into ice water and the white solid thus formed was filtered and dried. The product was recrystallised by dissolving it in minimum amount of ethyl acetate and a few drops of hexane were added till the solution became turbid. The contents were allowed to stand at rt to obtain the crown ditosylate **75** as a white crystalline solid.

Yield: 4.90 g (80%).

Solid, mp: 124-126°.

IR(KBr): 3050, 2900, 1600, 1353, 1187, 1119 cm⁻¹.

¹H NMR:

6.714-7.55 (AA'BB' system, *J* = 8 Hz, 8H, ArH), 3.78 (s, 4H, TsOCH₂), 3.23-3.35 (m, 20H, OCH₂CH₂O), 2.24 (s, 6H, TsCH₃)

¹³C NMR:

144.0, 132.0, 129.0, 128.0, 70.80, 70.50, 70.20, 68.50, 68.20, 44.0, 21.0 ppm.

Analysis:

Calcd for C₂₇H₃₈O₁₁S₂: C, 53.80; H, 6.35

Found: C, 53.71; H, 6.35.

Reaction of the ditosylate **75** with ethyl urethane:

To a solution of the crown ditosylate **75** (50 mg, 0.08 mmol) in dimethylformamide (2 ml) was added ethyl urethane (287 mg, 0.33 mmol)

and stirred at 60° overnight. The resulting reaction mixture was extracted with diethyl ether, dried and evaporated. The IR and NMR spectra of the crude product mixture showed that the starting material was recovered unchanged.

Reaction of the ditosylate 75 with benzylamine:

To a stirred solution of benzylamine (20 mg, 0.186 mmol) in dioxane (1 ml) was added ditosylate 75 (50 mg, 0.08 mmol) and refluxed overnight. To this 20% sodium hydroxide solution (5 ml) was added and the contents were extracted with ethyl acetate. The organic layer was dried and concentrated to give back unreacted starting material, as established by its IR spectrum

Preparation of the Crown diazide 74:

A mixture of the crown ditosylate 75 (3.0 g, 4.977 mmol) and sodium azide (1.95 g, 29.9 mmol) in dry dimethylformamide (15 ml) was heated to 120° for 3 days. The reaction mixture was cooled to rt, diluted with water and extracted with dichloromethane. The organic layer was dried and evaporated to give the product which was further purified by column chromatography using 50% ethyl acetate-hexane mixture as eluent.

Yield: 1.68 g (98%) as a colourless oil.

IR(neat): 2870, 2102, 1454, 1356, 1249, 1118 cm⁻¹.

¹H NMR:

6.340 (s, 4H, CH₂N₃), 3.46 (s, 4H, OCH₂CCH₂O), 3.64 (m, 16H, OCH₂CH₂O)

^{13}C NMR:

70.24, 51.71, 45.0 ppm.

Analysis:

Calcd for $\text{C}_{13}\text{H}_{24}\text{N}_6\text{O}_5$: C, 45.34, H, 7.02, N, 24.40

Found: C, 45.70; H, 7.24; N, 22.35.

(lower value for nitrogen may be because of thermal instability of the azide)

When the reaction was carried out for 3 days using 2.5 equivalents of sodium azide in dimethylformamide at rt as well as at 120° , the reaction was incomplete and yields were, thus lesser.

Attempted reductions of the crown diazide 74 to crown diamine 71:

1. Catalytic transfer hydrogenation:⁵²

A mixture of the crown diazide 74 (100 mg, 0.3 mmol) and 10% palladium on carbon in cyclohexene (10 ml) was stirred overnight at rt. The catalyst was filtered and the solvent was removed to obtain a residue. The residue did not contain any of the required product by IR and ^1H NMR analysis. No starting material was recovered as well.

IR (neat): 2900, 1460, 1280, 1120 cm^{-1} .

Hydrogenolysis the crown diazide 74 with 10% Pd/C in ethyl acetate also did not afford the required product.

2. Attempted reduction of the crown azide 74 using triphenylphosphine:⁵³

To a solution of the crown diazide 74 (100 mg, 0.3 mmol) in toluene (3 ml) was added a solution of triphenylphosphine (156 mg, 0.58 mmol) in

toluene (3 ml) and refluxed for 1 h. The solvent was removed and the residue was hydrolysed by heating for 1 h with a mixture of equal volumes of acetic acid (3 ml) and 40% hydrobromic acid (3 ml). The reaction mixture was diluted with water, neutralised with aq. ammonia, filtered and extracted with dichloromethane. The organic layer was dried and evaporated. The IR spectrum and tlc of the crude material showed the presence of the starting material only.

IR (neat): 2850, 2100, 1650, 1440, 1110 cm^{-1}

When the reaction was carried out using triphenylphosphine and 10% HCl in THF, the required product was not obtained

3. Reduction of the crown diazide 74 using LAH; Preparation of the crown diamine 71:

To a stirred suspension of LAH (1.12 g, 29.5 mmol) in dry THF (10 ml), the crown diazide 74 (1.7 g, 4.91 mmol) in dry THF (10 ml) was added dropwise at rt. After the addition was complete, the reaction mixture was refluxed overnight. The contents were cooled to rt and the reaction mixture was quenched by adding saturated sodium sulphate solution dropwise. A granular precipitate that formed was filtered off and washed with acetone. The filtrate was evaporated to obtain the product which was further purified by chromatography on a neutral alumina column using 50% ethyl acetate-acetone mixture as eluent to obtain the crown diamine 71 as a light yellow coloured syrup.

Yield: 1.40 g (98%).

IR(neat): 3315, 2866, 1662, 1450, 1359, 1120, 939, 879 cm^{-1}

^1H NMR:

8 3.60-3.65 (m, 16H, $\text{OCH}_2\text{CH}_2\text{O}$), 3.46 (s, 4H, $\text{OCH}_2\text{CCH}_2\text{O}$), 2.89 (s, 4H, CH_2NH_2)

^{13}C NMR

72.24, 70.77, 64.59, 44.59, 35.59 ppm

Analysis:

Calcd for $\text{C}_{13}\text{H}_{28}\text{N}_2\text{O}_5$ C, 53.40; H, 9.65; N, 9.58.

Found: C, 53.56; H, 9.64; N, 9.65.

When the crown azide 74 was treated with LAH in diethyl ether at rt as well as at reflux temperature and in THF at rt, no product was obtained

Reaction of the crown diamine 71 with hippuric acid: Preparation of the crown diamide 76:

To a stirred and ice cooled solution of hippuric acid (250 mg, 0.86 mmol) in dry dichloromethane (8 ml), DCC (445 mg, 2.15 mmol) was added and allowed to stir for 30 min at 5°. To this, the crown diamine 71 (385 mg, 2.15 mmol) in dry dichloromethane (8 ml) was added and stirred for 3 h under ice cooling and then at rt overnight. Water was added, the contents stirred for some time and the white solid thus formed was filtered off. The dichloromethane layer was separated from the aq. layer, dried and evaporated and the residue thus obtained was chromatographed using ethyl acetate as eluent to obtain the product 76 as a white coloured hygroscopic foamy solid.

Yield: 400 mg (76%).

IR (neat): 3300, 3050, 2850, 1650, 1530, 1300, 1110 cm^{-1} .

^1H NMR:

5 7.91-7.94 (d, 4H, *ArH*), 7.40-7.44 (d, 6H, *ArH*), 4 12-4.15 (d, 4H, COCH_2NH), 3.35-3.53 (m, 24H, $\text{OCH}_2\text{CH}_2\text{O}$)

^{13}C NMR

170 19, 167.50, 133 58, 13169, 128.47, 127 28, 127 10, 74.24, **71.42**, 70.50, 70.32, 69 72, 43.94 ppm

Analysis:

Calcd for $\text{C}_{31}\text{H}_{42}\text{N}_4\text{O}_9$: C, 60.56; H, 6.88; N, 9.11.

Found: C, 60.38; H, 6.95; N, 9.28.

Preparation of the crown N,N' -dibenzyldiamine 77:

To a stirred suspension of LAH (148 mg, 3.90 mmol) in dry THF (8 ml) was added dropwise the crown diamide 76 (400 mg, 0.65 mmol), in dry THF and the reaction mixture was refluxed overnight under nitrogen atmosphere. The reaction mixture was cooled to rt and then quenched by adding saturated sodium sulphate. The solid precipitate formed was filtered off and the cake was washed thoroughly with acetone. The combined organic layer was dried, concentrated and the residue was chromatographed to yield the pure product 77 as a brown coloured syrup.

Yield: 360 mg (96%).

IR(neat): 3317, 3028, 2868, 1454, 1358, 1120, 738, 700 cm^{-1} .

¹H NMR

8 7.29 (m, 10H, ArH), 3.76 (s, 4H, CH₂Ar) 3.59-3.63 (m, 20H, OCH₂CH₂O), 2.55-2.68 (m, 12H, CH₂NCH₂CH₂).

¹³C NMR

140.40, 128.20, 128.0, 126.00, 70.60, 70.20, 53.50, 51.80, **49.0, 47.0, 42.80** ppm.

Analysis:

Calcd for C₃₁H₁₅N₄O₅ C, 66.63; H, 9.02; N, 10.02

Found: C, 63.99; H, 8.77; N, 9.13

FAB mass:

m/z = 559 (M + H)⁺, 466, 345, 317, 285, 105, 91.

Preparation of hydrochloride salt of the crown dibenzylidiamine 77:

The crown N,N'-dibenzylidiamine 77 was dissolved in dichloromethane and HCl gas was passed till the solution became turbid. The solvent was evaporated to yield the salt as a white hygroscopic solid. Due to its high sensitivity to moisture, its characterization could not be carried out and hence the hexafluorophosphate salt was prepared.

¹H NMR (D₂O)

8 **7.31-7.34** (m, 10H, ArH), 4.14 (s, 4H, CH₂Ar), 3.15-3.53 (br, 32H, OCH₂CH₂O and CH₂NCH₂CH₂N)

Preparation of tetrakis(hexafluorophosphate) salt of the crown dibenzylidiamine 77:

To a solution of the hydrochloride salt of the crown dibenzylidiamine 77 in methanol, 4 eq. of ammonium hexafluorophosphate dissolved in methanol was added and to this solution, few drops of acetone were added until the solution becomes turbid and then allowed to stand at rt. The white solid thus formed was filtered, washed with methanol-acetone mixture and dried.

Solid, mp: Decomposes at 200°.

IR(KBr): 3337, 3038, 2430, 1624, 1431, 1271, 1186, 1132, 721 cm^{-1} .

^1H NMR (D_2O):

5 7.36 (s, 10H, ArH), 4.19 (s, 4H, CH_2Ar), 3.18-3.56 (m, 32H, $\text{OCH}_2\text{CH}_2\text{O}$ and $\text{CH}_2\text{NCH}_2\text{CH}_2\text{N}$).

Analysis:

Calcd for $\text{C}_{31}\text{H}_{54}\text{N}_4\text{O}_5\text{P}_4\text{F}_{24}$: C, 32.58, H, 4.76; N, 4.90

Found: C, 32.45; H, 4.72; N, 4.85.

**Attempted reactions for the cleavage of *N*-benzyl group of 77:
Preparation of the crown tetraamine 78:**

1. With formic acid:⁵⁴

To the crown $\text{N,N}'$ -dibenzylidiamine 77 (100 mg, 0.12 mmol) dissolved in 4.4% formic acid in methanol (10 ml) was added 10% Pd/C (200 mg) and stirred for 10 h. The catalyst was filtered and washed with methanol. The residue that was obtained after removal of solvent was found

to be the starting material from its IR spectrum. When the reaction was carried out using ammonium formate⁵⁶ and 10% Pd/C, sodium and liquid ammonia,⁵⁵ hydrogenation with 10% Pd/C in the presence of acetic acid⁵⁷ as well as with ethanolic HCl,⁵⁸ no cleavage of the TV-benzyl group occurred.

Preparation of Pearlman's catalyst (20% Pd(OH)₂/C):⁵⁹

Palladium chloride (1.0 g), carbon (2.40 g) and deionised water (20 ml) were mixed and stirred rapidly while being heated to 80°. To this, lithium hydroxide (LiOH H₂O, 50 mg) dissolved in water (2 ml) was added all at once and the heating stopped. The reaction mixture was stirred overnight at rt, filtered and the catalyst washed with 0.5% aq. acetic acid (20 ml). The cake was sucked as dry as possible and then dried in vacuum at 60°.

2. Cleavage of the crown N,N'-dibenzylidiamine 77 using Pearlman catalyst (20% Pd(OH)₂/C):

The crown N,N'-dibenzylidiamine 77 (90 mg, 0.161 mmol) in ethanol (5 ml) was hydrogenated using 20% Pd(OH)₂/C (200 mg) in a Parr apparatus at 65 psi for 4 h. The catalyst was filtered and the cake washed with methanol. The product, crown tetraamine 78 was obtained after evaporation of the solvent as a hygroscopic foamy solid.

Yield: 58 mg (95%).

IR (KBr): 3290, 2866, 1574, 1454, 1116, 943 cm⁻¹.

5 5.50 (br, NH), 3.35-3.51 (m, OCH₂CH₂O and NCH₂).

^{13}C NMR

73.69, 70.63, 70.45, 62.50, 59.23, 39.73 ppm.

The hygroscopic nature of the material precluded elemental analysis. When a catalytic amount or one equivalent (by weight) of 20% $\text{Pd}(\text{OH})_2/\text{C}$ was used, the cleavage was not effected.

Preparation of bis (tetraphenyborate) salt of the crown tetraamine 78:

To a solution of the crown tetraamine 78 (30 mg, 0.07 mmol) in 0.1M HCl was added an aq. solution of sodium tetraphenyborate (180 mg, 0.52 mmol). During the addition, a white solid was formed. After complete addition, the solid formed was filtered and dried. The product was recrystallised by dissolving it in minimum amount of ethyl acetate followed by adding ether till the solution became turbid. On cooling in the refrigerator a white solid was obtained which on drying became granular.

Yield: 60 mg (74%).

Solid, mp: 78-80°.

IR(KBr): 3425, 3055, 1579, 1479, 1427, 1109, 736, 707, 611 cm^{-1}

 ^1H NMR

5 7.80-7.06 (m, ArH), 3.61 (br, s).

Analysis:

Calcd for $\text{C}_{65}\text{H}_{80}\text{N}_4\text{O}_5\text{B}_2$: C, 76.61; H, 7.91; N, 5.49

Found: C, 76.19; H, 7.64; N, 5.18.

Preparation of the crown diaminetetraacetic acid 79:

Reaction of the crown diamine 71 with bromoacetic acid:⁴³

A solution of bromoacetic acid (238 mg, 1.71 mmol) in water (2.8 ml) was brought to pH 10 with 7 M KOH, maintaining the temperature below 5°. To this, a solution of the crown diamine 71 (100 mg, 0.34 mmol) in ethanol (5 ml) was added and the reaction mixture was heated to 70° for 4h. The pH of the mixture was maintained at 10 with 7 M KOH. After cooling to rt, the reaction mixture was acidified to pH 2 with 47% aq. HBr and was then evaporated. The resulting residue was dissolved in water (1 ml) and was loaded on a column of Amberlite 120 cation-exchange resin (washed with dil HCl). The column was eluted with water (500 ml) followed by 2% aq. NH₃ (500 ml). Both the water and aq. NH₃ fractions were concentrated to afford the product as a hygroscopic white, foamy solid. Further purification either by column chromatography or by preparative TLC was unsuccessful and hence a thorough characterization was not possible.

Yield 115 mg (64%, crude).

IR (KBr): 3408, 2922, 1743, 1439, 1249, 1109, 945 cm⁻¹.

¹H NMR (D₂O):

8.3.68-4.08 (m).

¹³C NMR:

172.59, 74.00, 73.87, 72.47, 62.65, 62.35, 62.00, 60.35, 60.24, 55.94, 45.88 ppm

Preparation of the crown tetraethyl ester 80:⁶¹

A mixture of the crown diamine 71 (100 mg, 0.34 mmol), ethyl bromoacetate (1.20 g, 7.18 mmol) and potassium carbonate was heated to 100° for 48 h. The reaction mixture was cooled to rt and the volatiles removed under vacuum. The residue thus obtained was extracted with dichloromethane and this organic layer was washed with bicarbonate solution, dried and concentrated to obtain the crude product which was further purified by column chromatography using ethyl acetate as the eluent.

Yield: 140 mg (64%).

IR(neat): 2872, 1734, 1662, 1464, 1371, 1186, 1116, 1032 cm^{-1} .

¹H NMR:

δ 4.09-4.19 (q, 8H, CH_2CH_3), 3.48-3.64 (m, 28H, $\text{OCH}_2\text{CH}_2\text{O}$ and NCH_2CO_2), 2.80 (s, 4H, CH_2N), 1.22-1.25 (t, 12H, CH_2CH_3)

¹³C NMR:

172.08, 70.60, 70.30, 60.30, 57.10, 56.80, 47.40 ppm.

Analysis:

Calcd for $\text{C}_{29}\text{H}_{52}\text{N}_2\text{O}_{13}$: C, 54.70, H, 8.23, N, 4.40.

Found: C, 53.63, H, 8.19, N, 3.40.

FAB mass

$m/z = 637 (\text{M} + \text{H})^+$, 307, 202, 130, 116.

Hydrolysis of the crown tetraethyl ester 80:⁶⁰

A mixture of the crown tetraethyl ester 80 (200 mg, 0.31 mmol) and KOH (350 mg, 6.24 mmol) in aq methanol (8 ml) was heated to 100°

overnight The reaction mixture was neutralised with conc HCl to pH 7, the solvent was removed under vacuum and the residue thus obtained was dissolved in methanol and filtered. Concentration of the solvent gave a hygroscopic foamy solid which was not amenable to further purification.

Yield: 145 mg (crude).

IR (KBr): 3410, 2191, 1734, 1635, 1402, 1248, 1107, 945 cm^{-1} .

^1H NMR (D_2O):

δ 3.50–3.80 (br).

Preparation of the crown tetrabenzyl ester 81:

A mixture of the crown diamine 71 (100 mg, 0.34 mmol), benzyl bromoacetate⁷¹ (470 mg, 2.05 mmol) and anhydrous potassium carbonate (285 mg, 2.05 mmol) in dry dimethylformamide (5 ml) was heated to 100° for 3 days. The mixture was concentrated under vacuum and the residue thus obtained was extracted with dichloromethane. The dichloromethane layer was dried and evaporated and the residue was chromatographed using ethyl acetate as eluent to obtain the product 81 as a brown coloured syrup with a fruity smell.

Yield: 165 mg (54%).

IR(neat): 3034, 2876, 1743, 1170, 1114, 736, 698 cm^{-1} .

^1H NMR:

δ 7.32 (s, 20H, ArH), 5.10 (s, 8H, CH_2Ar), 3.43–3.60 (m, 28H, $\text{OCH}_2\text{CH}_2\text{O}$ and NCH_2CO), 2.81 (s, 4H, CCH_2N).

^{13}C NMR:

171.76, 135.92, 128.56, 128.23, 77.80, 77.17, 76.53, 66.10,
57.27, 56.89, 47.43 ppm.

Analysis:

Calcd for $\text{C}_{49}\text{H}_{60}\text{N}_2\text{O}_{13}$: C, 66.49, H, 6.83, N, 3.16.

Found: C, 66.62; H, 6.88; N, 3.25.

When the reaction was carried out using benzyl iodoacetate in dimethylformamide under reflux conditions, the same product was obtained in 64% yield.

Hydrogenolysis of the crown ether tetrabenzyl ester 81:

The crown tetra-benzyl ester 81 (60 mg, 0.07 mmol) was hydrogenated with 20% $\text{Pd}(\text{OH})_2/\text{C}$ (100 mg) in dry ethanol (8 ml) at 65 psi in a Parr apparatus for 3 h. The catalyst was filtered off and washed with methanol. Upon evaporation of the solvent, the pure crown diaminetetraacetic acid 79 was obtained as a hygroscopic white solid.

Yield: 35 mg (95%).

IR (KBr): 3422, 2926, 1739, 1249, 1113 cm^{-1}

^1H NMR (D_2O):

6 3.20-3.80 (br, m).

^{13}C NMR.

170.45, 69.13, 71.22, 59.72, 57.90, 43.06 ppm.

Analysis:

Calcd for $C_{21}H_{36}N_2O_{13}$: C, 48.08; H, 6.91; N, 5.34.

Found: C, 48.15, H, 6.88; N, 5.31.

Preparation of the crown tetraaminehexaacetic acid 82 by direct alkylation with bromoacetic acid:⁴³

A solution of bromoacetic acid (850 mg, 6.12 mmol) in water (3 ml) was brought to pH 10 with 7 M KOH solution at 5° and then a solution of the tetraamine 78 (115 mg, 0.30 mmol) in ethanol (3 ml) was added. The reaction mixture was stirred at 70° for 12 h and during this time the pH of the reaction mixture was maintained at 10 by adding 7 M KOH solution. The reaction mixture was cooled to rt and acidified to pH 2 with 47% aq. HBr solution and then extracted with ether to remove the organic impurities. The resulting aq layer was concentrated, loaded on a column of cation exchange resin (IR 120, H^+ form) and the column was eluted first with water (100 ml) followed by 2% ammonia solution (100 ml). Both the fractions were combined and concentrated to yield the crude product 82, which once again was not amenable to further purification.

Yield: 140 mg (64%, crude)

IR(KBr): 3408, 2926, 1745, 1651, 1454, 1248, 1091, 945 cm^{-1} .

1H NMR (D_2O):

8.3.20-3.70 (br).

Preparation of the crown tetraaminehexaacetic acid hexaethyl ester **83**:⁶⁰

A mixture of the crown tetraamine **78** (75 mg, 0.20 mmol), ethyl bromoacetate (1.5 g, 9.00 mmol) and potassium carbonate (620 mg, 4.50 mmol) in dry dimethylformamide was heated to 100° for 3 days. DMF was removed in vacuum and the residue was extracted with dichloromethane. The dichloromethane extract was filtered, dried and concentrated and the residue was chromatographed using ethyl acetate as eluent to obtain the hexaester **83** as a dark brown syrup.

Yield: 100 mg (56.4%).

IR(neat): 2982, 1732, 1192, 1116, 1032, 733 cm⁻¹.

¹H NMR

5 4.20-4.13 (q, 12H, CH₂CH₃), 3.63-3.53 (m, 32H, OCH₂CH₂O & NCH₂CO₂), 2.80-2.75 (m, 12H, CH₂NCH₂CH₂N), 1.29-1.22 (t, 18H, CH₂CH₃)

¹³C NMR:

172.10, 171.20, 70.40, 70.0, 60.45, 60.16, 56.63, 55.23, 54.61, 52.94, 46.90, 14.31 ppm.

Analysis:

Calcd for C₄₁H₇₄N₄O₁₇: C, 55.01; H, 8.33, N, 6.26.

Found: C, 53.71; H, 7.62; N, 5.29.

FAB mass: m/z = 895 (M + H)⁺

Saponification of the hexaethyl ester **83**:⁶⁰

A mixture of the hexaethyl ester **83** (150 mg, 0.17 mmol) and KOH (500 mg) in aq. methanol (8 ml) was heated at 100° overnight. The reaction mixture was concentrated and loaded on a column of ion exchange resin (IR 120, H⁺ form). The column was eluted with water and then with 2% aq ammonia solution. These fractions were combined and concentrated in vacuum to afford the crude product, the crown tetraaminehexaacetic acid **82** as a yellow coloured hygroscopic solid. Its further purification was not successful for adequate characterization.

Yield: 90 mg (73% crude).

IR(KBr): 3423, 2935, 1728, 1614, 1462, 1390, 1116, 1030 cm⁻¹.

¹H NMR (D₂O):

6.3.00-3.90 (br).

Preparation of the crown tetraaminehexaacetic acid hexabenzyl ester **84**:

A mixture of the crown tetraamine **78** (100 mg, 0.26 mmol), benzyl bromoacetate (545 mg, 4.75 mmol) and anhydrous K₂CO₃ (330 mg, 2.38 mmol) in dry dimethylformamide (5 ml) was heated to 120° for 3 days. The solvent was removed in vacuum and the resulting residue was extracted with dichloromethane. The organic layer was dried, filtered and concentrated. The residue thus obtained was purified by column chromatography using ethyl acetate as the eluent to obtain the pure product **84** as a brown coloured syrup.

Yield: 40 mg (12%).

IR (neat): 3034, 2922, 1743, 1498, 1172, 736, 698 cm^{-1} .

^1H NMR:

6 7.28-7.36 (m, 30H, ArH), 5.09 (s, 12H, CH_2Ar), 3.45-3.55 (m, 32H, $\text{OCH}_2\text{CH}_2\text{O}$ & NCH_2CO_2), 2.78 (m, 12H, $\text{CH}_2\text{NCH}_2\text{CH}_2$).

^{13}C NMR

171.40, 170.60, 135.36, 128.90, 128.30, 128.0, 126.20, 70.50, 68.80, 66.30, 66.0, 55.0, 46.20 ppm.

Analysis:

Calcd for $\text{C}_{71}\text{H}_{86}\text{N}_4\text{O}_{17}$: C, 67.27; H, 6.83; N, 4.42

Found: C, 66.98, H, 6.62; N, 4.01.

When the reaction was carried out using benzyl iodoacetate in acetonitrile under reflux conditions and in the presence of cesium carbonate in DMF under reflux conditions, the yield of the product 84 was similar.

Hydrogenolysis of the crown tetraaminehexaacetic acid hexabenzyl ester 84:

Crown hexaester 84 (70 mg, 0.05 mmol) was hydrogenated using 20% $\text{Pd}(\text{OH})_2/\text{C}$ (150 mg) in ethanol (5 ml) at 65 psi for 5 h in a Parr apparatus. The catalyst was filtered off and washed with methanol. The methanol was evaporated to dryness to obtain the crown tetraaminehexaacetic acid 82 as an hygroscopic foamy material. Due to its hygroscopicity, its melting point could not be determined.

Yield: 30 mg (75%).

IR (KBr): 3429, 2926, 1736, 1651, 1385, 1249, 1109 cm^{-1} .

^1H NMR (D_2O):

δ 3.12-3.69 (br), 2.45-2.76 (br).

Analysis

Calcd for $\text{C}_{29}\text{H}_{50}\text{N}_4\text{O}_{17}$: C, 47.92, H, 6.93, N, **7.71**.

Found: C, **48.15**; H, 6.98; N, 7.78

Attempted reactions to prepare spiro crown compounds:

Preparation of the crown dibromide **72a**:

A mixture of the crown ditosylate **75** (1.0 g, 1.66 mmol) and sodium bromide (1.02 g, 9.95 mmol) in diethylene glycol (5 ml) was maintained at 150° overnight. The reaction mixture was cooled to 90° by adding ice slowly and then to 10° and finally poured into water and extracted with dichloromethane. The organic layer was washed with water, sodium bicarbonate solution and dried and evaporated to obtain the product **72a** as a pale yellow oil which was purified by column chromatography using ethyl acetate as eluent.

Yield: 670 mg (96%).

IR(neat): 2850, 1345, 1245, 1110 cm^{-1} .

^1H NMR:

δ 3.35-3.75 (m).

^{13}C NMR:

70.65, 70.47, 70.18, 69.77, 44.94, 35.90 ppm.

Analysis:

Calcd for $C_{13}H_{24}O_5Br_2$: C, 37.16; H, 5.75.

Found: C, 37.87, H, 5.93.

Preparation of the crown diiodide 72b:

A mixture of the crown ditosylate 75 (500 mg, 0.83 mmol) and sodium iodide (750 mg, 4.98 mmol) in diethylene glycol (5 ml) was heated to 150° for 12 h. The reaction mixture was cooled to rt, diluted with water and extracted with dichloromethane. The dichloromethane layer was washed with saturated sodium thiosulfate and with brine solution. The organic extract was dried, filtered and evaporated to obtain the product which was further purified by column chromatography using 50% ethyl acetate-hexane mixture as eluent to obtain the crown diiodide 72b as a light yellow coloured liquid.

Yield: 365 mg (85%).

IR (neat): 2845, 1440, 1340, 1230, 1110 cm^{-1} .

1H NMR:

5.372 (br, s, 18H, OCH_2CH_2O), 3.60 (s, 4H, OCH_2CCH_2O), 3.38 (s, 4H, CH_2I).

^{13}C NMR:

71.0, 70.65, 41.52, 13.58 ppm.

Reaction of the crown diiodide 72b with ethylenediamine:

a) A mixture of the crown diiodide 72b (90 mg, 0.21 mmol) and ethylenediamine (2 ml) was heated to 100° overnight. Excess

ethylenediamine was removed under vacuum to yield a complex **mixture**. The residue was dissolved in dichloromethane and the residue material was extracted with methanol.

Yield: 154 mg (crude)

Dichloromethane soluble product: 98 mg (syrup)

IR (neat): 3200, 2850, 1614, 1440, 1350, 1100 cm^{-1} .

^1H NMR:

δ 3.65-3.76 (m), 3.20 (s, br), 2.80 (s).

The above dichloromethane soluble product after neutralisation with ion exchange resin (IR 400, OH⁻ form) gave a syrup whose **homogeneity** could not be established.

Yield: 45 mg.

^1H NMR

δ 3.61-3.70 (m), 3.06 (s), 2.47-2.66 (m), 2.30 (s, br).

^{13}C NMR:

77.35, 73.58, 70.67, 70.48, 62.24, 59.28, 39.82, 39.59 ppm

Methanol soluble product: 56 mg (solid).

IR (KBr): 3350, 2900, 1640, 1100 cm^{-1} .

^1H NMR (D_2O):

δ 3.50-3.70 (m).

b) The reaction when carried out with the crown diiodide **72b** (115 mg, 0.26 mmol) and ethylenediamine (1 ml) in the presence of nickel (II) sulphate (35 mg, 0.13 mmol) yielded a complex mixture containing five

different products. The residue after removal of volatiles was taken up in different solvents to separate the products.

1 Hexane soluble product: 8 mg (syrup).

^1H NMR:

5 3.64-3.66 (m, crown-H), 3.20 (s), 3.55-3.75 (m).

After passing through ion exchange resin (IR 400, OH⁻ form), ^1H NMR was recorded again.

^1H NMR:

5 3.64-3.68 (m, crown-H), 3.07-3.08 (d), 2.50-2.70 (tt).

2 Dichloromethane soluble product: 25 mg (syrup).

IR(neat): 3398, 2868, 1668, 1454, 1114, 912, 731 cm^{-1} .

^1H NMR:

6 3.64-3.67 (m, crown-H), 3.30-3.40 (d), 2.80-2.90 (m), 2.50 (s, br).

3. Acetone soluble product: 30 mg (solid)

IR(KBr): 3445, 2967, 1664, 1381 cm^{-1} .

4. Methanol soluble product: 30 mg (solid).

IR (KBr): 2918, 1599, 1510, 1084, 1035 cm^{-1}

5. Pink coloured solid (insoluble in the above four solvents): 30 mg

IR(KBr): 3294, 2926, 1593, 1116, 1026, 704, 613 cm^{-1} .

Reaction of the crown diiodide 72b with *o*-phenylenediamine:

A mixture of the crown diiodide **72b** (100 mg, 0.19 mmol) and *o*-phenylenediamine (21 mg, 0.19 mmol) in dimethylformamide (5 ml) was heated to 120° for three days. The DMF was removed from the reaction

mixture under vacuum The residue obtained was taken in dichloromethane and undissolved material was extracted with methanol IR and NMR spectra of both the fractions showed only formylated product 87 no other tractable material.

Yield: 60 mg (crude).

Dichloromethane soluble: 12 mg (syrup)

IR(neat): 3300, 1650, 1100 cm^{-1}

^1H NMR

8 8.13 (s, 2H, CHO), 7 6.4-7.70 (m, 4H, ArH), 7.25-7.33 (m, 4H, ArH), 4.86 (s, 4H, CH_2N), 3.26-3.64 (m, 20H, $\text{OCH}_2\text{CH}_2\text{O}$)

Methanol soluble: 10 mg (white solid).

IR(KBr): 3300, 1650, 1100 cm^{-1}

^1H NMR:

5 8.00-8.10 (m), 7.70-7.80 (m), 7.50-7.60 (m), 7.20-7.30 (m), 3.56-3.65 (m).

Reaction of the crown diamine 71 with biacetyl:⁶¹

To a solution of the crown diamine 71 (500 mg, 1.71 mmol) in methanol (10 ml) was added conc. HCl (0.15 ml) and the reaction mixture was cooled to 5°. Biacetyl (200 mg, 2.32 mmol) was added and stirred for 30 min and then the reaction mixture was maintained at rt for 20 min followed by the addition of nickel (II) acetate (425 mg, 1.71 mmol). After 4 h, conc. HCl (0.15 ml) followed by ZnCl_2 (235 mg, 1.72 mmol) was added 10 min later, 3 ml of ether was added and the reaction mixture was kept at

rt. The solid thus formed was filtered and washed with **methanol-ether mixture**

Yield: 260 mg.

IR(KBr): 3134, 3047, 2926, 1610, 1498, 1354, 1093 cm^{-1} .

^1H NMR (D_2O):

5 3.51-3.57 (m,), 2.96 (s).

^{13}C NMR:

71.07, 69.60, 69.16, 68.99, 42.36, 40.25 ppm.

Reaction of the crown diamine 71 with bromoacetic acid: Preparation of the bis(bromoacetamide) 90:

To a stirred and ice cooled solution of the crown diamine 71 (200 mg, 0.68 mmol) in dichloromethane (10 ml) was added bromoacetic acid (240 mg, 1.72 mmol) followed by DCC (356 mg, 1.72 mmol) and this reaction mixture was stirred at 5° overnight. Water (10 ml) was added and after 15 min, the solid dicyclohexyl urea formed was filtered. The organic layer was separated, dried and evaporated to furnish a residue which was subjected to column chromatography using 50% acetone in ethyl acetate to afford the required product 90 as a foamy material.

Yield: 220 mg (60%).

IR (neat): 3200, 3050, 2900, 1160, 1560, 1100, 940, 740 cm^{-1}

^1H NMR

6 3.88 (s, 4H, CH_2COCH_2), 3.57-3.64 (m, 20H, $\text{OCH}_2\text{CH}_2\text{O}$), 3.35-3.38 (d, 4H, CH_2Br).

¹³C NMR

168.60, **74.77**, 71.06, 69.18, 68.47, 45.18, 38.76, 29.12 ppm

Reaction of the crown bis(bromoacetamide) 90 with the crown diamine 71:

A mixture of the bromoacetamide 90 (110 mg, 0.2 mmol) and the crown diamine 71 (50 mg, 0.17 mmol) in ethanol (10 ml) was refluxed overnight. Evaporation of the solvent afforded a brown coloured residue. TLC and IR spectrum of the residue showed a complex mixture whose purification was not attempted.

Yield: 80 mg (crude).

IR (neat): 3400, 3050, 2900, 1660, 1100 cm⁻¹.

¹H NMR

5 3.66(br, s).

¹³C NMR:

70.00-70.41 (m), 69.41-69.71 (m).

Reaction of the crown diamine 71 with oxalyl chloride:

To a stirred and ice cooled solution of the crown diamine 71 (50 mg, 0.17 mmol) in dichloromethane (5 ml) was added oxalyl chloride (48 mg, 0.37 mmol) and stirred at 5° for 1 h and then at rt overnight. Excess oxalyl chloride was removed from the reaction mixture and the residue was taken in dichloromethane and filtered. The undissolved material was dissolved in methanol. IR and NMR spectra of both fractions did not lead to any characterisable products.

Yield: 45 mg (crude).

Dichloromethane soluble product: 12 mg (syrup).

IR (neat): 3300, 2850, 1660, 1100 cm^{-1} .

^1H NMR:

δ 4.40 (br), 3.62 (s).

Methanol soluble product: 35 mg.

^1H NMR (D_2O):

5 3.50(br, m).

Reaction of the crown diamine 71 with diethyl oxalate:

A mixture of the diamine 71 (50 mg, 0.17 mmol) and diethyl oxalate (0.1 ml) in ethanol (2 ml) was refluxed overnight. The solvent was removed from the reaction mixture and the residue obtained was chromatographed eluting with 10% methanol in dichloromethane to afford a complex mixture, whose separation was not attempted.

Yield: 35 mg.

IR (neat): 2850, 1680, 1200, 1100, 920 cm^{-1} .

^1H NMR:

5 4.20-4.40 (t), 3.60 (br, s), 1.20-1.40 (m).

Preparation of the Copper (II) complex of the crown diamine92:

To the crown diamine 71 (175 mg, 0.60 mmol) in ethanol (3 ml), cupric chloride (306 mg, 1.80 mmol) in ethanol (2 ml) was added and the mixture was heated on a water bath. The undissolved cupric chloride was filtered off. To the filtrate, a small amount of ether was added and allowed to

stand at rt. The light yellow coloured crystals thus formed were filtered and dried

IR (KBr): 3450, 3200, 3100, 2850, 1607, 1100, 940 cm^{-1} .

Analysis:

Calcd for $\text{C}_{13}\text{H}_{30}\text{N}_2\text{O}_5\text{CuCl}_4$:

C, 31.24, H, 6.05; N, **5.61**; Cu, 12.71; Cl, 28.38.

Found: C, 30.52, H, **5.93**; N, **5.52**; Cu, **12.71**; Cl, 28.00.

The amounts of copper and chloride present in the complex **92** were estimated by gravimetry. UV spectra of the complex in methanol showed an absorption maxima at 893 nm.

Reaction of the crown diamine **71** with salicylaldehyde: Preparation of the crown salicylaldimine **93**:

A mixture of the diamine **71** (175 mg, 0.60 mmol) and salicylaldehyde (0.14 ml, **1.31** mmol) in ethanol (10 ml) was refluxed for 24h. The solvent removed and the residue thus obtained was subjected to column chromatography. Eluting with ethyl acetate yielded the required product **93**

Yield: 139 mg (65%), starting material recovered is 50 mg.

IR (neat): 3300, 2850, 1600, 1340, 1100, 740 cm^{-1} .

^1H NMR

8.38 (s, br, 2H, Ar-OH), **7.27-7.35** (m, 6H, ArH), 6.84-6.97 (m, 4H, ArH and N=CH), 3.63-3.70 (m, 24H, $\text{OCH}_2\text{CH}_2\text{O}$)

Preparation of the crown salicylaldimine nickel (II) complex 94:⁶²

To the crown diamine 71 (200 mg, 0.68 mmol) in ethanol (10 ml) salicylaldehyde (250 mg, 2.05 mmol) was added and refluxed for 2 days. To this reaction mixture, nickel (II) sulphate (100 mg) was added and refluxed for 30 min. The solvent was evaporated, the residue thus obtained was dissolved in dichloromethane and filtered. The residue obtained was chromatographed on basic alumina using ethyl acetate and acetone as eluent to afford the product 94 as a dark green coloured solid.

Yield: 80 mg.

IR(neat): 3350, 2850, 1610, 1540 cm^{-1} .

¹H NMR:

5.69–7.25 (m, 6H, ArH), 6.50 (t, 2H, ArH), 3.41–3.64 (m, 24H, $\text{OCH}_2\text{CH}_2\text{O}$).

UV (methanol): The UV spectrum of 94 shows an absorption maxima at 585.4 nm. The extinction coefficient ($\epsilon = 68$) is small, typical of a d-d transition.

Preparation of the gadolinium complex of the crown diaminetetraacetic acid 95:⁷³

A mixture of the crown diaminetetraacetic acid 79 (33 mg, 0.06 mmol) and gadolinium oxide (11.4 mg, 0.03 mmol) in water was refluxed overnight. Initially, gadolinium oxide was not soluble completely but on heating, it dissolved. The reaction mixture was cooled to rt and filtered and the water was removed under reduced pressure to dryness. The light pink

coloured solid thus obtained could not be satisfactorily recrystallised from various solvents.

Yield: 37 mg

IR (KBr): 3400, 2920, 1600, 1410, 1107 cm^{-1} .

^1H NMR (D_2O):

5 4.70(br), 3 59 (br), 3 22(br)

The ESR spectrum was recorded at room temperature. The g value obtained was 2.06. Literature⁶³ g value for Gd^{3+} (gadolinium trichloride doped in lanthanum trichloride) is 1.991.⁶³

Determination of Acid Dissociation and Stability Constants.

The dissociation constants of the crown diamine 71, the crown diaminetetraacetic acid 79 and the crown tetraaminehexaacetic acid 82 and the stability constants of these ligands with metal ions were obtained from potentiometric titration experiments. Potentiometric measurements of the ligands in the presence and in the absence of the metal ions were carried out with a pH meter equipped with glass and calomel reference electrodes and calibrated with standard pH 7 and pH 4 solutions. The readings are correct to ± 0.01 units. The temperature was maintained at $25.00 \pm 0.05^\circ$, and the ionic strength was adjusted to 0.10 by using tetraethylammonium perchlorate or sodium perchlorate as the supporting electrolyte. Anhydrous lanthanide metal chlorides were prepared according to the literature.⁷³ The concentrations of the experimental solutions were $2 - 5 \times 10^{-3}$ M in metal ions and ligands. Standard perchloric acid and carbonate and carbon dioxide free standard sodium hydroxide solutions were used for titrations. All the metal ion solutions were standardised by titrating against standard EDTA. All the experimental solutions were prepared using double distilled water.

Potentiometric titrations were performed in a nitrogen atmosphere by taking a known volume of ligand solution and the pH was measured after each addition of the titer base or acid in the presence as well as in the absence of the metal ions. All the experimental titrations were checked for consistency. The pH meter was calibrated for pH 7 and pH 4 before each titration.

All the calculations were performed using the computer program SCOGS 2 reported by Perrin and Stunzi.⁶⁵ All the calculations were carried out in two steps. First, the dissociation constants of the ligands were determined and in the second step, these constants were used for the determination of the stability constants. In all the computer calculations the standard deviations were of the order of 2×10^{-2} . All the dissociation and stability constants are correct to ± 0.01 log units. Initially all the dissociation constants were calculated according to Jonassen⁶⁶ Schwarzenbach and Martell⁶⁹ and were used as input data in the SCOGS 2 program.

The experiments were performed first with the crown diamine 71 followed by crown diaminetetraacetic acid 79 and crown tetraaminehexaacetic acid 82 and the data obtained are given below.

1. Crown Diamine 71.

1. Determination of acid dissociation constants ($\mu = 0.10$ M, Et_4NClO_4):

5 ml of 5×10^{-3} M crown diamine 71 solution was **titrated with** 0.0953 M perchloric acid. The ionic strength of the solution was maintained with tetraethylammonium perchlorate at $\mu = 0.10$. The titration data consists of two rows in which the first row gives the amount of titrant acid or base added (in ml) and the second row is the pH measured after each addition of the titrant.

acid added	PH
000	10.22
001	10.13
0.02	10.03
0.03	9.93
0.04	9.82
0.05	9.72
006	9.60
0.07	9.48
0.08	9.36
0.09	9.23
0.10	9.10
0.11	8.98
0.12	8.87
0.13	8.75

0.15	8.51
0.16	8.39
0.17	8.27
0.18	8.15
0.19	8.04
0.20	7.94
0.22	7.73
0.24	7.57
0.25	7.50
0.26	7.41
0.28	7.20
0.30	6.98
0.32	6.68
0.34	6.28
0.36	5.63

0.38	4.06
0.39	3.62
0.40	3.39
0.41	3.24
0.42	3.13
0.43	3.04
0.44	2.97
0.45	2.90
0.46	2.85
0.47	2.80
0.48	2.76
0.49	2.72
0.50	2.69
0.51	2.65
0.52	2.62

Initially the dissociation constants were determined by Jonassen's method⁶⁶ as $pK_1 = 3.31$ and $pK_2 = 8.76$. These pK 's values were given as rough estimates in the input to the SCOGS 2 program and after refinement the values were obtained as

$$pK_1 = 2.86$$

$$pK_2 = 8.79$$

2. Determination of stability constants ($\mu = 0.10 \text{ M}$, Et_4NClO_4):

a) With Cu^{2+} :

To 5 ml of $5 \times 10^{-3} \text{ M}$ diamine 71 solution, 0.04 ml of 0.1613 M copper (II) perchlorate solution was added and then titrated with 0.0953 M perchloric acid

acid added	pH
0.00	8.70
0.01	8.50
0.02	8.30
0.03	8.08
0.04	7.87
0.05	7.69
0.06	7.54
0.07	7.41
0.08	7.30
0.09	7.20
0.10	7.10
0.11	7.02

0.12	6.94
0.13	6.86
0.14	6.77
0.15	6.69
0.16	6.60
0.17	6.50
0.18	6.40
0.19	6.30
0.21	6.08
0.23	5.90
0.25	5.74
0.27	5.59
0.29	5.43

0.31	5.24
0.33	4.97
0.35	4.37
0.37	3.55
0.39	3.20
0.41	3.01
0.43	2.87
0.45	2.77
0.47	2.69
0.49	2.63
0.51	2.57
0.53	2.53

When this data was processed it was observed that the crown diamine 71 forms 1 : 1 and 2 : 1 complexes with Cu^{2+} and the values obtained are $\log K_1 = 9.95$, $\log K_2 = 4.80$ and $\log K_N = 14.75$.

b) With Co^{2+} :

To 4 ml of 5×10^{-3} M diamine 71 solution, 0.40 ml of 0.0159 M cobalt (II) perchlorate solution was added and then titrated with 0.0953 M perchloric acid solution.

acid added	PH
000	8.05
0.01	7.97
0.02	7.90
0.04	7.76
0.06	7.63
0.08	7.52
0.10	7.46
0.12	7.35
0.13	7.30

0.14	7.25
0.15	7.19
0.17	7.06
0.18	6.90
0.20	6.68
0.21	6.39
0.22	5.99
0.23	5.48
0.24	4.18
0.25	3.65

0.26	3.39
0.27	3.23
0.28	3.11
0.29	3.02
0.30	2.94
0.31	2.87
0.32	2.82
0.33	2.77
0.34	2.72
0.35	2.68

The stability constants obtained are $\log K_1 = 9.74$, $\log K_2 = 3.95$, $\log K_3 = 1.35$ and $\log K_N = 15.04$. It was observed that the crown diamine 71 forms 1:1, 2:1 and 3 : 1 complexes with Co^{2+} .

c) With Ni^{2+} :

To 4 ml of 5×10^{-3} M diamine 71 solution, 0.30 ml of 0.0193 M nickel (II) perchlorate solution was added and titrated with 0.095 M perchloric acid solution

acid added	PH
0.00	9.16
0.01	8.99
0.02	8.82
0.03	8.65
0.04	8.48
0.05	8.32
0.06	8.15
0.07	8.00
0.08	7.87
0.09	7.75

0.10	7.65
0.11	7.55
0.12	7.45
0.13	7.37
0.14	7.30
0.15	7.23
0.16	7.16
0.17	7.10
0.18	7.02
0.20	6.87
0.22	6.69

0.24	6.44
0.26	6.00
0.28	4.78
0.30	3.41
0.31	3.21
0.32	3.09
0.33	2.99
0.34	2.92
0.35	2.85
0.36	2.79
0.37	2.74

The stability constants obtained are $\log K_1 = 8.38$, $\log K_2 = 3.64$ and the $\log K_N = 12.02$. It is seen that the crown diamine 71 forms 1 : 1 and 2 : 1 complexes with Ni^{2+} .

d) With La^{3+} :

To 4 ml of 5×10^{-3} M diamine 71 solution, 0.20 ml of 0.0262 M lanthanum (III) chloride solution was added and titrated with 0.0953 M perchloric acid solution. The pH data is summarized below

acid added	PH
0.00	8.89
0.01	8.80
0.02	8.70
0.03	8.61
0.05	8.43
0.07	8.26
0.08	8.12
0.10	7.91
0.12	7.67
0.14	7.43

0.16	7.16
0.18	6.86
0.20	6.58
0.21	6.38
0.22	6.26
0.23	6.14
0.24	6.00
0.25	5.94
0.26	5.73
0.27	5.09
0.28	3.88

0.29	3.47
0.30	3.26
0.31	3.12
0.32	3.01
0.33	2.93
0.35	2.80
0.37	2.70
0.38	2.66
0.40	2.59
0.41	2.56
0.42	2.53

The stability constants obtained show that the crown diamine 71 forms 1 : 1, 2 : 1 and 3 : 1 complexes with La^{3+} and the values obtained are $\log K_1 = 3.01$, $\log K_2 = 4.51$, $\log K_3 = 3.60$ and $\log K_N = 11.12$.

e) With Ce^{3+} :

To 5 ml of $5 \times 10^{-3} \text{ M}$ diamine 71 solution, 0.20 ml of 0.04015 M cerium (III) chloride solution was added and titrated with 0.0953 M perchloric acid solution.

acid added	pH
0.00	7.80
0.01	7.68
0.02	7.57
0.03	7.47
0.04	7.37
0.05	7.26
0.06	7.16
0.07	7.02
0.08	6.91

0.09	6.77
0.10	6.62
0.11	6.43
0.12	6.22
0.13	5.97
0.14	5.70
0.15	5.34
0.16	5.04
0.17	4.70
0.18	4.06

0.19	3.58
0.20	3.37
0.21	3.23
0.22	3.13
0.23	3.04
0.24	2.98
0.25	2.92
0.26	2.87
0.27	2.82
0.28	2.78

The stability constants obtained are $\log K_1 = 5.97$, $\log K_2 = 4.05$, $\log K_3 = 4.60$ and $\log K_N = 14.62$.

0 With Nd^{3+} :

To 5 ml of 5×10^{-3} M diamine 71 solution, 0.20 ml of 0.0415 M neodymium (III) chloride solution was added and titrated with 0.0953 M perchloric acid solution

acid added	pH
0.00	8.10
0.02	8.00
0.04	7.92
0.06	7.84
0.08	7.76
0.10	7.70
0.12	7.63

0.16	7.49
0.18	7.39
0.20	7.29
0.22	7.15
0.24	6.96
0.26	6.70
0.28	6.40
0.30	6.01

0.32	5.81
0.34	4.85
0.36	3.62
0.38	3.26
0.40	3.07
0.42	2.94
0.44	2.84
0.46	2.76

The stability constants obtained are

$$\log K_1 = 3.48$$

$$\log K_2 = 2.34$$

$$\log K_3 = 5.90$$

$$\log K_N = 11.80.$$

g) With Sm^{3+} :

To 4 ml 5×10^{-3} M diamine 71 solution, 0.20 ml of 0.0241 M samarium (III) chloride solution was added and titrated with 0.0953 M perchloric acid solution

acid added	pH
0.00	8.18
0.01	8.02
0.02	7.86
0.03	7.77
0.04	7.60
0.05	7.47
0.06	7.34
0.07	7.22
0.08	7.09
0.10	6.78

0.11	6.60
0.12	6.44
0.14	6.24
0.15	6.19
0.17	6.12
0.19	6.00
0.20	5.91
0.21	5.85
0.23	5.71
0.24	5.54
0.25	5.10

0.26	4.07
0.27	3.62
0.28	3.35
0.30	3.09
0.32	2.93
0.33	2.87
0.34	2.81
0.35	2.76
0.36	2.72
0.37	2.68
0.38	2.65

The stability constants obtained are

$$\log K_1 = 3.55$$

$$\log K_2 = 6.01$$

$$\log K_3 = 5.21$$

$$\log K_N = 14.77.$$

h) With Eu^{3+} :

To 4 ml of 5×10^{-3} M **diamine** 71 solution, 0.30 ml of 0.02275 M europium (III) chloride solution was added and titrated with 0.0953 M perchloric acid solution.

acid added	pH
000	7.83
001	7.73
0.02	7.65
0.03	7.56
0.04	7.48
0.05	7.41
006	7.35
007	7.30
0.09	7.14
0 10	7.09
0 11	7.03

0.12	6.94
0.13	6.85
0.14	6.73
0.15	6.60
0.16	6.47
0.17	6.34
0.18	6.20
0 19	6.09
0.20	6.00
0.21	5.86
0.22	5.72
0.23	5.62

0.24	5.59
0.25	5.29
0.26	5.13
0.27	4.25
0.28	3.68
0.29	3.41
0.30	3.24
0.31	3.12
0.32	3.03
0.33	2.95
0.34	2.88
0.35	2.83

The stability constants obtained are

$$\log K_1 = 3.85$$

$$\log K_2 = 4.45$$

$$\log K_3 = 4.90$$

$$\log K_N = 13.42.$$

i) With Gd^{3+} :

To 4 ml of 5×10^{-3} M **diamine 71** solution, 0.20 ml of 0.02677 M gadolinium (III) chloride solution was added and titrated with 0.0953 M perchloric acid solution

acid added	pH
0.00	7.72
0.01	7.52
0.02	7.34
0.03	7.27
0.04	7.15
0.05	7.00
0.06	6.93
0.07	6.79
0.08	6.69
0.09	6.59
0.10	6.48
0.11	6.31

0.12	6.18
0.14	6.02
0.15	5.94
0.16	5.89
0.17	5.85
0.18	5.80
0.19	5.77
0.20	5.72
0.21	5.68
0.22	5.62
0.23	5.52
0.24	5.37
0.25	5.16

0.26	4.28
0.27	3.66
0.28	3.39
0.29	3.25
0.30	3.14
0.31	3.06
0.32	2.98
0.33	2.92
0.34	2.87
0.35	2.81
0.36	2.77
0.37	2.73
0.38	2.69

The stability constants obtained are $\log K_1 = 3.62$, $\log K_2 = 6.41$, $\log K_3 = 5.37$ and $\log K_N = 15.40$.

2. Determination of stability constants of the crown diamine 71 ($\mu = 0.10$ M, NaClO_4):

a) With Cu^{2+} :

5×10^{-3} M crown diamine 71 solution was prepared using sodium perchlorate as the supporting electrolyte with $\mu = 0.10$ M. To 4 ml of this diamine solution, 0.05 ml of 0.1613 M copper (II) perchlorate solution was added and titrated with 0.0953 M perchloric acid solution.

acid added	pH
0.00	7.37
0.02	7.16
0.04	6.97
0.05	6.88
0.06	6.79
0.07	6.71
0.08	6.62
0.09	6.54
0.10	6.46
0.11	6.39
0.12	6.30
0.14	6.15
0.15	6.08

0.16	6.01
0.17	5.95
0.18	5.90
0.20	5.83
0.21	5.77
0.22	5.71
0.23	5.64
0.24	5.57
0.25	5.50
0.26	5.43
0.27	5.33
0.28	5.24
0.29	5.11
0.30	4.94

0.31	4.65
0.32	4.16
0.33	3.71
0.34	3.44
0.35	3.27
0.36	3.14
0.37	3.04
0.38	2.97
0.39	2.90
0.40	2.84
0.41	2.79
0.42	2.74

The stability constant obtained is $\log K = 10.05$ and the stoichiometry corresponds to a ligand to metal ratio of **2 : 1**

b) With Sm^{3+} :

To 4 ml of 4×10^{-3} M diamine 71 solution, 0.20 ml of 0.0241 M samarium (III) chloride solution was added and titrated with 0.0953 M perchloric acid solution.

acid added	pH
0.00	8.16
0.01	8.02
0.02	7.89
0.03	7.78
0.04	7.68
0.05	7.58
0.06	7.50
0.08	7.34

0.10	7.20
0.12	7.05
0.14	6.82
0.16	6.46
0.18	6.12
0.20	5.72
0.21	5.25
0.22	4.34
0.23	3.70

0.24	3.38
0.25	3.19
0.26	3.05
0.28	2.84
0.30	2.70
0.32	2.60
0.34	2.52
0.36	2.45
0.38	2.39

The stability constant obtained is $\log K = 12.91$ and the stoichiometry corresponds to a ligand to metal ratio of **3 : 1**.

c) With Gd^{3+} :

To 4 ml of 4×10^{-3} M **diamine** 71 solution, 0.16 ml of 0.02677 M gadolinium (III) chloride solution was added and titrated with 0.0953 M perchloric acid solution.

acid added	PH
0.00	8.52
0.01	8.30
0.02	8.10
0.03	7.93
0.04	7.79
0.05	7.65
0.06	7.53
0.07	7.42

0.08	7.33
0.09	7.22
0.10	7.10
0.12	6.84
0.14	6.50
0.16	6.55
0.18	6.28
0.20	6.10
0.22	5.82

0.24	4.32
0.26	3.29
0.28	3.00
0.30	2.81
0.32	2.69
0.34	2.59
0.36	2.50
0.38	2.43
0.40	2.37

The stability constant obtained is $\log K = 12.94$ and the stoichiometry corresponds to a ligand to metal ratio of 3 : 1

2. Crown diaminetetraacetic acid 79.

1. Determination of the dissociation constants of crown diaminetetraacetic acid 79 ($\mu = 0.10$ M, NaClO_4):

4 ml of 1.8645×10^{-3} M crown diaminetetraacetic acid solution 79 was titrated with 0.3125 M sodium hydroxide solution and pH measured is given below.

base added	pH	0.070	360	0.125	9.12
0.000	2.55	0.075	3 86	0.130	9.54
0.010	2.62	0.080	4.22	0.140	9.98
0.020	2.71	0.085	4.73	0 150	10.26
0.030	2.80	0.090	5.38	0.160	10.47
0 040	2.93	0.095	5.91	0.170	10.62
0.045	2.99	0.100	6.30	0.180	10.73
0.050	3.07	0.105	6.61	0.190	10.81
0.055	3.17	0 110	6.91	0.200	10.87
0.060	3.28	0.115	7.32	0.220	10.97
0.065	3.42	0.120	8 17	0.240	11 05

The titration curve (pH vs a where a = number of moles of base added per mole of the ligand 79) showed the saturation point at a = 6 instead of 4 (see results and discussion) Hence, initially the crown diaminetetraacetic acid 79 solution was neutralised with 6 eq of 0.125 M

sodium hydroxide solution and then titrated with perchloric acid solution. The titration data was used for calculating the acid dissociation constants

4 ml of 1.8645×10^{-3} M crown diaminetetraacetic acid 79 solution was neutralised by adding 0.39 ml of 0.125 M NaOH and titrated with 0.0953 M perchloric acid solution.

acid added	pH
0.00	10.18
0.01	10.03
0.02	9.85
0.03	9.62
0.04	9.35
0.05	8.90
0.06	8.14
0.07	7.39
0.08	7.01
0.09	6.73

0.10	6.46
0.11	6.19
0.12	5.84
0.13	5.36
0.14	4.83
0.15	4.40
0.16	4.06
0.17	3.79
0.18	3.59
0.19	3.43
0.20	3.31

0.21	3.20
0.22	3.12
0.23	3.05
0.24	2.98
0.25	2.93
0.26	2.88
0.27	2.83
0.28	2.79
0.29	2.75
0.30	2.72

Initially, the acid dissociation constants of 79 were calculated using Schwarzenbach's and Martell's methods⁶⁹ and the values obtained are $pK_1 = 1.60$, $pK_2 = 2.82$, $pK_3 = 6.20$ and $pK_4 = 9.43$. These numbers were used as rough estimates in the program and after refinement, the values obtained are $pK_1 = 2.12$, $pK_2 = 2.80$, $pK_3 = 6.07$ and $pK_4 = 9.35$.

2. Determination of the stability constants of the crown diaminetetraacetic acid 79 ($\mu = 0.10$ M, NaClO_4):

a) With Cu^{2+} :

To 4 ml of 1.8645×10^{-3} M crown diaminetetraacetic acid 79 solution, 0.046 ml of 0.1613 M copper (II) perchlorate solution was added and then titrated with 0.3125 M sodium hydroxide solution

base added	pH
0.00	2.27
0.01	2.31
0.02	2.35
0.04	2.44
0.06	2.56
0.08	2.72
0.10	2.95
0.11	3.11

0.12	3.31
0.13	3.61
0.140	4.09
0.150	5.16
0.160	7.46
0.165	9.24
0.170	9.89
0.175	10.17
0.180	10.35

0.185	10.45
0.195	10.63
0.205	10.73
0.215	10.82
0.225	10.88
0.235	10.93
0.245	10.98
0.255	11.01

The stability constant obtained is $\log K = 12.67$ and the stoichiometry corresponds to a ligand-metal ratio of 1 : 1.

b) With Ni^{2+} :

To 4 ml of 1.8645×10^{-3} M crown diaminetetraacetic acid 79 solution, 0.4 ml of 0.01927 M nickel (II) perchlorate solution was added and titrated with 0.3125 M sodium hydroxide solution and pH measured is listed below.

base added	pH
0.000	2.33
0.020	2.41
0.040	2.51
0.060	2.63
0.080	2.79
0.100	3.06
0.120	3.60
0.125	3.83
0.130	4.14

0.12	3.31
0.135	4.66
0.140	5.55
0.145	7.60
0.150	8.55
0.155	9.21
0.160	9.70
0.165	10.04
0.170	10.21
0.175	10.33

0.185	10.45
0.180	10.43
0.190	10.59
0.200	10.70
0.210	10.77
0.230	10.84
0.250	10.94
0.270	11.01
0.280	11.04

The stability constant obtained is $\log K = 12.25$ and the stoichiometry corresponds to a ligand-metal ratio of 1 : 1

c) With La^{3+} :

To 4 ml of 1.5538×10^{-3} M crown diaminetetraacetic acid **79** solution, 0.25 ml of 0.02623 M lanthanum (III) chloride solution was added and titrated with 0.3125 M sodium hydroxide solution.

base added	PH
0.000	2.64
0.025	2.86
0.050	3.28
0.065	3.90
0.075	4.48
0.085	4.80
0.095	5.09
0.100	5.25

0.110	5.75
0.115	6.59
0.120	7.83
0.125	8.50
0.130	8.86
0.135	9.50
0.140	9.91
0.145	10.18
0.150	10.33

0.160	10.56
0.170	10.45
0.180	10.43
0.190	10.59
0.200	10.70
0.210	10.77
0.230	10.84
0.240	10.94
0.260	11.01

The stability constants obtained are
 with 4 displaceable protons, $\log K = 9.96$
 with 3 displaceable protons, $\log K = 11.05$.

d) With Ce^{3+} :

To 4 ml of 1.5538×10^{-3} M crown **diaminetetraacetic** acid 79 solution, 0.15 ml of 0.04015 M cerium (III) chloride solution was added and titrated with 0.3125 M sodium hydroxide solution.

base added	pH
0 000	2.64
0.020	282
0.040	3.11
0.050	3.35
0.055	3.52
0.060	3.73
0.065	3.96
0.070	4.14
0.075	4.29
0.080	4.42

0.085	4.54
0.090	4.68
0.095	482
0.100	4.99
0.105	5.23
0.110	5.70
0.115	6.76
0.120	7.75
0.125	841
0.130	9.12
0.135	9.71

0.140	10.03
0.145	10.22
0.150	10.33
0.160	10.53
0.170	10.65
0.180	10.74
0.200	10.88
0.220	10.97
0.240	11.03
0260	11 10
0.280	11 15

The stability constants obtained are
 with 4 displaceable protons, $\log K = 9.80$
 with 3 displaceable protons, $\log K = 11.20$.

e) With Nd^{3+} :

To 4 ml of 1.5538×10^{-3} M crown diaminetetraacetic acid 79 solution, 0.15 ml of 0.04149 M neodymium (III) chloride solution was added and titrated with 0.3125 M sodium hydroxide solution.

base added	pH
0.000	2.57
0.020	2.73
0.040	2.99
0.060	3.46
0.065	3.62
0.070	3.77
0.075	3.91
0.080	4.03
0.085	4.14
0.090	4.26

0.100	4.50
0.105	4.66
0.110	4.87
0.115	5.19
0.120	5.91
0.125	7.06
0.130	7.69
0.135	8.25
0.140	9.06
0.145	9.64
0.150	9.93

0.155	10.15
0.160	10.31
0.170	10.51
0.180	10.63
0.190	10.71
0.200	10.78
0.220	10.83
0.240	10.93
0.260	11.00
0.280	11.06
0.300	11.11

The stability constants obtained are
 with 4 displaceable protons, $\log K = 10.44$
 with 3 displaceable protons, $\log K = 11.45$.

η) With Sm^{3+} :

To 4 ml of 1.5538×10^{-3} M crown diaminetetraacetic acid 79 solution, 0.26 ml of 0.0241 M samarium (III) chloride solution was added and titrated with 0.3125 M sodium hydroxide solution.

base added	pH
0.000	2.61
0.020	2.78
0.040	3.06
0.060	3.51
0.070	3.75
0.075	385
0.080	396
0.085	4.07
0.090	4.17
0.095	4.30

0.100	4.46
0.105	4.69
0.110	5.07
0.115	5.75
0.120	6.67
0.125	7.26
0.130	7.99
0.135	885
0.140	9.46
0.145	982
0.150	10.04

0.155	10.21
0.160	10.32
0.170	10.48
0.180	10.59
0.200	10.74
0.220	10.84
0.240	10.92
0.260	10.98
0.280	11.04
0.300	11.09

The stability constants obtained are

with 4 displaceable protons, $\log K \approx 9.82$

with 3 displaceable protons, $\log K \approx 10.80$.

g) With Eu^{3+} :

To 4 ml of $1.5538 \times 10^{-3} \text{ M}$ crown diaminetetraacetic acid 79 solution, 0.28 ml of 0.02275 M europium (III) chloride solution was added and titrated with 0.3125 M sodium hydroxide solution.

base added	pH
0.000	262
0.020	2.79
0.040	3.05
0.060	3.48
0.070	3.71
0.075	3.81
0.080	3.92
0.085	4.02
0.090	4.13
0.095	4.27

0.100	4.44
0.105	4.68
0.110	5.10
0.115	5.87
0.120	7.00
0.125	7.55
0.130	8.38
0.135	9.04
0.140	9.60
0.145	9.92
0.150	10.13

0.155	10.27
0.160	10.37
0.170	10.52
0.180	10.62
0.190	10.70
0.200	10.77
0.220	10.87
0.240	10.95
0.260	11.02
0.280	11.08
0.300	11.13

The stability constants obtained are
 with 4 displaceable protons, $\log K = 9.56$
 with 3 displaceable protons, $\log K = 10.70$.

h) With Gd^{3+} :

To 4 ml of 1.5538×10^{-3} M crown diaminetetraacetic acid 79 solution, 0.23 ml of 0.02677 M gadolinium (III) chloride solution was added and titrated with 0.3125 M sodium hydroxide solution

base added	pH
0.00	2.50
0.02	2.68
0.04	2.92
0.06	3.34
0.07	3.56
0.08	3.77
0.09	4.00

0.10	4.28
0.11	4.78
0.12	6.36
0.13	7.91
0.14	9.37
0.15	10.07
0.16	10.43
0.17	10.64

0.18	10.76
0.19	10.84
0.20	10.91
0.21	10.97
0.22	11.00
0.24	11.10
0.26	11.14

The stability constants obtained are

with 4 displaceable protons, $\log K = 10.77$

with 3 displaceable protons, $\log K = 11.84$

2. Determination of the stability constants of the crown diaminetetraacetic acid **79** ($\mu = 0.10$ M, Et_4NClO_4):

a) With Cu^{2+} :

To 4 ml of 1.8645×10^{-3} M crown diaminetetraacetic acid **79** solution, 0.05 ml of 0.1613 M copper (II) perchlorate solution was added and titrated with 0.3125 M sodium hydroxide solution.

base added	pH
0.00	2.26
0.02	2.35
0.04	2.46
0.06	2.61
0.07	2.70
0.08	2.82
0.09	2.97

0.10	3.16
0.11	3.44
0.12	3.88
0.13	4.84
0.14	6.23
0.15	8.07
0.16	9.84
0.17	10.40

0.18	10.63
0.19	10.77
0.20	10.87
0.21	10.96
0.22	11.02
0.23	11.07

The stability constant obtained is $\log K = 12.81$.

b) With Sm^{3+} :

To 4 ml of 2.175×10^{-3} M crown diaminetetraacetic acid 79 solution, 0.40 ml of 0.0241 M samarium (III) chloride solution was added and then titrated with 0.3125 M sodium hydroxide solution.

base added	pH
0.00	2.49
0.02	2.59
0.04	2.70
0.06	2.87
0.08	3.09
0.10	3.37
0.11	3.51
0.12	3.65
0.13	3.77

0.14	3.89
0.15	4.05
0.16	4.20
0.17	4.44
0.18	4.87
0.19	4.74
0.20	7.12
0.21	8.36
0.22	9.25
0.23	9.87

0.24	10.26
0.25	10.50
0.26	10.65
0.27	10.74
0.28	10.80
0.29	10.85
0.30	10.91
0.31	10.96

The stability constants obtained are
 with 4 displaceable protons, $\log K = 10.40$
 with 3 displaceable protons, $\log K = 11.30$

c) With Eu^{3+} :

To 4 ml of 1.8645×10^{-3} M crown diaminetetraacetic acid 79 solution, 0.30 ml of 0.02275 M europium (III) chloride solution was added and titrated with 0.3125 M sodium hydroxide solution.

base added	pH
0.00	2.58
0.02	2.72
0.04	2.90
0.06	3.19
0.07	3.38
0.08	3.55
0.10	3.90

0.11	4.10
0.12	4.38
0.13	4.92
0.14	6.30
0.15	8.20
0.16	9.34
0.17	9.93
0.18	10.26

0.19	10.46
0.20	10.59
0.21	10.67
0.22	10.71
0.23	10.77
0.24	10.84
0.25	10.87
0.26	10.93

The stability constants obtained are
 with 4 displaceable protons, $\log K = 9.83$
 with 3 displaceable protons, $\log K = 11.04$.

d) With Gd^{3+} :

To 4 ml of 2.175×10^{-3} M crown diaminetetraacetic acid 79 solution, 0.30 ml of 0.02677 M gadolinium (III) chloride solution was added and titrated with 0.3125 M sodium hydroxide solution

base added	pH
0.00	2.55
0.02	2.62
0.04	2.75
0.06	2.90
0.07	3.00
0.08	3.11
0.09	3.25
0.10	3.39
0.11	3.52

0.12	3.64
0.13	3.77
0.14	3.92
0.15	4.07
0.16	4.26
0.17	4.54
0.18	5.03
0.19	6.34
0.20	8.58
0.21	9.47

0.22	996
0.23	1023
0.24	1040
0.25	1052
0.26	1061
0.27	1065
0.28	1070
0.29	1076
0.30	10.80
0.31	10.83

The stability constants obtained are
 with 4 displaceable protons, $\log K = 10.38$
 with 3 displaceable protons, $\log K = 11.20$.

3. Crown tetraaminehexaacetic acid 82.

1. Determination of acid dissociation constants of the crown tetraamine-hexaacetic acid 82 ($\mu = 0.10$ M, NaClO_4):

4 ml of 1.8989×10^{-3} M crown tetraaminehexaacetic acid 82 was titrated with 0.3125 M sodium hydroxide solution.

base added	PH
0,00	2.76
0.01	2.88
0,02	3.02
0.03	3.25
0.04	3.55
0.05	3.96
0.06	4.52
0.07	5.30

0080	640
0 090	7.86
0.100	891
0.110	9.56
0.120	10.04
0.130	10.33
0.135	10.42
0.140	10.49
0.145	10.55

0.150	10.59
0.160	10.70
0.170	10.78
0.180	10.85
0.190	10.91
0.200	10.95
0.210	10.99
0.220	11.03

When the data was processed in the program SCOGS 2 it was found that the last acid dissociation constant did not converge and hence the pK_6 was calculated according to Martell.⁶⁹ Thus, the pK 's obtained are $\text{pK}_1 = 2.14$, $\text{pK}_2 = 3.16$, $\text{pK}_3 = 4.53$, $\text{pK}_3 = 7.06$, $\text{pK}_5 = 9.51$ and $\text{pK}_6 = 10.85$. These acid dissociation constants were used in the determination of the stability constants.

2. Determination of the stability constants of the crown tetraaminehexaacetic acid 82 ($\mu = 0.10$ M, NaClO_4):

a) With Cu^{2+} :

To 3 ml of 1.8383×10^{-3} M crown tetraaminehexaacetic acid 82 solution, 0.035 ml of 0.1613 M copper (II) perchlorate solution was added and titrated with 0.3125 M sodium hydroxide solution.

base added	pH
0.00	2.64
0.01	2.77
0.02	2.95
0.03	3.20
0.04	3.58
0.05	4.14
0.06	484

0.07	5.38
0.08	5.98
0.09	7.00
0.10	8.36
0.11	9.22
0.12	9.84
0.13	10.25
0.14	10.51

0.15	1066
0.16	1078
0.17	1086
0.18	1094
0.19	11.00
0.20	11.05

The stability constants obtained are listed below.

log K (No. of displaceable protons)			
6	5	4	3
6.97	13.10	21.76	2240

b) With La^{3+} :

To 4 ml of 1.8989×10^{-3} M crown tetraaminehexaacetic acid 82 solution, 0.30 ml of 0.02623 M lanthanum (III) chloride solution was added and titrated with 0.3125 M sodium hydroxide solution

base added	pH
0.00	2.77
0.01	2.90
0.02	3.05
0.03	3.24
0.04	3.48
0.05	3.76
0.06	4.05

0.07	4.47
0.08	5.13
0.09	6.33
0.10	7.24
0.11	7.71
0.12	8.02
0.13	8.22
0.14	8.50

0.15	8.83
0.16	9.26
0.17	9.78
0.18	10.20
0.19	10.48
0.20	10.67
0.21	10.80
0.22	10.90

The stability constants obtained are

log K (No. of displaceable protons)			
6	5	4	3
4.44	11.33	19.60	21.32

c) With Ce^{3+} :

To 3 ml of 1.8383×10^{-3} M crown tetraaminehexaacetic acid 82 solution, 0.14 ml of 0.04015 M cerium (III) chloride solution was added and titrated with 0.3125 M sodium hydroxide solution.

base added	PH
0.00	2.85
0.01	3.04
0.02	3.30
0.03	3.62
0.04	4.08
0.05	4.91

0.06	6.12
0.07	6.89
0.08	7.39
0.09	7.76
0.10	8.30
0.11	9.10
0.12	9.79

0.13	10.33
0.14	10.57
0.15	10.77
0.16	10.91
0.17	11.01
0.18	11.09
0.19	11.17

The stability constants obtained are

log K (No. of displaceable protons)			
6	5	4	3
5.30	9.01	18.91	21.06

d) With Nd³⁺:

To 3.5 ml of 1.8383×10^{-3} M crown tetraaminehexaacetic acid 82 solution, 0.15 ml of 0.014149 M neodymium (III) chloride solution was added and titrated with 0.3125 M sodium hydroxide solution

base added	pH
0.00	2.78
0.01	2.94
0.02	3.14
0.03	3.38
0.04	3.68
0.05	4.10
0.06	4.76

0.07	5.80
0.08	6.62
0.09	7.16
0.10	7.53
0.11	7.90
0.12	8.53
0.13	9.30
0.14	9.89

0.15	10.35
0.16	10.65
0.17	10.83
0.18	10.96
0.19	11.06
0.20	11.15

The stability constants obtained are

log K (No. of displaceable protons)			
6	5	4	3
5.48	10.00	19.70	21.46

e) With Sm^{3+} :

To 4 ml of 1.8989×10^{-3} M crown **tetraaminehexaacetic** acid 82 solution, 0.30 ml of 0.0241 M samarium (III) chloride solution was added and titrated with 0.3125 M sodium hydroxide solution.

base added	PH
0.00	2.78
0.01	2.91
0.02	3.07
0.03	3.25
0.04	3.43
0.05	3.69
0.06	4.07

0.07	4.63
0.08	5.43
0.09	6.23
0.10	6.68
0.11	7.02
0.12	7.32
0.13	7.68
0.14	8.25

0.15	8.93
0.16	9.49
0.17	9.95
0.18	10.28
0.19	10.50
0.20	10.66
0.21	10.80
0.22	10.90

The stability constants obtained are

log K (No. of displaceable protons)			
6	5	4	3
6.30	11.14	19.35	21.25

0 With Eu^{3+} :

To 3.5 ml of 1.8383×10^{-3} M crown tetraaminehexaacetic acid 82 solution, 0.30 ml of 0.02275 M europium (III) chloride solution was added and titrated with 0.3125 M sodium hydroxide solution.

base added	pH
0.00	2.74
0.01	2.90
0.02	3.10
0.03	3.31
0.04	3.59
0.05	4.02
0.06	4.67

0.07	5.61
0.08	6.40
0.09	6.83
0.10	7.18
0.11	7.50
0.12	8.08
0.13	8.84
0.14	9.51

0.15	10.04
0.16	10.44
0.17	10.69
0.18	10.83
0.19	10.97
0.20	11.07

The stability constants obtained are

log K (No. of displaceable protons)			
6	5	4	3
6.64	10.48	19.75	21.53

g) **With Gd³⁺:**

To 3 ml of 1.8383×10^{-3} M crown tetraaminehexaacetic acid 82 solution, 0.20 ml of 0.04015 M gadolinium (III) chloride solution was added and titrated with 0.3125 M sodium hydroxide solution.

base added	PH
0.00	2.76
0.01	2.94
0.02	3.18
0.03	3.48
0.04	3.86
0.05	4.50

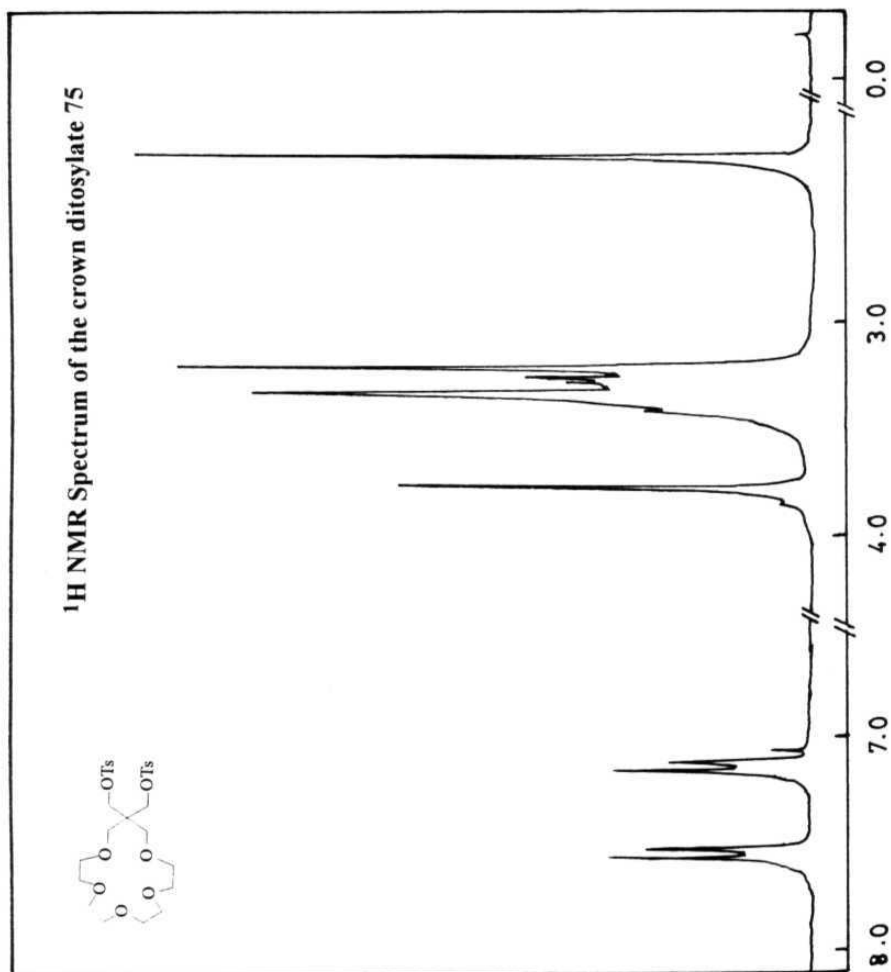
0.06	5.52
0.07	6.42
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0.09	7.33
0.10	7.91
0.11	8.85
0.12	9.59

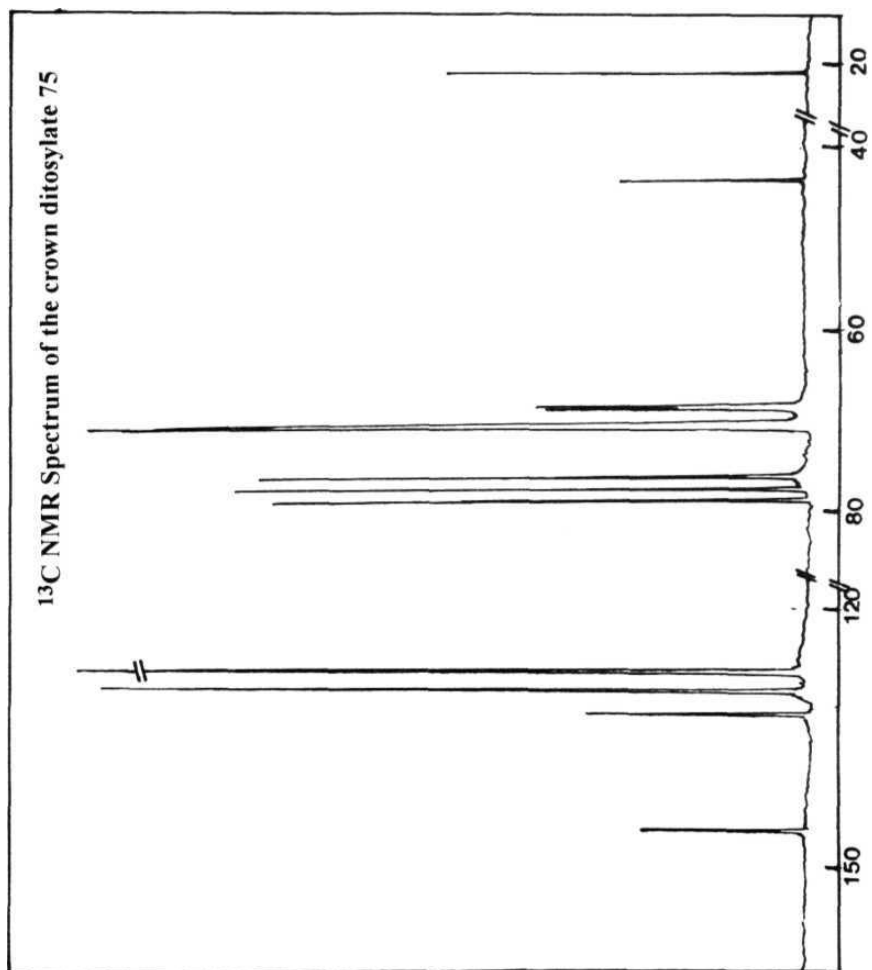
0.13	10.18
0.14	10.54
0.15	10.77
0.16	10.93
0.17	11.03
0.18	11.13

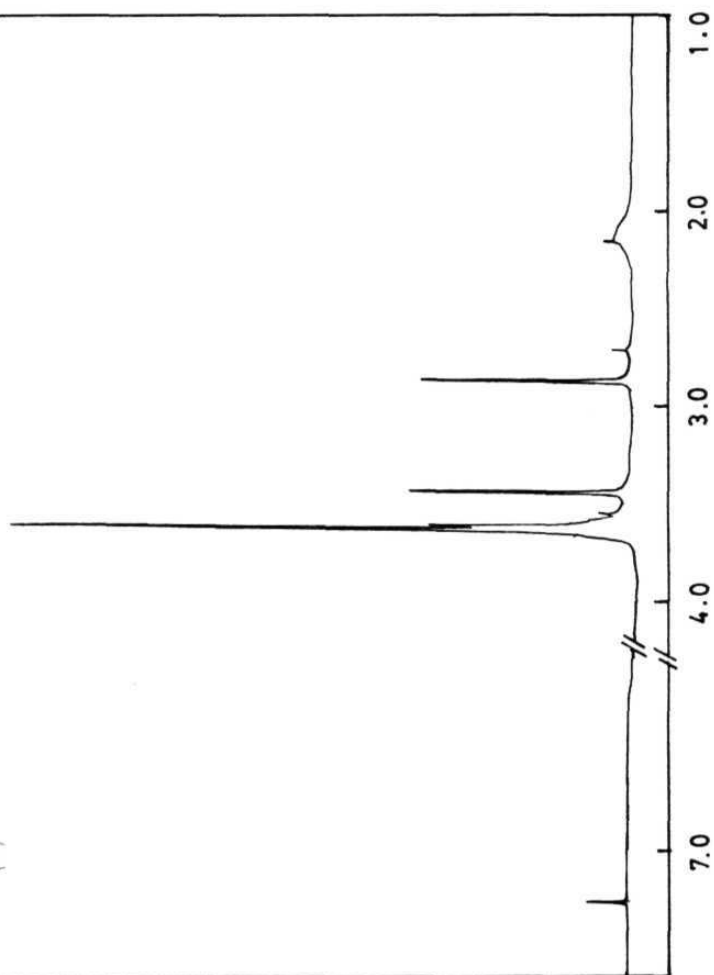
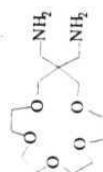
The stability constants obtained are

log K (No. of displaceable protons)			
6	5	4	3
6.56	10.87	19.75	21.65

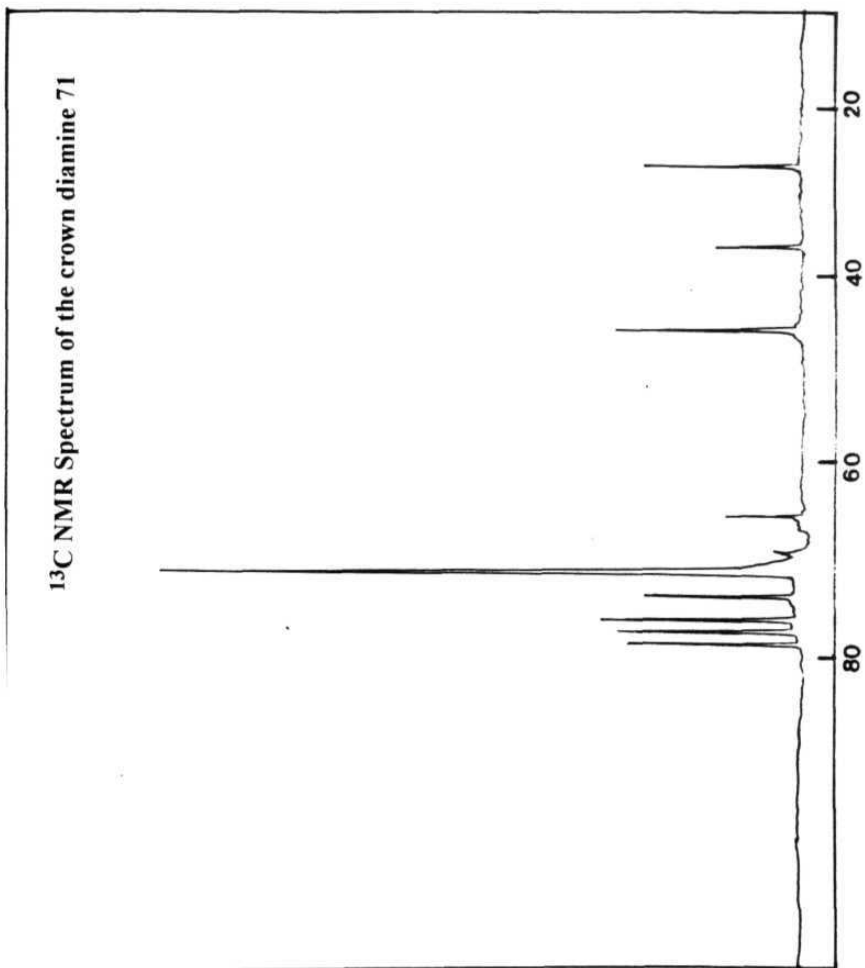
SPECTRA



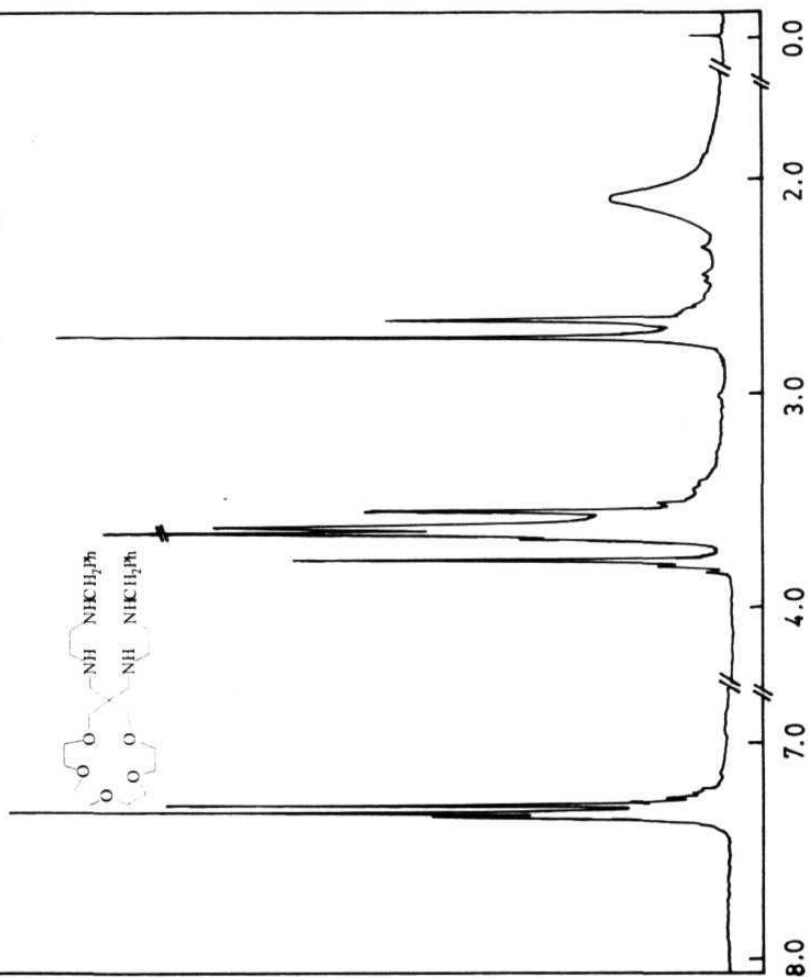


¹H NMR Spectrum of the crown diamine 71

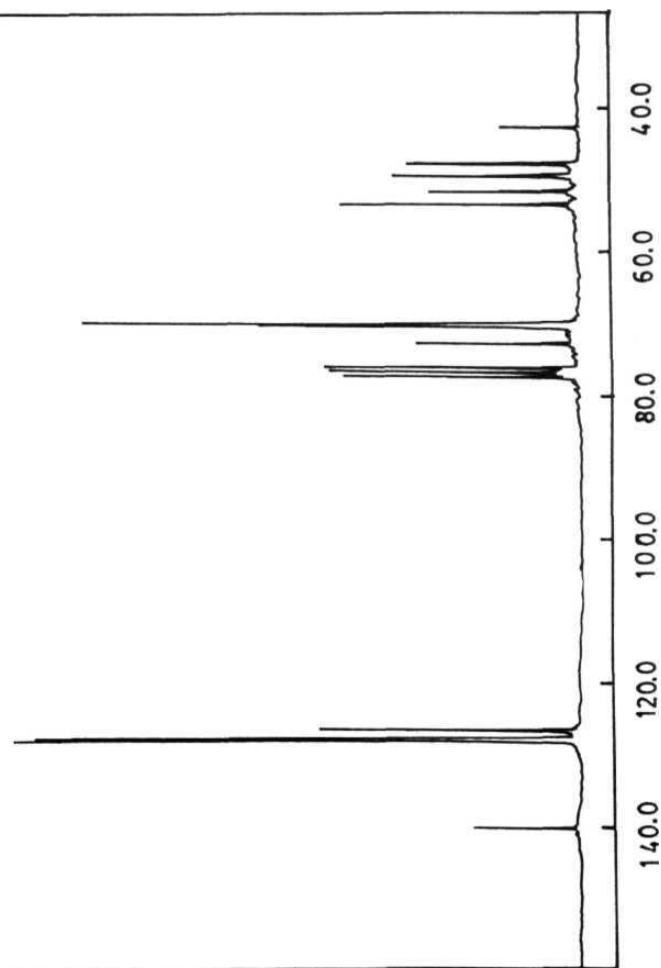
^{13}C NMR Spectrum of the crown diamine 71

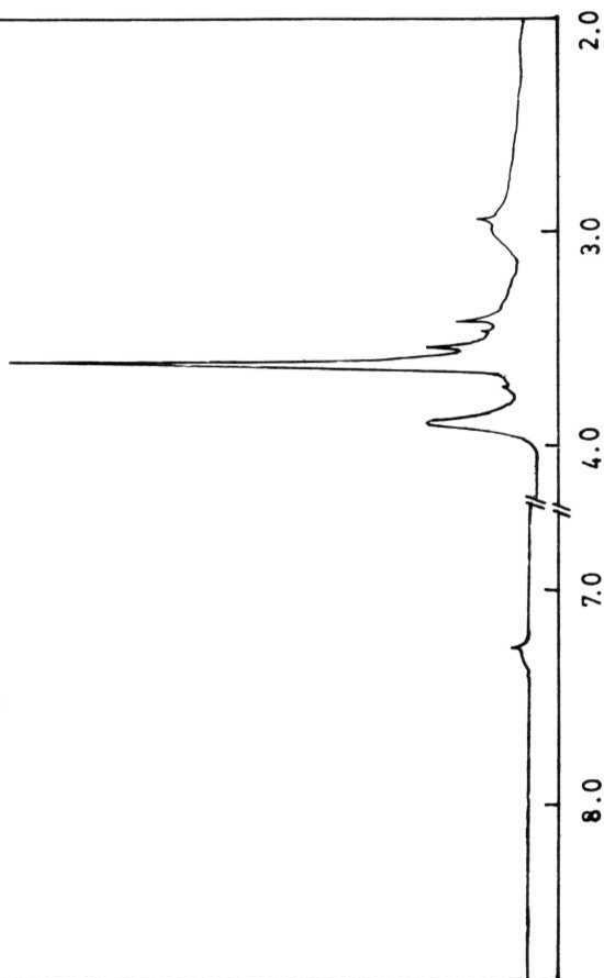
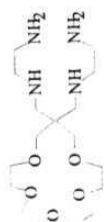


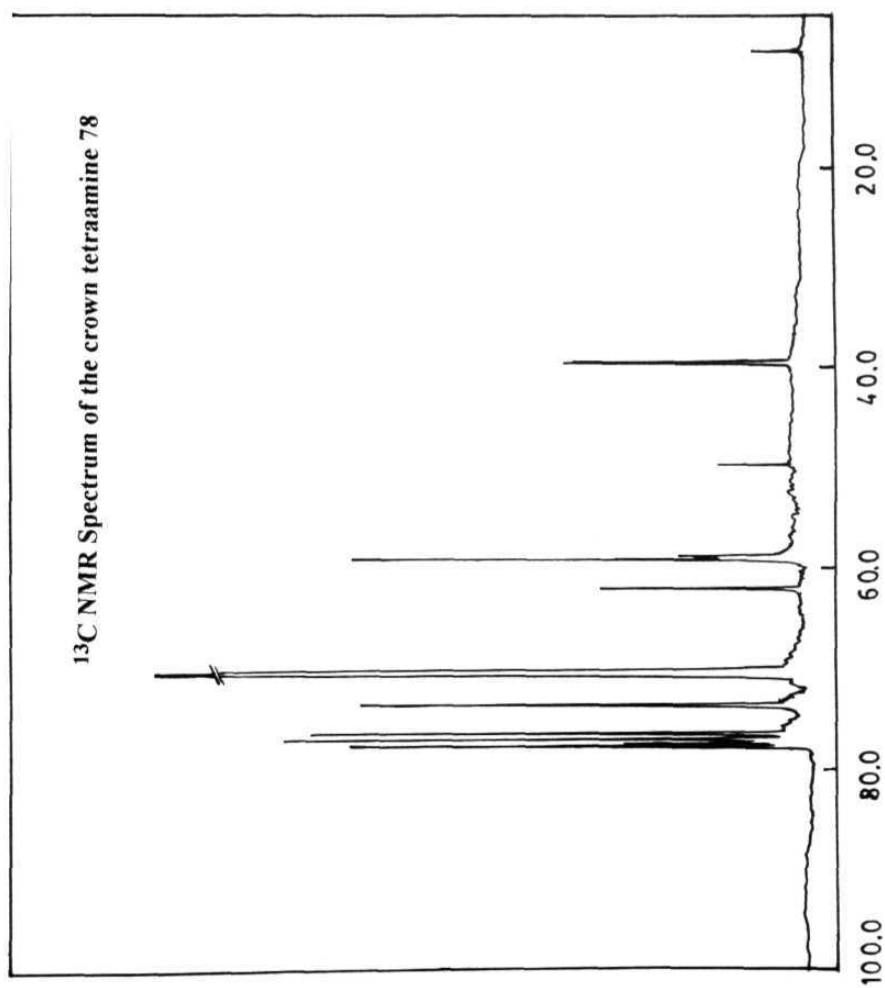
¹H NMR Spectrum of the crown N,N'-dibenzyltetraamine 77

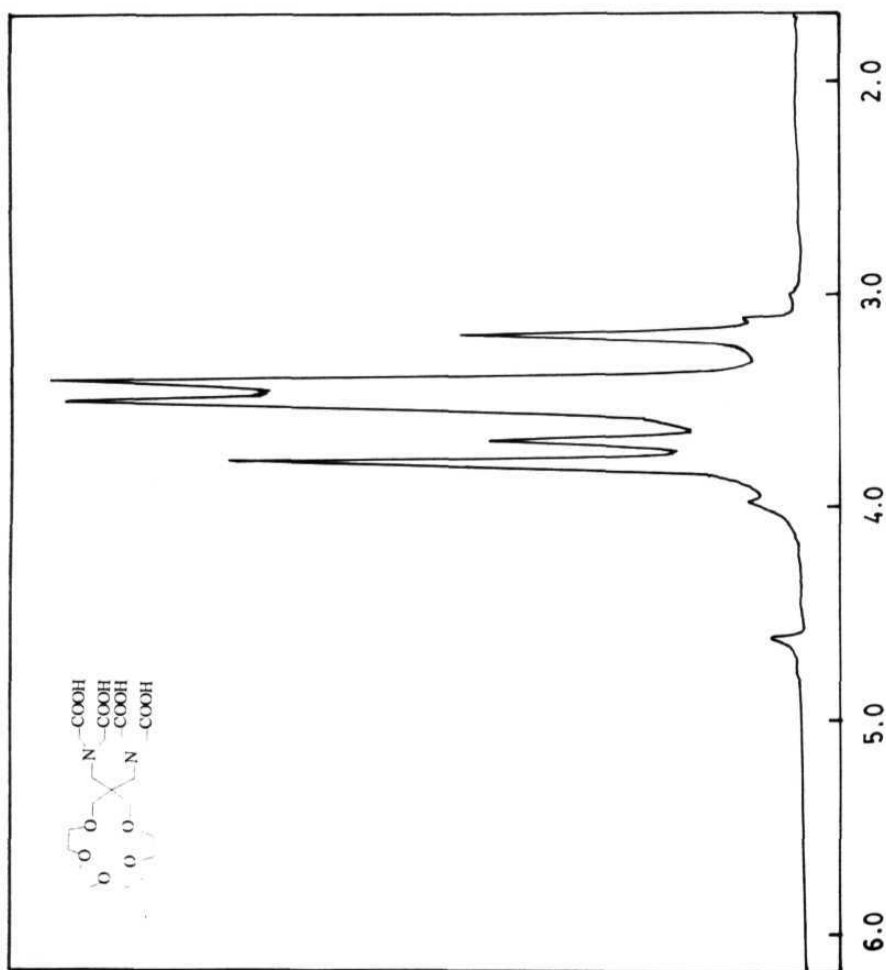


^{13}C NMR Spectrum of the crown N,N' -dibenzyltetraamine 77

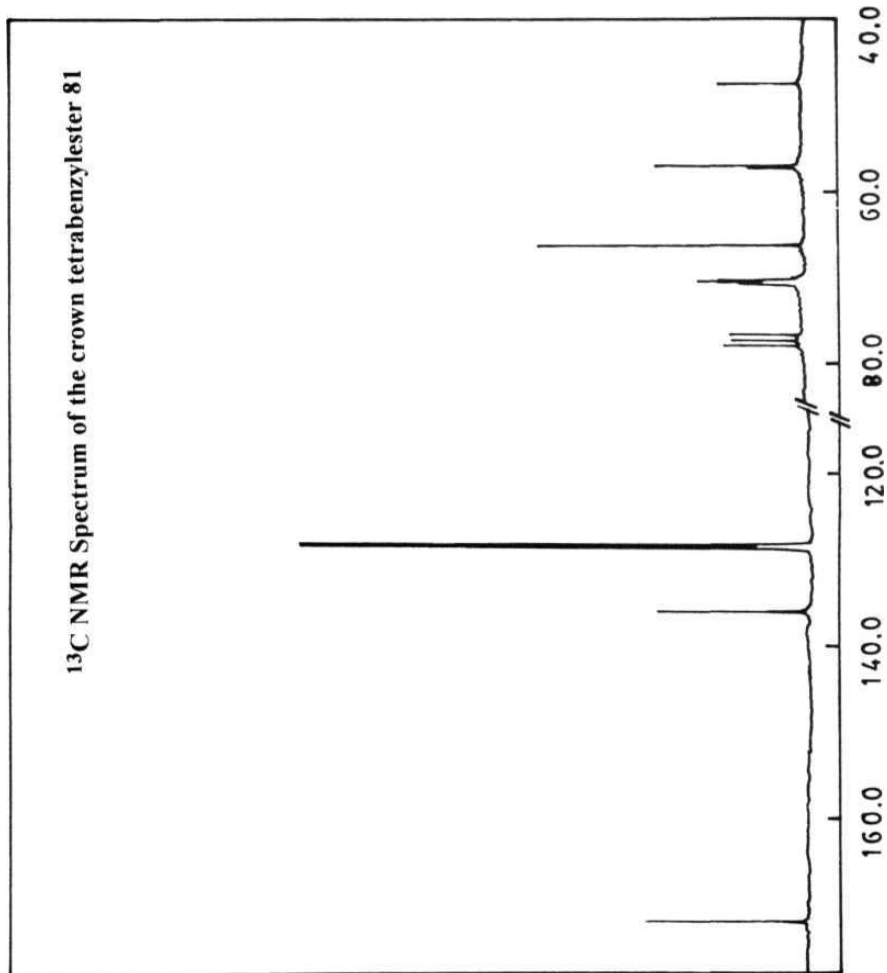


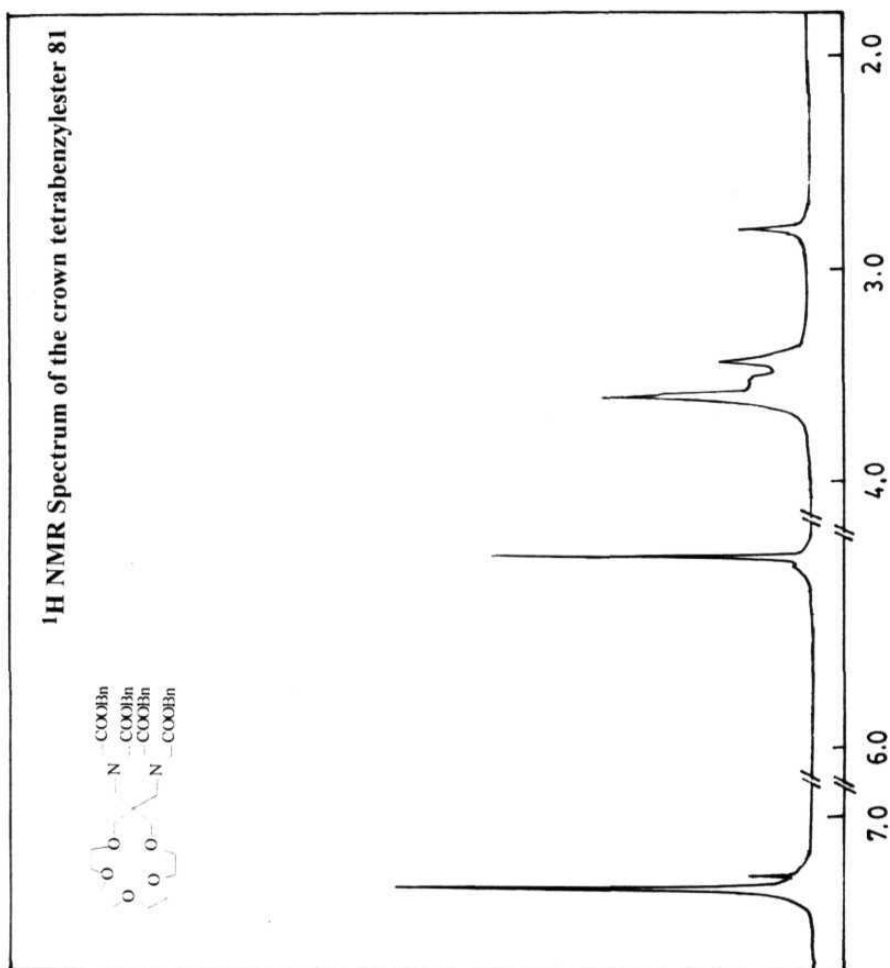
^1H NMR Spectrum of the crown tetraamine 78



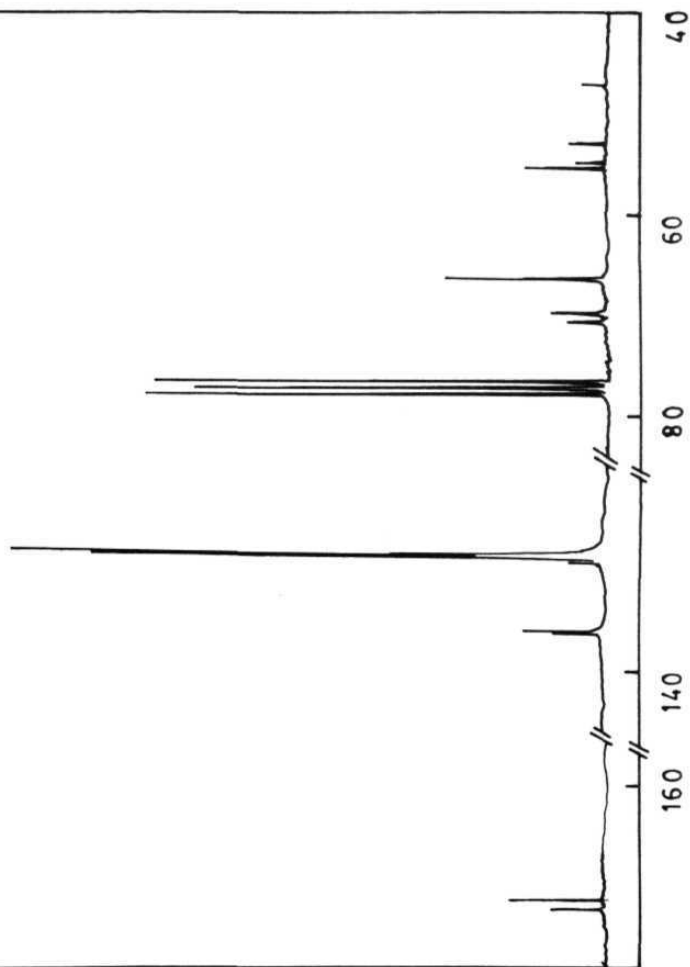


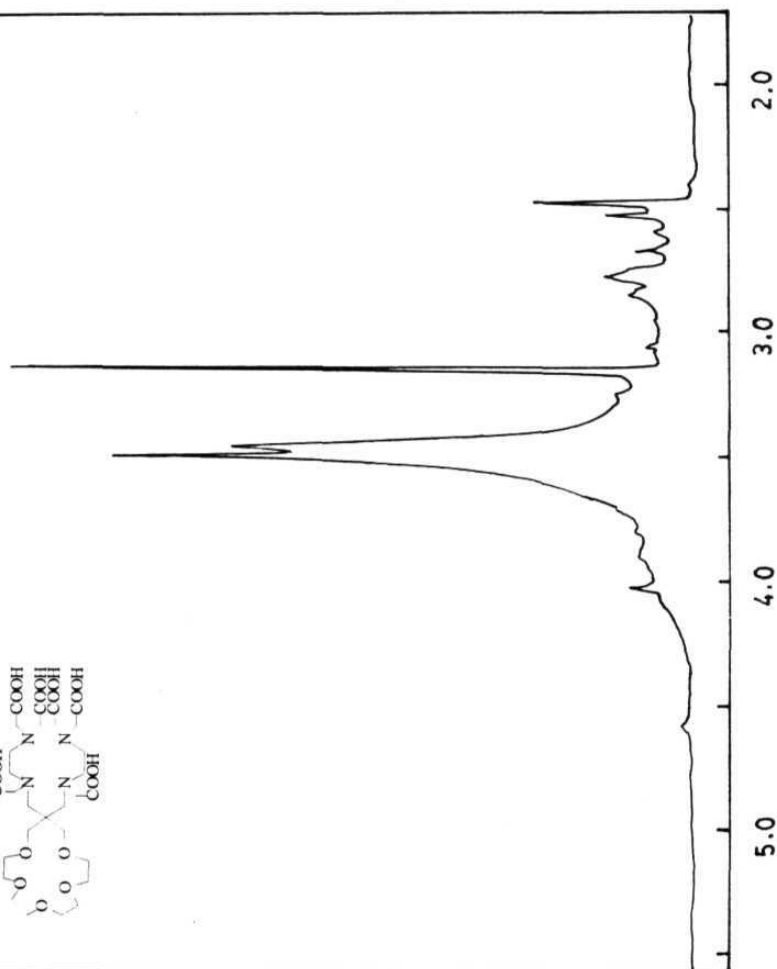
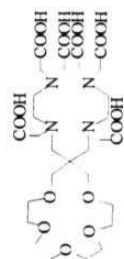
^1H NMR Spectrum of the crown diaminetetraacetic acid 79



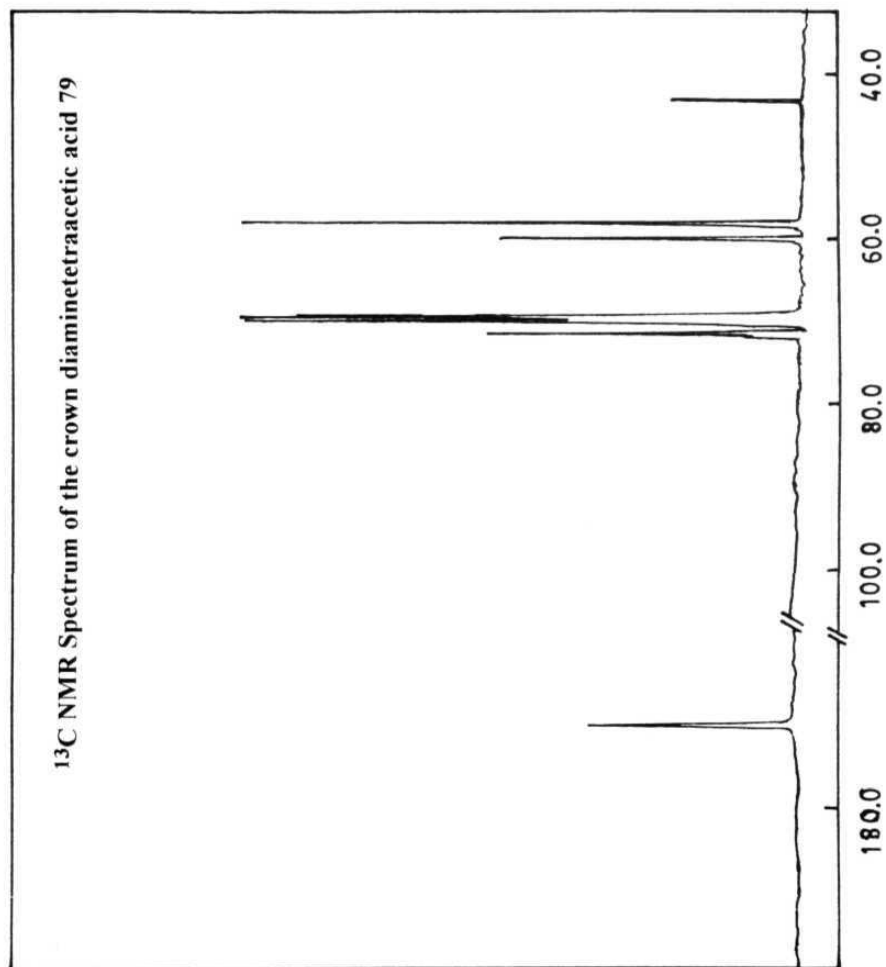


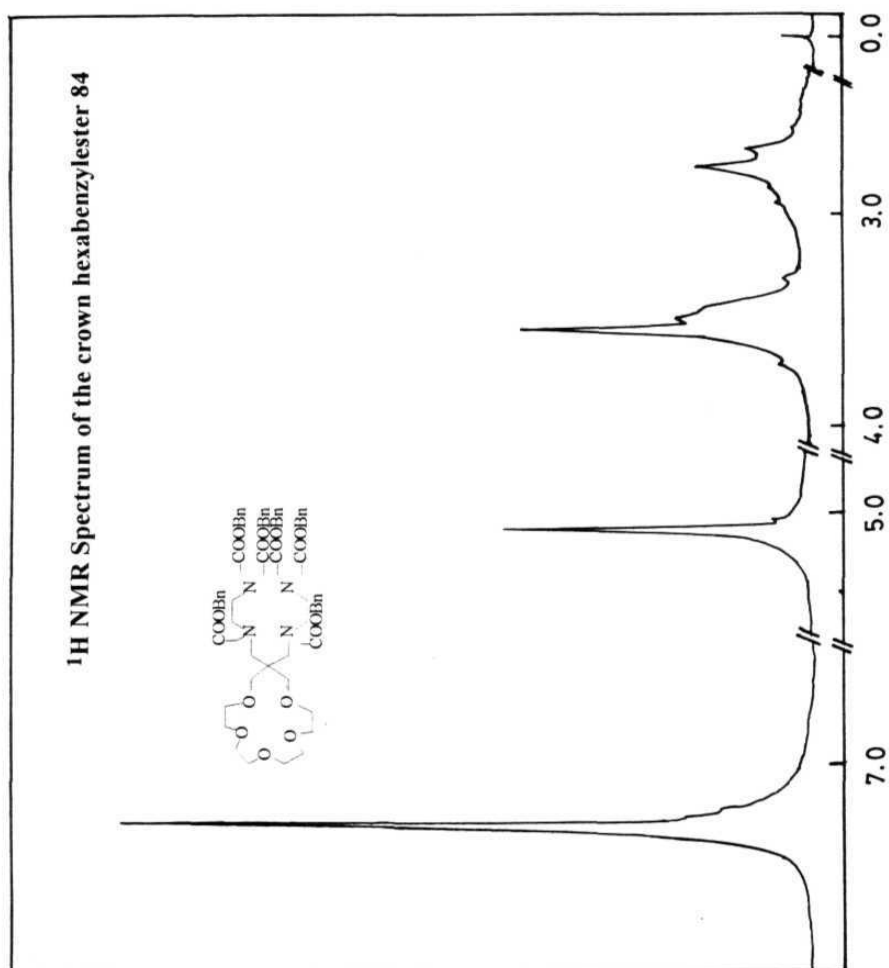
^{13}C NMR Spectrum of the crown hexabenzylester 84



¹H NMR Spectrum of the crown tetraaminehexaacetic acid 82

^{13}C NMR Spectrum of the crown diaminetetraacetic acid 79





REFERENCES

REFERENCES

1. Pedersen, C J. *J. Am. Ghent. Soc.* **1967**, *89*, 2495, 7017.
2. Lehn, J M *Angew. Chem., Int. Ed. Engl.* **1988**, *27*, **89**.
3. Lehn, J M. Ed. *Comprehensive Supramolecular Chemistry*, vols 1-10, Pregamon press, Oxford, **1996**.
4. Izatt, R M ; Pawlak, K.; Bradshaw, J S *Chem. Rev.* **1991**, *97*, 1721
5. a) Hiraoka, M. Ed *Crown Ethers and Analogous Compounds, Studies in Organic Chemistry 45*, Elsevier: Amsterdam, **1992**; Chapter 4 b) Cobben, P. L. H. M , Egberink, R J M , Bommer, J G.; Bergveld, P.; Verboom, W , Reinhoudt, D. N *J. Am. Chem. Soc.* **1992**, *114*, 10573.
6. Dozol, J F. Use of Liquid Membranes for Treatment of Nuclear Wastes In *Future Industrial Prospects of Membrane Processes*; Cecille, L, Toussaint, J.-C, Eds.; Elsevier: London, **1989**.
7. Van Straaten-Nijenhuis, W. F , De Jong, F.; Reinhoudt, D. N *Recl. Trav. Chim. Pays-Bas* **1993**, *772*, 317
8. a) Parker D. *Chem. Soc. Rev.* **1990**, *79*, 271. b) Cooper, S R. Ed *Crown Compounds Toward Future Applications*, VCH Publishers, Inc New York, **1992**; Chapter 4.
9. Patai, S.; Rappoport, Z. Eds *Crown Ethers and Analogs*, John Wiley and Sons, Chichester, **1989**, Chapter 2
10. March, J *Advanced Organic Chemistry: Reactions, Mechanisms and Structure*, John Wiley and Sons, New York, **1992**, Chapter 10, p 363.
11. Weber, E J. *Org. Chem.* **1982**, *47*, 3478

12. van Veggel, F. C. J. M., Bos, M.; Harkema, S.; van de Bovenkamp, H.; Verboom, W.; Reedijk, J.; Reinhoudt, D. N. *J. Org. Chem.* **1991**, 56, 225.
13. a) Ghosh, P.; Tyeklar, Z.; Karlin, K. D.; Jacobson, R. R.; Zubieta, J. J. *Am. Chem. Soc.* **1987**, 109, 6889 b) Casella, L.; Gullotti, M.; Pallanza, G.; Rigoni, L. *J. Am. Chem. Soc.* **1988**, 110, 4221. c) Tyeklar, Z., Karlin, K. D. *Acc. Chem. Res.* **1989**, 22, 241.
14. Costero, A. M.; Rodriguez, S. *Tetrahedron Lett.* **1992**, 33, 623
75. Nakatsuji, Y.; Kobayashi, H.; Okahara, M. *J. Org. Chem.* **1986**, 51, 3789.
16. Bloch, F.; Hansen, W. W.; Packard, M. *Phys. Rev.* **1948**, 70, 474
17. Young, I. R.; Clarke, G. J.; Gales, D. R. *Comput. Tomogr.* **1981**, 5, 534.
18. Carr, D. H.; Brown, J.; Bydder, G. M. *Lancet* **1984**, 1, 484.
19. Koenig, S. H.; Baglin, C.; Brown, R. D. III; Brewer, C. F. *Magn. Reson. Med.* **1984**, 1, 496
20. Lauffer, R. B. *Chem. Rev.* **1987**, 87, 901
21. May, P. M.; Bulman, R. A. *Prog. Med. Chem.* **1983**, 20, 225.
22. Bulman, R. A. "The Chemistry of Chelating Agents in Medical Sciences" in *Structure and Bonding*, Springer-Verlag: Berlin, **1987**, 67, p 93.
23. Cacheris, W. P.; Nickle, S. K.; Sherry, A. D. *Inorg. Chem.* **1987**, 26, 958.
24. Cotton, F. A.; Wilkinson, G. *Advanced Inorganic Chemistry*, 5 ed., Wiley-Interscience: New York, **1988**.

25. a) McMurry, T. J., Brechbiel, M , Kumar, **K** *Bioconjugate Chem.* **1992**, 3, 108. b) Studer M; Meares, **C F** *Ibid.* **1992**, 3, 337 and references cited therein.
26. a) Stetter, H ; Frank, **W** *Angew. Chem., Int. Ed. Engl.* **1976**, 15, 686 b) Stetter, H , Frank, W, Mertens, **R** *Tetrahedron* **1981**, 37, 767 c) Deigado, R , Frausto de Silva, **J R** *Talanta* **1982**, 29, 815.
27. a) Desreux, **J F** *Inorg. Chem.* **1980**, 19, 1319. b) Wang, X , Jin, T , Comblin, V , Lopez-Mut, A.; Merciny, E , Desreux, **J. F** *Inorg. Chem.* **1992**, 31, 1095.
28. a) Loncin, **M F** , Desreux, **J F** , Merciny, E *Inorg. Chem.* **1986**, 25, 2646 b) Brucher, E.; Laurenczy, G ; Makra, **Z** *Inorg. Chim. Acta* **1987**, 139, 141. c) Sherry, A D, Brown, **R D III**; Geraldles, **C F G C** , Koenig, S. H., Kuan, K ; Spiller, **M** *Inorg. Chem.* **1989**, 28, 620.
29. a) Kodama, M; Koike, T., Mahatma, A B , Kimura, E *Inorg. Chem.* **1991**, 30, 1270. b) Alexander, V. *Chem. Rev.* **1995**, 95, 273.
30. a) Taliaferro, **C H** ; Motekaitis, **R J** , Martell, A E. *Inorg. Chem.* **1984**, 23, 1188 b) Motekaitis, **R J**.; Sun, Y.; Martell, A. E. *Inorg. Chim. Acta* **1989**, 159, 29
31. Rocklage, **S M**.; Cacheris, **W P**.; Quay, **S C** ; Hahn, **F E** , Raymond, K. N. *Inorg. Chem.* **1989**, 28, 477.
32. Sun, Y ; Martell, A E., Reibenspies, J. H.; Welch, **M J** *Tetrahedron* **1991**, 47, 357
33. Dischino, **D D** , Delaney, E. J ; Emswiler, **J E**.; Gaughan, G. T., Prasad, **J S**.; Srivastava, S. K ; Tweedle, **M F** *Inorg. Chem.* **1991**, 30, 1265

34. Konings, M. S.; Dow, W. C ; Love, D B , Raymond, K. N.; Quay, S C.; Rocklage, S. M *Inorg. Chem.* **1990**, *29*, 1488.
35. Carvalho, J. F ; Kim, S -H ; Chang, C A *Inorg. Chem.* **1992**, *31*, 4065
36. Cram, D. J. *Science* **1988**, *240*, 760
37. a) Adam, K R ; Dancey, K P ; Leong, A J , Lindoy, L F ; McCool, B J.; McPartlin, M., Tasker, P A. *J. Am. Chem. Soc.* **1988**, *110*, 8471 b) Chang, C. A ; Ochaya, V. *Inorg. Chem.* **1986**, *25*, 355.
38. Cacheris, W P , Quay, S C.; Rocklage, S M *Magn. Reson. Imaging* **1990**, *8*, 467
39. Aime, S , Botta, M, Crich, S G , Giovenzana, G B , Jommi, G.; Pagliarin, R., Sisti, M. *J. Chem. Soc. (Chem. Commun.* **1995**, 1885
40. a) Deshpande, S V ; DeNardo, S J.; Meares, C F.; McCall, M J.; Adams, G. P., Moi, M. K., DeNardo, G L *J. Nucl. Med.* **1988**, *29*, 217
b) Moi, M K.; Meares, C F , McCall, M J , Cole, W C ; DeNardo, S *J. Anal. Biochem.* **1985**, *148*, 249 c) Moi, M K, Yanuck, M , Deshpande, S. V ; Hope, H.; DeNardo, S J ; Meares, C F *Inorg. Chem.* **1987**, *26*, 3458.
41. Moi, M. K.; Meares, C F ; DeNardo, S J *J. Am. Chem. Soc.* **1988**, *110*, 6266.
42. a) Sundberg, M. W.; Meares, C F ; Goodwin, D. A.; Diamanti, C I *J. Med. Chem.* **1974**, *17*, 1304 b) Sundberg, M W, Meares, C F, Goodwin, D A., Diamanti, C. I *Nature* **1984**, *250*, 587
43. Brechbiel, M. W , Gansow, O A., Atcher, R. W ; Schlom, J.; Esteban, J., Simpson, D. E.; Colcher, D. *Inorg. Chem.* **1986**, *25*, 2772.

44. Westerberg, D. A ; Carney, P L ; Rogers, P. E.; Kline, S. J , Johnson, D. **K J. Med. Chem.** **1989**, 32, 236.
45. a) Sundaralingam, M , Bergstorm, R , Strasburg, G , Rao, S T.; Roychowdhury, T.; Greaser, M , Wang, B C *Science* **1985**, 227, 945
b) Herzberg, O , James, M. N G *Nature* **1985**, 313, 653
46. Babu, Y. S.; Sack, J S ; Greenhough, T J., Bugg, C E.; Means, A R , Cook, W J *Nature* **1985**, 315, 37
47. a) Schauer, C K.; Anderson, O P. *J. Am. Chem. Soc.* 1987, 109, 3646.
b) Schauer, C K.; Anderson, O P. *Inorg. Chem.* **1988**, 27, 3118.
48. Tsien, R Y *Biochemistry* **1980**, 19, 2396
49. Kimura, T.; Maruyama, T , Okamura, M , Sugiyama, T, Ando, T.; Ohno, A *Bull. Chem. Soc. Jpn.* **1994**, 67, 1615.
50. De Clercq, E.; Yamamoto, N , Pauwels, R , Baba, M ; Schols, D , Nakashima, H.; Balzarini, J., Debyser, Z.; Murrer, B. A., Schwartz, D., Thornton, D.; Bridger, G.; Fricker, S., Henson, G., Abrams, M., Picker, D. *Proc. Natl. Acad. Sci.* **1992**, 89, 5286.
51. Ouchi, M ; Inoue, Y.; Liu, Y ; Nagamune, S , Nakamura, S , Wada, K , Hakushi, T *Bull. Chem. Soc. Jpn.* **1990**, 63, 1260
52. Johnstone, R A. W ; Wilby, A. H , Entwistle, J D. *Chem. Rev.* **1985**, 85, 129.
53. Horner, V L.; Gross, A *Liebigs Ann. Chem.* **1955**, 591, 117.
54. ElAmin, B , Anantharamaiah, G.; Royer, G , Means, G. *J. Org. Chem.* 1979, 44, 3442.
55. du Vigneaud, V., Beherens, O K *J. Biol. Chem.* 1937, 117, 27.
56. Ram, S ; Spicer, L D. *Synth. Commun.* **1987**, 17, 415

57. Huegi, B S.; Ebnother, A. M.; Rissi, E.; Gradient, F., Hauser, D.; Roemer, D ; Hill, R C , Buescher, H H ; Petcher, T. J. J. *Med. Chem.* **1983**, 26, 42.
58. Bouzard, D., Di Cesare, P., Essiz, M; Jacquet, J. P.; Kiechel, J R.; Remuzon, P.; Weber, A.; Oki, T.; Masuyoshi, M; Kessler, R. E , Fung-Tomc, J ; Desiderio, J *J. Med. Chem.* **1990**, 33, 1344.
59. Pearlman, W M *Tetrahedron Lett.*, **1967**, 17, 1663.
60. Takenouchi, K., Watanabe, K , Kota, Y , Koike, T.; Kimura, E *J. Org. Chem.* **1993**, 58, 1955.
61. Tait, A M ; Busch, D. H. *Inorganic Synthesis*. vol 18, p 22
62. Whitmore, B. C.; Eisenberg, R *Inorg. Chem.* **1983**, 22, 1
63. Hutchison, C A.; Judd, B. R., Pope, D F D *Proc. phys. Soc.* **1957**, B70, 514.
64. Bunzli, J. -C. G in *Handbook on the Physics and Chemistry of Rare Earths*, Gschneider, K A. Jr.; Eyring, L, Eds Elsevier Science Publishers B. V.: Amsterdam, **1987**, vol.9, Chapter 60.
65. Perrin, D D., Sayce, I. G. in *"Computational Methods for the Determination of Formation Constants"* Leggett, D.J. Ed. Plenum Press New York, **1985**, p.71.
66. Jonassen, H. B., LeBlanc, R. B., Meibohm, A W.; Rogan, R M *J. Am. Chem. Soc.* **1950**, 72, 2430.
67. Chaberek, S.; JR.; Martell, A. E. *J. Am. Chem. Soc.* **1952**, 74, 6228
68. Newman, M. S , Busch, D. H , Cheney, G. E.; Gustafson, C R *Inorg. Chem.*, **1972**, //, 2890.

69. a) Schwarzenbach, G.; Ackermann, H *Helv. Chim. Acta* **1947**, *30*, 1798
 b) Calvin, M., Martell, A. E. "The Chemistry of the Metal Complexes", Prentice-Hall, Inc., New York **1952**, Chapter 4
70. a) Schwarzenbach, G.; Ackermann, H *Helv. Chim. Acta* **1948**, *31*, 1029
 b) Harris, W. R.; Martell, A. E. *Inorg. Chem.* **1976**, *15*, 713
71. Clarke, H. T. *J. Chem. Soc.* **1910**, 97, 416
72. Aime, S., Botta, M., Ermondi, G., Fredeli, F., Uggeri, F. *Inorg. Chem.* **1992**, *31*, 1100
73. Vinarov, I. V.; Kornelli, M. E. *Khim., Prom., Nauk. -Tekhn. Zh* **1962**, No 1, 28-30. (C. A. **1963**, 59, 1257f).
74. a) Hendricson, R. R., Mack, M. P.; Palmer, R. A., Ottolenghi, A., Ghirardelli, R. G. *Toxicol. Appl. Pharmacol.* **1978**, *44*, 263
 b) Hasegawa, S., Sasagawa, S., Nambu, N.; Nagai, T. *Chem. Pharm. Bull.* **1977**, *25*, 3125
 c) Tso, W. W., Fung, W. P. *Inorg. Chim. Acta* **1981**, *55*, 129.